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UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Docket No.
1038-1000 MIS
Total Pages in this Submission
3

jc535 U.S. PTO 01/07/00

TO THE ASSISTANT COMMISSIONER FOR PATENTS
Box Patent Application
Washington, D.C. 20231

Transmitted herewith for filing under 35 U.S.C. 111(a) and 37 C.F.R. 1.53(b) is a new utility patent application for an invention entitled:

CHIMERIC IMMUNOGENS

jc564 U.S. PTO 09/479240 01/07/00

and invented by:

Michel H. Klein et al

If a CONTINUATION APPLICATION, *check appropriate box and supply the requisite information:*

☒ **Continuation** ☐ **Divisional** ☐ **Continuation-in-part (CIP)** of prior application No.: **08/467,961**

Which is a:

☐ **Continuation** ☒ **Divisional** ☐ **Continuation-in-part (CIP)** of prior application No.: **08/001,554**

Which is a:

☐ **Continuation** ☐ **Divisional** ☐ **Continuation-in-part (CIP)** of prior application No.: _____

Enclosed are:

Application Elements

1. ☒ Filing fee as calculated and transmitted as described below
2. ☒ Specification having **42** pages and including the following:
   - a. ☒ Descriptive Title of the Invention
   - b. ☐ Cross References to Related Applications *(if applicable)*
   - c. ☐ Statement Regarding Federally-sponsored Research/Development *(if applicable)*
   - d. ☐ Reference to Microfiche Appendix *(if applicable)*
   - e. ☒ Background of the Invention
   - f. ☒ Brief Summary of the Invention
   - g. ☒ Brief Description of the Drawings *(if drawings filed)*
   - h. ☒ Detailed Description
   - i. ☒ Claim(s) as Classified Below
   - j. ☒ Abstract of the Disclosure

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Application Elements (Continued)

3. ☒ Drawing(s) *(when necessary as prescribed by 35 USC 113)*
   a. ☒ Formal Number of Sheets 39
   b. ☐ Informal Number of Sheets _____
4. ☒ Oath or Declaration
   a. ☐ Newly executed *(original or copy)* ☐ Unexecuted
   b. ☒ Copy from a prior application (37 CFR 1.63(d)) *(for continuation/divisional application only)*
   c. ☒ With Power of Attorney ☐ Without Power of Attorney
   d. ☐ *DELETION OF INVENTOR(S)*
   Signed statement attached deleting inventor(s) named in the prior application, see 37 C.F.R. 1.63(d)(2) and 1.33(b).
5. ☒ Incorporation By Reference *(usable if Box 4b is checked)*
   The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby incorporated by reference therein.
6. ☐ Computer Program in Microfiche *(Appendix)*
7. ☒ Nucleotide and/or Amino Acid Sequence Submission *(if applicable, all must be included)*
   a. ☒ Paper Copy
   b. ☒ Computer Readable Copy *(identical to computer copy)*
   c. ☒ Statement Verifying Identical Paper and Computer Readable Copy

Accompanying Application Parts

8. ☐ Assignment Papers *(cover sheet & document(s))*
9. ☐ 37 CFR 3.73(B) Statement *(when there is an assignee)*
10. ☐ English Translation Document *(if applicable)*
11. ☒ Information Disclosure Statement/PTO-1449 ☒ Copies of IDS Citations
12. ☒ Preliminary Amendment
13. ☒ Acknowledgment postcard
14. ☐ Certificate of Mailing
    ☐ First Class ☐ Express Mail *(Specify Label No.):* _____

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Accompanying Application Parts (Continued)

15. ☐ Certified Copy of Priority Document(s) *(if foreign priority is claimed)*

16. ☐ Additional Enclosures *(please identify below):*

Fee Calculation and Transmittal

CLAIMS AS FILED

For	#Filed	#Allowed	#Extra		Rate	Fee
Total Claims	18	- 20 =	0	x	$18.00	$0.00
Indep. Claims	3	- 3 =	0	x	$78.00	$0.00
Multiple Dependent Claims (check if applicable) ☐						$0.00
					BASIC FEE	$760.00
OTHER FEE ***(specify purpose)***						$0.00
					TOTAL FILING FEE	$760.00

☒ A check in the amount of **$760.00** to cover the filing fee is enclosed.

☐ The Commissioner is hereby authorized to charge and credit Deposit Account No. as described below. A duplicate copy of this sheet is enclosed.

- ☐ Charge the amount of as filing fee.
- ☐ Credit any overpayment.
- ☐ Charge any additional filing fees required under 37 C.F.R. 1.16 and 1.17.
- ☐ Charge the issue fee set in 37 C.F.R. 1.18 at the mailing of the Notice of Allowance, pursuant to 37 C.F.R. 1.311(b).

Signature

Michael I. Stewart - Reg. No. 24,973

Dated: **January 6, 2000**

cc:

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Our Ref: 1038-1000 MIS:as

In re patent application

No.

Applicant: Michel H. Klein et al

Title: CHIMERIC IMMUNOGENS

Filed:

Group No. 1645

Examiner: Mark Navarro

January 6, 2000

PRELIMINARY AMENDMENT

The Commissioner of Patents
and Trademarks,
Washington, D.C. 20231,
U.S.A.

Dear Sir:

Please amend this application in the following manner:

In the Disclosure:

On page 4, lines 13, 15, 17 and 19, change "antigenic" to "immunogenic" at each occurrence;

Page 4, line 20, delete the word "generally";

Page 5, lines 11 and 13, change "antigenic" to "immunogenic" at each occurrence;

Page 5, line 16, insert "The first and second pathogens are selected from bacterial and viral pathogens." After "system."

On page 6, line 23, change "Figures 1A and 1B" to read "Figures 1A to 1E";

On page 6, line 23, change "Figures 3A to 3B" to read "Figures 3A to 3E";

On page 6, line 33, change "Figures 5A to 5B" to read "Figures 5A to 5E";

On page 7, line 8, change "Figures 9A to 9B" to read "Figures 9A to 9D";

Page 14, line 18, insert "SE ID NO:21" after "AGGACAAAAG".

Page 15, line 13, change "1887" to "1886";

Page 19, line 31 and page 23, line 10, underline the terms "Spodoptera frugiperda" to signify that they should be printed in italicized form.

On pages 24, lines 20 and 26, capitalize the term "TRITON X-100" and add "(Trademark for a non-ionic detergent which is octadienyl phenol (ethylene glycol)$_{10}$" following the term in line 20;

Page 26, line 32, change "homogenates" to "lavages".

Add the Sequence Listing enclosed.

In the Claims:

Cancel claims 1 to 58.

Add new claims 59 to 76 as follows:

59. (New) A multimeric hybrid gene encoding a chimeric protein including a protein from parainfluenza virus (PIV) and a protein from respiratory syncytial virus (RSV), comprising a nucleotide sequence encoding a PIV-3 protein or a fragment thereof having fusion activity or a PIV-3 HN protein or a fragment thereof having hemagglutinin-neurominidase activity linked to a nucleotide sequence coding for a RSV G protein or a fragment thereof having attachment activity or a RSV F protein or a fragment thereof having fusion activity.

60. (New) The hybrid gene of claim 59 which is selected from the group consisting of F_{PIV-3}-F_{RSV}, F_{RSV}-HN_{PIV-3} and F_{PIV-3}-G_{RSV} hybrid genes.

61. (New) The hybrid gene of claim 59 contained in an expression vector.

62. (New) The hybrid gene of claim 61 in the form of a plasmid selected from the group consisting of pAC DR7 (ATCC 75387), pD2 RF-HN (ATCC 75388) and pD2 F-G (ATCC 75389).

63. (New) Eukaryotic cells containing the multimeric hybrid gene of claim 59 for expression of the chimeric protein encoded by the hybrid gene.

64. (New) The cells of claim 63 which are mammalian cells, insect cells, yeast cells or fungal cells.

65. (New) A vector for antigen delivery containing the gene of claim 59.

66. (New) The vector of claim 65 which is viral vector.

67. (New) The vector of claim 66 wherein said viral vector is selected from the group consisting of poxviral, adenoviral and retroviral viral vectors.

68. (New) The vector of claim 65 which is a bacterial vector.

69. (New) The vector of claim 68 wherein said bacterial vector is selected from salmonella and mycobacteria.

70. (New) A process for the preparation of a chimeric protein including a protein from parainfluenza virus (PIV) and a protein from respiratory syncytial virus (RSV), which comprises:

isolating a first nucleotide sequence encoding a PIV-3 protein or a fragment thereof having fusion activity or a PIV-3 HN protein or a fragment thereof having hemagglutinin-neurominidase activities,

isolating a second nucleotide sequence encoding a RSV-G protein or a fragment thereof having attachment activity or a RSV protein or a fragment thereof having fusion activity,

linking said first and second nucleotide sequences to form a multimeric hybrid gene, and

expressing the multimeric hybrid gene in a cellular expression system.

71. (New) The process of claim 70 wherein said multimeric hybrid gene is selected from the group consisting of F_{PIV-3}-F_{RSV}, F_{RSV}-HN_{PIV-3} and F_{PIV-3}-G_{RSV} hybrid genes.

72. (New) The process of claim 71 wherein said multimeric hybrid gene is contained in an expression vector comprising a gene selected from the group consisting of pAC DR7 (ATCC 75387), pD2 RF-HN (ATCC 75388) and pD2 F-G (ATCC 75389).

73. (New) The process of claim 71 wherein said cellular expression system is provided by mammalian cells, insect cells, yeast cells or fungal cells.

74. (New) The process of claim 70 including separating a chimeric protein from a culture of said eukaryotic cellular expression and purifying the separated chimeric protein.

75. (New) A chimeric protein including a protein from parainfluenza virus (PIV) and a protein from respiratory syncytial virus (RSV), comprising a PIV-3 F protein or a fragment thereof having fusion activity or a PIV-3 HN protein or a fragment thereof having hemagglutinin-neurominidase activity.

76. (New) The chimeric protein of claim 75 which is selected from the group consisting of F_{PIV-3}-F_{RSV}, F_{RSV}-HN_{PIV-3} and F_{PIV-3}-G_{RSV} hybrid genes.

In the Drawings:

Cancel the informal drawings submitted with the application and substitute therefor the formal drawings enclosed.

REMARKS

The disclosure has been amended to correspond to the changes made to the parent application.

Formal drawings have been substituted for the informal drawings. It is noted that Figure 5 has been amended as shown on the enclosed print of the original drawing in red, to correct a spelling error. The Sequence Listing enclosed herewith, both in hard copy and computer-readable form, includes the changes effected in the drawings.

It is hereby stated that the hard copy and computer-readable form of the Sequence Listing are the same.

The claims have been amended in the interests of expedited prosecution. In this regard, it is noted that the definition of the chimeric protein is the same as allowed in the diagnostic claims of the parent case. The claims directed to the hybrid gene and method of making the chimeric protein recombinantly utilizes corresponding language.

The PTO-1449 submitted herewith lists all prior art cited by or to the PTO in the parent and related filings. Copies of each of the references are enclosed for the Examiner's convenience.

Respectfully submitted,

M.I. Stewart
Reg. No. 24,973

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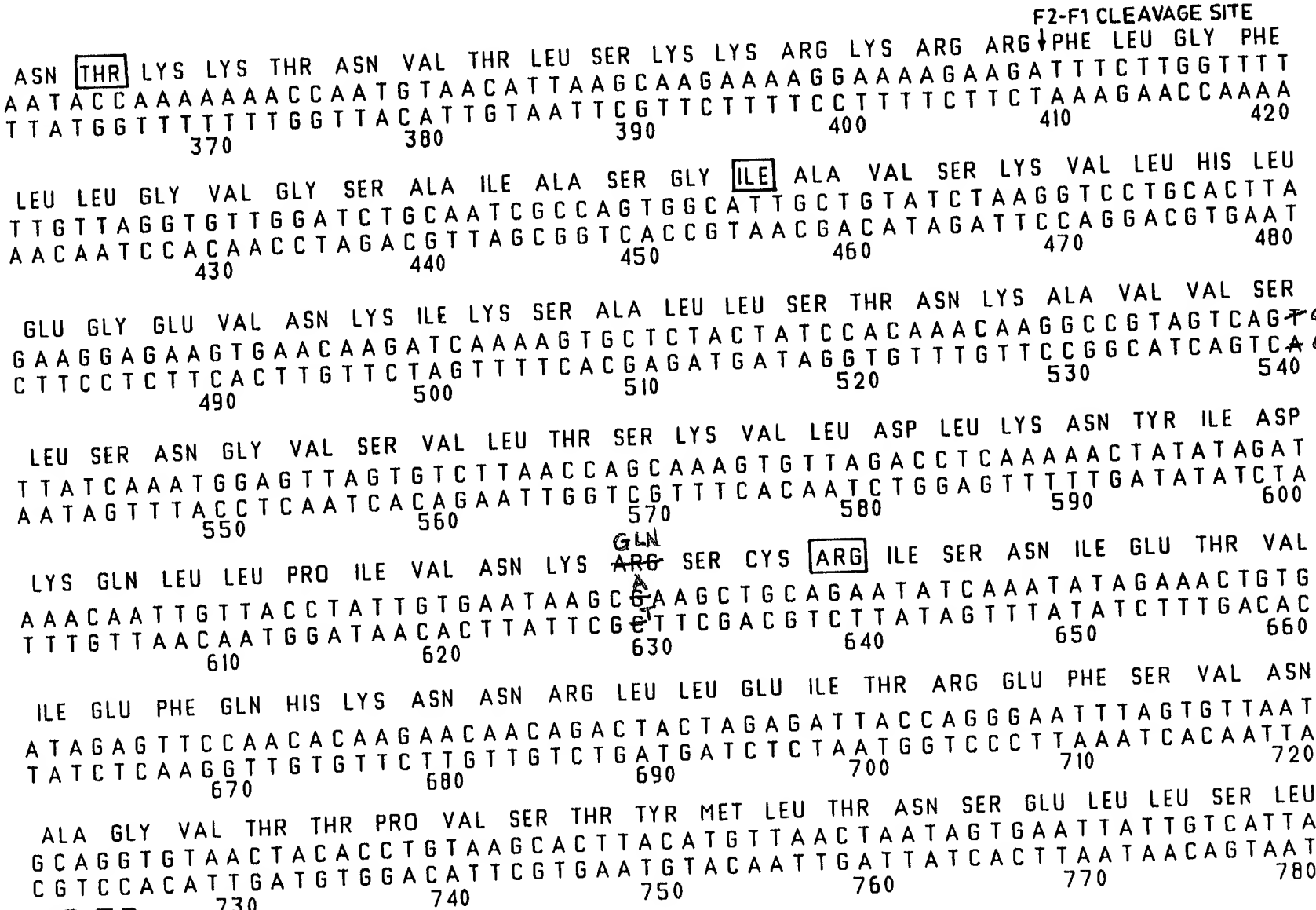

F2-F1 CLEAVAGE SITE
ASN THR LYS LYS THR ASN VAL THR LEU SER LYS LYS ARG LYS ARG ARG PHE LEU GLY PHE
AATACCAAAAAAACCAATGTAACATTAAGCAAGAAAAGGAAAAGAAGATTTCTTGGTTTT
TTATGGTTTTTTTGGTTACATTGTAATTCGTTCTTTTCCTTTTCTTCTAAAGAACCAAAA
370 380 390 400 410 420
LEU LEU GLY VAL GLY SER ALA ILE ALA SER GLY ILE ALA VAL SER LYS VAL LEU HIS LEU
TTGTTAGGTGTTGGATCTGCAATCGCCAGTGGCATTGCTGTATCTAAGGTCCTGCACTTA
AACAATCCACAACCTAGACGTTAGCGGTCACCGTAACGACATAGATTCCAGGACGTGAAT
430 440 450 460 470 480
GLU GLY GLU VAL ASN LYS ILE LYS SER ALA LEU LEU SER THR ASN LYS ALA VAL VAL SER
GAAGGAGAAGTGAACAAGATCAAAAGTGCTCTACTATCCACAAACAAGGCCGTAGTCAGTC
CTTCCTCTTCACTTGTTCTAGTTTTCACGAGATGATAGGTGTTTGTTCCGGCATCAGTCAG
490 500 510 520 530 540
LEU SER ASN GLY VAL SER VAL LEU THR SER LYS VAL LEU ASP LEU LYS ASN TYR ILE ASP
TTATCAAATGGAGTTAGTGTCTTAACCAGCAAAGTGTTAGACCTCAAAAACTATATAGAT
AATAGTTTACCTCAATCACAGAATTGGTCGTTTCACAATCTGGAGTTTTTGATATATCTA
550 560 570 580 590 600
LYS GLN LEU LEU PRO ILE VAL ASN LYS GLN ARG SER CYS ARG ILE SER ASN ILE GLU THR VAL
AAACAATTGTTACCTATTGTGAATAAGCAAGCTGCAGAATATCAAATATAGAAACTGTG
TTTGTTAACAATGGATAACACTTATTCGTTCGACGTCTTATAGTTTATATCTTTGACAC
610 620 630 640 650 660
ILE GLU PHE GLN HIS LYS ASN ASN ARG LEU LEU GLU ILE THR ARG GLU PHE SER VAL ASN
ATAGAGTTCCAACACAAGAACAACAGACTACTAGAGATTACCAGGGAATTTAGTGTTAAT
TATCTCAAGGTTGTGTTCTTGTTGTCTGATGATCTCTAATGGTCCCTTAAATCACAATTA
670 680 690 700 710 720
ALA GLY VAL THR THR PRO VAL SER THR TYR MET LEU THR ASN SER GLU LEU LEU SER LEU
GCAGGTGTAACTACACCTGTAAGCACTTACATGTTAACTAATAGTGAATTATTGTCATTA
CGTCCACATTGATGTGGACATTCGTGAATGTACAATTGATTATCACTTAATAACAGTAAT
730 740 750 760 770 780

FIG.5B.

TITLE OF INVENTION

CHIMERIC IMMUNOGENS

FIELD OF INVENTION

The present invention relates to the engineering and expression of multimeric hybrid genes containing sequences from the gene coding for immunogenic proteins or protein fragments of numerous pathogens.

BACKGROUND TO THE INVENTION

The advantage of the approach taken by the present invention is to produce single immunogens containing protective antigens from a range of pathogens. Such chimeras greatly simplify the development of combination vaccines, in particular, with the view ultimately to produce single dose multivalent vaccines. Multivalent vaccines are currently made by separately producing pathogens and/or their pertinent antigens and combining them in various formulations. This is a labour intensive, costly and complex manufacturing procedure. In contrast, the availability of a single immunogen capable of protecting against a range of diseases would solve many of the problems of multivalent vaccine production. Several chimeric immunogens of the type provided herein may be combined to decrease the number of individual antigens required in a multivalent vaccine.

Human Parainfluenza virus types 1,2,3 and Respiratory syncytial virus types A and B are the major viral pathogens responsible for causing severe respiratory tract infections in infants and young children. It is estimated that, in the United States alone, approximately 1.6 million infants under one year of age will have a clinically significant RSV infection each year and an additional 1.4 million infants will be infected with PIV-3. Approximately 4000 infants less than one year of age in the United States die each year from complications arising from severe respiratory tract disease caused by infection with RSV and PIV-3. The WHO

and NIALD vaccine advisory committees ranked RSV number two behind HIV for vaccine development while the preparation of an efficacious PIV-3 vaccine is ranked in the top ten vaccines considered a priority for vaccine development.

Safe and effective vaccines for protecting infants against these viral infections are not available and are urgently required. Clinical trials have shown that formaldehyde-inactivated and live-attenuated viral vaccines failed to adequately protect vaccinees against these infections. In fact, infants who received the formalin-inactivated RSV vaccine developed more serious lower respiratory tract disease during subsequent natural RSV infection than did the control group. [Am. J. Epidemiology 89, 1969, p.405-421; J. Inf. Dis. 145, 1982, p.311-319]. Furthermore, RSV glycoproteins purified by immunoaffinity chromatography using elution at acid pH induced immunopotentiation in cotton rats. [Vaccine, 10(7), 1992, p.475-484]. The development of efficacious PIV-3 and RSV vaccines which do not cause exacerbated pulmonary disease in vaccinees following injection with wild-type virus would have significant therapeutic implications. It is anticipated that the development of a single recombinant immunogen capable of simultaneously protecting infants against diseases caused by infection with both Parainfluenza and Respiratory syncytial viruses could significantly reduce the morbidity and mortality caused by these viral infections.

It has been reported that a protective response against PIV-3 and RSV is contingent on the induction of neutralizing antibodies against the major viral surface glycoproteins. For PIV, these protective immunogens are the HN protein which has a molecular weight of 72 kDa and possesses both hemagglutination and neuraminidase activities and the fusion (F) protein, which has a molecular weight of 65 kDa and which is responsible for

both fusion of the virus to the host cell membrane and cell-to-cell spread of the virus. For RSV, the two major immunogenic proteins are the 80 to 90 kDa G glycoprotein and the 70 kDa fusion (F) protein. The G and F proteins are thought to be functionally analogous to the PIV HN and F proteins, respectively. The PIV and RSV F glycoproteins are synthesized as inactive precursors (FO) which are proteolytically cleaved into N-terminal F2 and C-terminal F1 fragments which remain linked by disulphide bonds.

Recombinant surface glycoproteins from PIV and RSV have been individually expressed in insect cells using the baculovirus system [Ray et al., (1989), Virus Research, 12: 169-180; Coelingh et al., (1987), Virology, 160: 465-472; Wathen et al., (1989), J. of Inf. Dis. 159: 253-263] as well as in mammalian cells infected with recombinant poxviruses [Spriggs, et al., (1987), J. Virol. 61: 3416-3423; Stott et al., (1987), J. Virol. 61: 3855-3861]. Recombinant antigens produced in these systems were found to protect immunized cotton rats against live virus challenge. More recently, hybrid RSV F-G [Wathan et al., (1989), J. Gen Virol. 70: 2625-2635; Wathen, published International Patent application WO 89/05823] and PIV-3 F-HN [Wathen, published International Patent Application WO 89/10405], recombinant antigens have been engineered and produced in mammalian and insect cells. The RSV F-G hybrid antigen was shown to be protective in cotton rats [Wathan et al., (1989), J. Gen. Virol. 70: 2637-2644] although it elicited a poor anti-G antibody response [Connors et al., (1992), Vaccine 10: 475-484]. The protective ability of the PIV-3 F-HN protein was not reported in the published patent application. These antigens were engineered with the aim to protect against only the homologous virus, that is either RSV or PIV-3. However, it would be advantageous and economical to engineer and produce a single

recombinant immunogen containing at least one protective antigen from each virus in order simultaneously to protect infants and young children against both PIV and RSV infections. The chimeric proteins provided herein for such purpose also may be administered to pregnant women or women of child bearing age to stimulate maternal antibodies to both PIV and RSV. In addition, the vaccine also may be administered to other susceptible individuals, such as the elderly.

SUMMARY OF INVENTION

In its broadest aspect, the present invention provides a multimeric hybrid gene, comprising a gene sequence coding for an antigenic region of a protein from a first pathogen linked to a gene sequence coding for an antigenic region of a protein from a second pathogen and to a chimeric protein encoded by such multimeric hybrid gene. Such chimeric protein comprises an antigenic region of a protein from a first pathogen linked to an antigenic region of a protein from a second pathogen.

The first and second pathogens generally are selected from bacterial and viral pathogens and, in one embodiment, may both be viral pathogens. Preferably, the first and second pathogens are selected from those causing different respiratory tract diseases, which may be upper and lower respiratory tract diseases. In a preferred embodiment, the first pathogen is parainfluenza virus and the second pathogen is respiratory syncytial virus. The PIV protein particularly is selected from PIV-3 F and HN proteins and the RSV protein particularly is selected from RSV G and F proteins. Another aspect of the invention provides cells containing the multimeric hybrid gene for expression of a chimeric protein encoded by the gene. Such cells may be bacterial cells, mammalian cells, insect cells, yeast cells or fungal cells. Further, the present invention provides a live vector for antigen delivery containing the multimeric

hybrid gene, which may be a viral vector or a bacterial vector, and a physiologically-acceptable carrier therefor. Such live vector may form the active component of a vaccine against diseases caused by multiple pathogenic infections. Such vaccine may be formulated to be administered in an injectable form, intranasally or orally.

In an additional aspect of the present invention, there is provided a process for the preparation of a chimeric protein, which comprises isolating a gene sequence coding for an antigenic region of a protein from a first pathogen; isolating a gene sequence coding for an antigenic region of a protein from a second pathogen; linking the gene sequences to form a multimeric hybrid gene; and expressing the multimeric hybrid gene in a cellular expression system. Such cellular expression system may be provided by bacterial cells, mammalian cells, insect cells, yeast cells or fungal cells. The chimeric protein product of gene expression may be separated from a culture of the cellular expression system and purified.

The present invention further includes a vaccine against diseases caused by multiple pathogen infections, comprising the chimeric protein encoded by the multimeric hybrid gene and a physiologically-acceptable carrier therefor. Such vaccine may be formulated to be administered in an injectable form, intranasally or orally.

The vaccines provided herein may be used to immunize a host against disease caused by multiple pathogenic infections, particularly those caused by a parainfluenza virus and respiratory syncytial virus, by administering an effective amount of the vaccine to the host. As noted above, for human PIV and RSV , the host may be infants and young children, pregnant women as well as those of a

child-bearing age, and other susceptible persons, such as the elderly.

The chimeric protein provided herein also may be used as a diagnostic reagent for detecting infection by a plurality of different pathogens in a host, using a suitable assaying procedure.

It will be appreciated that, while the description of the present invention which follows focuses mainly on a chimeric molecule which is effective for immunization against diseases caused by infection by PIV and RSV, nevertheless the invention provided herein broadly extends to any chimeric protein which is effected for immunization against diseases caused by a plurality of pathogens, comprising an antigen from each of the pathogens linked in a single molecule, as well as to genes coding for such chimeric molecules.

In this application, by the term "multimeric hybrid genes" we mean genes encoding antigenic regions of proteins from different pathogens and by the term "chimeric proteins" we mean immunogens containing antigenic regions from proteins from different pathogens.

BRIEF DESCRIPTION OF DRAWINGS

Figure 1 shows the nucleotide (SEQ ID No: 1) and amino acid (SEQ ID No: 2) sequence of a PCR-amplified PIV-3 F gene and F protein, respectively;

Figure 2 shows the restriction map of the PIV-3 F gene;

Figure 3 shows the nucleotide (SEQ ID No: 3) and amino acid (SEQ ID No: 4) sequences of the PIV-3 HN gene and HN protein, respectively;

Figure 4 shows the restriction map of the PIV-3 HN gene;

Figure 5 shows the nucleotide (SEQ ID No: 5) and amino acid (SEQ ID No: 6) sequences of the RSV F gene and RSV F protein, respectively;

Figure 6 shows the restriction map of the RSV F gene;

Figure 7 shows the nucleotide (SEQ ID No: 7) and amino acid (SEQ ID No: 8) sequences of the RSV G gene and RSV G protein, respectively;

Figure 8 shows the restriction map of the RSV G gene;

Figure 9 shows the steps involved in the construction of an expression vector containing a chimeric F_{PIV-3} - F_{RSV} gene;

Figure 10 shows the steps involved in the construction of an expression vector containing a F_{PIV-3} gene lacking the 5'-untranslated sequence and transmembrane anchor and cytoplasmic tail coding regions;

Figure 11 shows the steps involved in the construction of an expression vector containing a chimeric F_{PIV-3} - F_{RSV} gene containing a truncated PIV-3 F gene devoid of 5'-untranslated region linked to a truncated RSV F1 gene;

Figure 12 shows the steps involved in construction of a modified pAC 610 baculovirus expression vector containing a chimeric F_{PIV-3} - F_{RSV} gene consisting of the PIV-3 F gene lacking both the 5'-untranslated sequence as well as transmembrane and cytoplasmic tail coding region linked to the truncated RSV F1 gene;

Figure 13 shows immunoblots of cell lysates from Sf9 cells infected with recombinant baculoviruses;

Figure 14 shows the steps involved in constructing a baculovirus transfer vector (pD2);

Figure 15 shows the steps involved in construction of a chimeric F_{RSV} - HN_{PIV-3} gene;

Figure 16 shows an SDS-PAGE gel and immunoblot of purified F_{RSV} - HN_{PIV-3} chimeric protein;

Figure 17 illustrates mutagenesis of a PIV-3 F gene; and

Figure 18 shows the steps involved in the construction of a chimeric F_{PIV-3} - G_{RSV} gene.

GENERAL DESCRIPTION OF INVENTION

In the present invention, a chimeric molecule protective against two different major childhood diseases is provided. The present invention specifically relates to the formulation of various recombinant Parainfluenza virus (PIV)/Respiratory syncytial virus (RSV) immunogens to produce safe and efficacious vaccines capable of protecting infants and young children, as well as other susceptible individuals, against diseases caused by infection with both PIV and RSV. However, as described above, the present invention extends to the construction of multimeric hybrid genes containing genes coding for protective antigens from many pathogens. Such vaccines may be administered in any desired manner, such as a readily-injectable vaccine, intranasally or orally.

In the present invention, the inventors have specifically engineered several model PIV/RSV chimeric genes containing relevant sequences from selected genes coding for PIV-3 and RSV surface glycoproteins linked in tandem. All genes in the chimeric constructs described herein were obtained from recent clinical isolates of PIV-3 and RSV. The chimeric gene constructs may include gene sequences from either PIV-3 F or HN genes linked in tandem to either RSV F or G genes in all possible relative orientations and combinations.

The chimeric gene constructs provided herein may consist of either the entire gene sequences or gene segments coding for immunogenic and protective epitopes thereof. The natural nucleotide sequence of these genes may be modified by mutation while retaining antigenicity and such modifications may include the removal of putative pre-transcriptional terminators to optimize their expression in eukaryotic cells. The genes were

designed to code for hybrid PIV-RSV surface glycoproteins linked in tandem in a single construct to produce gene products which elicit protective antibodies against both parainfluenza and respiratory syncytial viruses. Such multimeric hybrid genes consist of a gene sequence coding for a human PIV-3 F or HN protein or an immunogenic epitope-containing fragment thereof linked to a gene sequence coding for a human RSV G or F protein or an immunogenic epitope-containing fragment thereof. Specific gene constructs which may be employed include F_{PIV-3} - F_{RSV}, F_{RSV} - HN_{PIV-3} and F_{PIV-3} - G_{RSV} hybrid genes.

In addition, the present invention also extends to the construction of other multimeric genes, such as trimeric genes containing PIV and RSV genes or gene segments, linked in all possible relative orientations. For example:

F_{PIV} - HN_{PIV} - F or G_{RSV}

F_{PIV} - F_{RSV} - G_{RSV}

HN_{PIV} -F_{RSV} - G_{RSV}

The multimeric genes provided herein also may comprise at least one gene encoding at least one immunogenic and/or immunostimulating molecule.

The multimeric hybrid genes provided herein may be sub-cloned into appropriate vectors for expression in cellular expression systems. Such cellular expression systems may include bacterial, mammalian, insect and fungal, such as yeast, cells.

The chimeric proteins provided herein also may be presented to the immune system by the use of a live vector, including live viral vectors, such as recombinant poxviruses, adenoviruses, retroviruses, Semliki Forest viruses, and live bacterial vectors, such as Salmonella and mycobacteria (e.g. BCG).

Chimeric proteins, such as a PIV/RSV chimera, present in either the supernatants or cell lysates of

transfected, transformed or infected cells then can be purified in any convenient manner.

To evaluate the immunogenicity and protective ability of the chimeric proteins, suitable experimental animals are immunized with either varying doses of the purified chimeric proteins, such as the PIV/RSV chimera, and/or live recombinant vectors as described above. Such chimeric proteins may be presented to the immune system by either the use of physiologically-acceptable vehicles, such as aluminum phosphate, or by the use of delivery systems, such as ISCOMS and liposomes. The chimeras also may be formulated to be capable of eliciting a mucosal response, for example, by conjugation or association with immunotargeting vehicles, such as the cholera toxin B subunit, or by incorporation into microparticles. The vaccines may further comprise means for delivering the multimeric protein specifically to cells of the immune system, such as toxin molecules or antibodies. To further enhance the immunoprotective ability of the chimeric proteins, they may be supplemented with other immunogenic and/or immunostimulating molecules. The chimeric PIV/RSV proteins specifically described herein may be formulated with an adjuvant, such as aluminum phosphate, to produce readily-injectable vaccines for protection against the diseases caused by both PIV-3 and RSV. The chimeric proteins also may be administered intranasally or orally. The chimeric proteins may be used in test kits for diagnosis of infection by PIV-3 and RSV.

The invention is not limited to the preparation of chimeric PIV-3 and RSV proteins, but is applicable to the production of chimeric immunogens composed of either the entire sequences or regions of the immunogenic proteins from at least two pathogens sequentially linked in a single molecule. Chimeric antigens also may be synthesized to contain the immunodominant epitopes of

several proteins from different pathogens. These chimeric antigens may be useful as vaccines or as diagnostic reagents.

SEQUENCE IDENTIFICATION

Several nucleotide and amino acid sequences are referred to in the disclosure of this application. The following table identifies the sequences and the location of the sequence:

SEQ ID No.	Identification	Location
1	Nucleotide sequence for PCR-amplified PIV-3 F gene	Fig. 1, Example 1
2	Amino acid sequence for PCR-amplified PIV-F protein	Fig. 1, Example 1
3	Nucleotide sequence for PIV-3 HN gene	Fig. 3, Example 1
4	Amino acid sequence for PIV-3 HN protein	Fig. 3, Example 1
5	Nucleotide sequence for RSV F gene	Fig. 5, Example 1
6	Amino acid sequence for RSV F protein	Fig. 5, Example 1
7	Nucleotide sequence for RSV G gene	Fig. 7, Example 1
8	Amino acid sequence for RSV G protein	Fig. 7, Example 1
9	BsrI - BamHI oligo-nucleotide cassette	Fig. 9, Example 2
10	BspHI - BamHI oligo-nucleotide cassette	Fig. 9, Example 2
11	EcoRI - Ppu MI oligo-nucleotide cassette	Fig. 9, Example 2
12	BrsI - BamHI oligo-nucleotide cassette	Fig. 10, Example 3

13	EcoRI -Bsr BI oligo-nucleotide cassette	Fig. 10, Example 3
14	EcoRV - EcoRI oligo-nucleotide cassette	Fig. 11, Example 5
15	EcoRV - BamHI oligo-nucleotide cassette	Fig. 14, Example 8
16	BspHI - BspHI oligo-nucleotide cassette	Fig. 15, Example 9
17	Nucleotide sequence for PIV-3 F gene	Example 15
18	Mutagenic oligo-nucleotide #2721	Fig. 17, Example 15
19	Nucleotide sequence for part of oligo-nucleotide #2721	Example 15
20	Oligonucleotide probe	Example 15

DEPOSIT INFORMATION

Certain plasmid DNAs described and referred to herein have been deposited with the American Type Culture Collection (ATCC) located at Rockville, Maryland, USA, pursuant to the Budapest Treaty and prior to the filing of this application. The deposited purified plasmids will become available to the public upon grant of this U.S. patent application or upon publication of its corresponding European patent application, whichever first occurs. The invention described and claimed herein is not to be limited in scope by the plasmid DNAs of the constructs deposited, since the deposited embodiment is intended only as an illustration of the invention. The following purified plasmids were deposited at the ATCC with the noted accession numbers on December 17, 1992:

Plasmid	Example No.	Accession No.
pAC DR7	5	75387
pD2RF-HN	9	75388
pD2F-G	16	75389

Any equivalent plasmids that can be used to produce equivalent antigens as described in this application are within the scope of the invention.

EXAMPLES

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific Examples. These Examples are described solely for purposes of illustration and are not intended to limit the scope of the invention. Changes in form and substitution of equivalents are contemplated as circumstances may suggest or render expedient. Although specific terms have been employed herein, such terms are intended in a descriptive sense and not for purposes of limitations.

Methods for cloning and sequencing the PIV-3 and RSV genes as well as the procedures for sub-cloning the genes into appropriate vectors and expressing the gene constructs in mammalian and insect cells are not explicitly described in this disclosure but are well within the scope of those skilled in the art.

Example 1:

This Example outlines the strategy used to clone and sequence the PIV-3 F, HN and RSV F, G genes (from a type **A** isolate). These genes were used in the construction of the F_{PIV-3} - F_{RSV}, F_{RSV}-HN_{PIV-3}, and F_{PIV-3}-G_{RSV} chimeric genes detailed in Examples 2 to 4, 9 and 15, respectively.

Two PIV-3 F gene clones initially were obtained by PCR amplification of cDNA derived from viral RNA extracted from a recent clinical isolate of PIV-3. Two other PIV-3 F gene clones as well as the PIV-3 HN, RSV F and RSV G genes were cloned from a cDNA library prepared from mRNA isolated from MRC-5 cells infected with clinical isolates of either PIV-3 or RSV (type A isolate). The PIV-3 F (both PCR amplified and non-PCR amplified), PIV-3 HN, RSV F and RSV G gene clones were sequenced by the dideoxynucleotide chain termination

procedure. Sequencing of both strands of the genes was performed by a combination of manual and automated sequencing.

The nucleotide (SEQ ID No: 1) and amino acid (SEQ ID No: 2) sequences of the PCR amplified PIV-3 F gene and F protein, respectively, are presented in Figure 1 and the restriction map of the gene is shown in Figure 2. Sequence analysis of the 1844 nucleotides of two PCR amplified PIV-3 F gene clones confirmed that the clones were identical. Comparison of the coding sequence of the PCR-amplified PIV-3 F gene clone with that of the published PIV-3 F gene sequence revealed a 2.6% divergence in the coding sequence between the two genes resulting in fourteen amino acid substitutions.

The nucleotide sequence of the non-PCR amplified PIV-3 F gene clone differed from the PCR amplified gene clone in the following manner: the non-PCR amplified clone had ten additional nucleotides (AGGACAAAAG) at the 5′ untranslated region of the gene and differed at four positions, 8 (T in PCR-amplified gene to C in non-PCR amplified gene) , 512 (C in PCR-amplified gene to T in non-PCR amplified gene) , 518 (G in PCR-amplified gene to A in non-PCR amplified gene) and 1376 (A in PCR-amplified gene to G in non-PCR amplified gene). These changes resulted in three changes in the amino acid sequence of the F protein encoded by the non-PCR amplified PIV-3 F gene. Serine (position 110), glycine (position 112), and aspartic acid (position 398) in the primary amino acid sequence of the F protein encoded by the PCR amplified PIV-3 F gene was changed to phenylalanine (position 110), glutamic acid (position 112) and glycine (position 398), respectively, in the primary amino acid sequence of the F protein encoded by the PCR amplified clone.

Figure 3 shows the nucleotide (SEQ ID No: 3) and amino acid (SEQ ID No: 4) sequences of the PIV-3 HN gene and protein, respectively and the restriction map of the

gene is presented in Figure 4. Analysis of the 1833 nucleotide sequence from two HN clones confirmed that the sequences were identical. A 4.4% divergence in the coding sequence of the PIV-3 HN gene was noted when the sequence was compared to the published PIV-3 HN coding sequence. This divergence resulted in seventeen amino acid substitutions in the amino acid sequence of the protein encoded by the PIV-3 HN gene.

The nucleotide (SEQ ID No: 5) and amino acid (SEQ ID No: 6) sequences of the RSV F gene and RSV F protein, respectively, are shown in Figure 5 and the restriction map of the gene is shown in Figure 6. Analysis of the ~~1887~~ nucleotide sequence from two RSV F clones verified complete sequence homology between the two clones. Comparison of this nucleotide sequence with that reported for the RSV F gene revealed approximately 1.8% divergence in the coding sequence resulting in eleven amino acid substitutions.

The nucleotide (SEQ ID No: 7) and amino acid (SEQ ID No: 8) sequences of the RSV G gene and RSV G protein, respectively, are presented in Figure 7 while the restriction map of the gene is outlined in Figure 8. Comparison of the 920 nucleotide sequence of the G gene clone with the published G sequence (type A isolate) revealed a 4.2% divergence in the nucleotide sequence and a 6.7% divergence in the amino acid sequence of the gene product. This divergence resulted in twenty amino acid substitutions.

The full-length PIV-3 F (non-PCR amplified) , PIV-3 HN, RSV F and RSV G genes were cloned into λgt11 and subcloned into the multiple cloning site of a Bluescript M13-SK vector, either by blunt end ligation or using appropriate linkers. The PCR-amplified PIV-3 F gene was directly cloned into the Bluescript vector. The cloning vectors containing the PIV-3 F-PCR amplified, PIV-3 F non-PCR amplified, PIV-3 HN, RSV F and RSV G genes were

named pPI3F, pPI3Fc, pPIVHN, pRSVF and pRSVG, respectively.

Example 2:

This Example illustrates the construction of a Bluescript-based expression vector (pMCR20) containing the chimeric F_{PIV-3} - F_{RSV} gene. This chimeric gene construct contains the 5′ untranslated region of the PIV-3 F gene but lacks the hydrophobic anchor and cytoplasmic tail coding regions of both the PIV-3 and RSV F genes. The steps involved in the construction of this plasmid are summarized in Figure 9.

To prepare the PIV-3 portion of the chimeric gene (Figure 9, step 1), the full length PIV-3 gene lacking the transmembrane region and cytoplasmic tail coding regions was retrieved from plasmid pPI3F by cutting the polylinker with BamHI, blunt-ending the linearized plasmid with Klenow polymerase and cutting the gene with BsrI. A BsrI-BamHI oligonucleotide cassette (SEQ ID No: 9) containing a PpuMI site and three successive translational stop codons were ligated to the truncated 1.6 Kb [BamHI]-BsrI PIV-3 F gene fragment and cloned into the EcoRV-BamHI sites of a Bluescript M13-SK expression vector containing the human methallothionen promoter and the poly A and IVS sequences of the SV40 genome (designated pMCR20), to generate plasmid pME1.

To engineer the RSV F gene component of the chimeric construct (Figure 9, step 2), the RSV F gene lacking the transmembrane region and cytoplasmic tail coding regions was retrieved from plasmid pRSVF by cutting the polylinker with EcoRI and the gene with BspHI. A synthetic BspHI-BamHI oligonucleotide cassette (SEQ ID No: 10) containing three successive translational stop codons was ligated to the 1.6 Kb truncated RSV F gene and cloned into the EcoRI-BamHI sites of the Bluescript based expression vector, pMCR20 to produce plasmid pES13A. Plasmid pES13A then was cut with EcoRI and PpuMI to

remove the leader and F2 coding sequences from the truncated RSV F gene. The leader sequence was reconstructed using an EcoRI-PpuMI oligocassette (SEQ ID No: 11) and ligated to the RSV F1 gene segment to generate plasmid pES23A.

To prepare the chimeric F_{PIV-3}-F_{RSV} gene (Figure 9, step 3) containing the 5′ untranslated region of the PIV-3 F gene linked to the truncated RSV F1 gene fragment, plasmid pME1 (containing the 1.6 Kb truncated PIV-3 F gene) first was cut with PpuMI and BamHI. The PpuMI-BamHI restricted pME1 vector was dephosphorylated with intestinal alkaline phosphatase. The 1.1 Kb RSV F1 gene fragment was retrieved from plasmid pES23A by cutting the plasmid with PpuMI and BamHI. The 1.1 Kb PpuMI-BamHI RSV F1 gene fragment was cloned into the PpuMI-BamHI sites of the dephosphorylated pME1 vector to generate plasmid pES29A. This chimeric gene construct contains the 5′ untranslated region of the PIV-3 F gene but lacks the nucleotide sequences coding for the hydrophobic anchor domains and cytoplasmic tails of both the PIV-3 and RSV F proteins.

Example 3:

This Example illustrates the construction of a Bluescript-based expression vector containing the PIV-3 F gene lacking both the 5′ untranslated and transmembrane anchor and cytoplasmic tail coding regions. The steps involved in constructing this plasmid are outlined in Figure 10.

Plasmid pPI3F containing the full length PIV-3 F gene was cut with BamHI, blunt ended with Klenow polymerase and then cut with BsrI to remove the transmembrane and cytoplasmic tail coding regions. The Bluescript-based expression vector, pMCR20, was cut with SmaI and BamHI. A synthetic BsrI-BamHI oligonucleotide cassette (SEQ ID No: 12) containing a translational stop codon was ligated with the 1. 6 Kb blunt ended-BsrI PIV-3

F gene fragment to the SmaI-BamHI restricted pMCR20 vector to produce plasmid pMpFB. The PIV-3 F gene of this construct lacked the DNA fragment coding for the transmembrane and cytoplasmic anchor domains but contained the 5′ untranslated region. To engineer a plasmid containing the PIV-3 F gene devoid of both the 5′ untranslated region and the DNA fragment coding for the hydrophobic anchor domain, plasmid pMpFB was cut with EcoRI and BstBI. An EcoRI-BstBI oligocassette (SEQ ID No: 13) containing the sequences to reconstruct the signal peptide and coding sequences removed by the EcoRI-BstBI cut was ligated to the EcoRI-BstBI restricted pMpFB vector to produce plasmid pMpFA.

Example 4:

This Example illustrates the construction of the chimeric F_{PIV-3}-F_{RSV} gene composed of the truncated PIV-3 F gene devoid of the 5′ untranslated region linked to the truncated RSV F1 gene. The steps involved in constructing this plasmid are summarized in Figure 11.

To prepare this chimeric gene construct, plasmid pES29A (Example 2) was cut with BstBI and BamHI to release the 2.5 Kb BstBI-BamHI PI3-3 F-RSV F1 chimeric gene fragment. This BstBI-BamHI fragment was isolated from a low melting point agarose gel and cloned into the BstBI-BamHI sites of the dephosphorylated vector pMpFA to produce plasmid pES60A. This construct contained the PIV-3 F gene lacking both the 5′ untranslated region and the hydrophobic anchor and cytoplasmic tail coding sequences linked to the F1 coding region of the truncated RSV F gene. This chimeric gene was subsequently subcloned into the baculovirus transfer vector (see Example 5).

Example 5:

This Example illustrates the construction of the modified pAC 610 baculovirus transfer vector containing the native polyhedrin promoter and the chimeric F_{PIV-3}-F_{RSV}

gene consisting of the PIV-3 F gene lacking both the 5' untranslated sequence and the nucleotide sequence coding for the hydrophobic anchor domain and cytoplasmic tail linked to the truncated RSV F1 gene. Construction of this plasmid is illustrated in Figure 12.

The pAC 610 baculovirus expression vector was modified to contain the native polyhedrin promoter in the following manner. Vector pAC 610 was cut with EcoRV and BamHI. The 9.4 Kb baculovirus transfer vector lacking the EcoRV-BamHI DNA sequence was isolated from a low melting point agarose gel and treated with intestinal alkaline phosphatase. In a 3-way ligation, an EcoRV-EcoRI oligonucleotide cassette (SEQ ID No: 14) containing the nucleotides required to restore the native polyhedrin promoter was ligated with the 1.6 Kb EcoRI-BamHI truncated RSV F gene fragment isolated from construct pES13A (Example 2, step 2) and the EcoRV-BamHI restricted pAC 610 phosphatased vector to generate plasmid pES47A. To prepare the pAC 610 based expression vector containing the chimeric F_{PIV-3}-F_{RSV} gene, plasmid pES47A was first cut with EcoRI and BamHI to remove the 1.6 Kb truncated RSV F gene insert. The 2.8 Kb F_{PIV-3}-F_{RSV} chimeric gene was retrieved by cutting plasmid pES60A (Example 4) with EcoRI and BamHI. The 2.8 Kb EcoRI-BamHI chimeric gene was ligated to the EcoRI-BamHI restricted pES47A vector to generate plasmid pAC DR7 (ATCC 75387).

Example 6

This Example outlines the preparation of plaque-purified recombinant baculoviruses containing the chimeric F_{PIV-3} - F_{RSV} gene.

Spodoptera frugiperda (Sf9) cells were co-transfected with 1.0 μg wild-type AcMNPV DNA and 2.5 μg of F_{PIV-3}-F_{RSV} plasmid DNA (plasmid pAC DR7 - Example 5). Putative recombinant baculoviruses (purified once by serial dilution) containing the F_{PIV-3} - F_{RSV} chimeric gene were identified by dot-blot hybridization. Lysates of

insect cells infected with the putative recombinant baculoviruses were probed with the ^{32}P-labelled F_{PIV-3}-F_{RSV} chimeric gene insert. Recombinant baculoviruses were plaque-purified twice before being used for expression studies. All procedures were carried out according to the protocols outlined by M.D. Summers and G.E. Smith in "A Manual of Methods for Baculovirus Vectors and Insect Cell Culture Procedures", Texas Agricultural Experiment Station, Bulletin 1555, 1987.

Example 7:

This Example illustrates the presence of the chimeric F_{PIV-3} -F_{RSV} protein in supernatants and cell lysates of infected Sf9 cells.

Insect cells were infected with the plaque-purified recombinant baculoviruses prepared as described in Example 6 at a m.o.i. of 8. Concentrated supernatants from cells infected with the recombinant viruses were positive in a PIV-3 F specific ELISA. In addition, when lysates from ^{35}S-methioninelabelled infected cells were subjected to SDS-polyacrylamide gel electrophoresis and gels were analyzed by autoradiography, a strong band with apparent molecular weight of approximately 90 kDa was present in lysates of cells infected with the recombinant viruses but was absent in the lysates from wild-type infected cells. The presence of the chimeric F_{PIV-3} - F_{RSV} protein in the lysates of cells infected with the recombinant baculoviruses was confirmed further by Western blot analysis using monospecific anti-PIV-3 F and anti-RSV F antisera and/or monoclonal antibodies (Mabs). Lysates from cells infected with the recombinant baculoviruses reacted with both anti-PIV-3 and anti-RSV antisera in immunoblots. As shown in the immunoblot of Figure 13, lysates from cells infected with either the RSV F or F_{PIV-3} - F_{RSV} recombinant baculoviruses reacted positively with the anti-F RSV Mab. As expected, lysates from cells infected with wild type virus did not react

with this Mab. In addition, only lysates from cells infected with the chimeric F_{PIV-3} - F_{RSV} recombinant viruses reacted with the anti-PIV-3 F_1 antiserum.

Example 8

This Example illustrates modification of the baculovirus transfer vector pVL1392 (obtained from Invitrogen), wherein the polyhedrin ATG start codon was converted to ATT and the sequence CCG was present downstream of the polyhedrin gene at positions +4,5,6. Insertion of a structural gene several base pairs downstream from the ATT codon is known to enhance translation. The steps involved in constructing this modified baculovirus transfer vector are outlined in Figure 14.

The baculovirus expression vector pVL1392 was cut with EcoRV and BamHI. The 9.5 kb restricted pVL1392 vector was ligated to an EcoRV-BamHI oligonucleotide cassette (SEQ ID No: 15) to produce the pD2 vector.

Example 9:

This Example illustrates the construction of the pD2 baculovirus expression vector containing the chimeric F_{RSV} -HN_{PIV-3} gene consisting of the truncated RSV F and PIV-3 HN genes linked in tandem. The steps involved in constructing this plasmid are summarized in Figure 15.

To engineer the F_{RSV}-HN_{PIV-3} gene, the RSV F gene lacking the nucleotide sequence coding for the transmembrane domain and cytoplasmic tail of the RSV F glycoprotein was retrieved from plasmid pRSVF (Example 1) by cutting the polylinker with EcoRI and the gene with BspHI. The PIV-3 HN gene devoid of the DNA fragment coding for the hydrophobic anchor domain was retrieved from plasmid pPIVHN (Example 1) by cutting the gene with BspHI and the polylinker with BamHI. The 1.6 Kb EcoRI-BspHI RSV F gene fragment and the 1.7 Kb BspHI-BamHI PIV-3 HN gene fragment were isolated from low melting point agarose gels. For cloning purposes, the two BspHI sites

in the Bluescript based mammalian cell expression vector, pMCR20, were mutated. Mutations were introduced in the BspHI sites of the pMCR20 by cutting the expression vector with BspHI, treating both the BspHI restricted vector and the 1. 1 Kb fragment released by the BspHI cut with Klenow polymerase and ligating the blunt-ended 1.1 Kb fragment to the blunt-ended Bluescript-based expression vector to generate plasmid pM'. Since insertion of the 1.1 Kb blunt-end fragment in the mammalian cell expression vector in the improper orientation would alter the Amp' gene of the Bluescript-based expression vector, only colonies of HB101 cells transformed with the pM' plasmid DNA with the 1.1 Kb blunt-ended fragment in the proper orientation could survive in the presence of ampicillin. Plasmid DNA was purified from ampicillin-resistant colonies of HB101 cells transformed with plasmid PM' by equilibrium centrifugation in cesium chloride-ethidium bromide gradients. The 1.6 Kb EcoRI-BspHI RSV F and 1.7 Kb BspHI-BamHI PIV-3 HN gene fragments were directly cloned into the EcoRI-BamHI sites of vector pM' in a 3-way ligation to generate plasmid pM' RF-HN.

To restore specific coding sequences of the RSV F and PIV-3 HN genes removed by the BspHI cut, a BspHI-BspHI oligonucleotide cassette (SEQ ID No: 16) containing the pertinent RSV F and PIV-3 HN gene sequences was ligated via the BspHI site to the BspHI-restricted plasmid pM' RF-HN to produce plasmid pM RF-HN. Clones containing the BspHI-BspHI oligonucleotide cassette in the proper orientation were identified by sequence analysis of the oligonucleotide linker and its flanking regions.

To clone the chimeric F_{RSV}-HN_{PIV-3} gene into the baculovirus expression vector pD2 (Example 8), the F_{RSV}-HN_{PIV-3} truncated gene first was retrieved from plasmid pM RF-HN by cutting the plasmid with EcoRI. The 3.3 Kb F_{RSV}-

HN_{PIV-3} gene then was cloned into the EcoRI site of the baculovirus transfer vector plasmid pD2 to generate plasmid pD2 RF-HN (ATCC 75388). Proper orientation of the 3.3 Kb EcoRI F_{RSV}-HN_{PIV-3} chimeric gene insert in plasmid pD2 RF-HN was confirmed by sequence analysis.

Example 10:

This Example outlines the preparation of plaque-purified recombinant baculoviruses containing the chimeric F_{RSV}-HN_{PIV-3} gene.

Spodoptera frugiperda (Sf9) cells were co-transfected with 1 μg wild-type AcNPV DNA and 2 μg of F_{RSV}-HN_{PIV-3} plasmid DNA (plasmid pD2 RF-HN-Example 9). Putative recombinant baculoviruses (purified once by serial dilution) containing the F_{RSV}-HN_{PIV-3} chimeric gene were identified by dot-blot hybridization. Lysates of insect cells infected with the putative recombinant baculoviruses were probed with the ^{32}P-labelled RSV F or PTV-3 HN gene oligonucleotide probes. Recombinant baculoviruses were plaque-purified three times before being used for expression studies. All procedures were carried out according to the protocols outlined by Summers and Smith (Example 6).

Example 11:

This Example illustrates the presence of the chimeric F_{RSV}-HN_{PIV-3} protein in supernatants of infected Sf9 and High 5 cells.

Insect cells (Sf9 and High 5), maintained in serum free medium EX401, were infected with the plaque purified recombinant baculoviruses of Example 10 at a m.o.i. of 5 to 10 pfu/cell. Supernatants from cells infected with the recombinant baculoviruses tested positive for expressed protein in both the RSV-F and PIV-3 HN specific ELISAS. In addition, supernatants from infected cells reacted positively with both an anti-F RSV monoclonal antibody and anti-HN peptide antisera on immunoblots. A distinct band of approximately 105 kDa was present in the

immunoblots. These results confirm the secretion of the chimeric F_{RSV}-HN_{PIV-3} protein into the supernatant of Sf9 and High 5 cells infected with the recombinant baculoviruses.

Example 12:

This Example illustrates the purification of the chimeric F_{RSV}-HN_{PIV-3} protein from the supernatants of infected High 5 cells.

High 5 cells, maintained in serum free medium, were infected with the plaque purified recombinant baculoviruses of Example 10 at a m.o.i of 5 pfu/cell. The supernatant from virus infected cells was harvested 2 days post-infection. The soluble F_{RSV}-HN_{PIV-3} chimeric protein was purified from the supernatants of infected cells by immunoaffinity chromatography using an anti-HN PIV-3 monoclonal antibody. The anti-HN monoclonal antibody was coupled to CNBr-activated Sepharose 4B by conventional techniques. The immunoaffinity column was washed with 10 bed volumes of washing buffer (10mM Tris-HCl pH 7.5, 150 mM NaCl, 0.02% v/v Triton-X 100) prior to use. After sample loading, the column was washed with 10 bed volumes of washing buffer followed by 3 bed volumes of high salt buffer (10mm Tris-HCl pH 7.5, 500mM NaCl, 0.02% v/v Triton-X 100) . The chimeric F_{RSV}-HN_{PIV-3} protein was eluted from the immunoaffinity column with 100 MM glycine, pH 2.5, in the presence of 0.02% Triton X-100. Eluted protein was neutralized immediately with 1M Tris-HCl, pH 10.7.

Polyacrylamide gel electrophoretic analysis (Fig. 16, panel A) of the immunoaffinity-purified F_{RSV}-HN_{PIV-3} protein revealed the presence of one major protein band with an apparent molecular weight of 105 kDa. The purified protein reacted with both an anti-RSV F monoclonal antibody and anti-HN peptide antisera on immunoblots (Fig. 16, panel B, lanes 1 and 2, respectively).

Example 13:

This Example illustrates the immunogenicity of the F_{RSV}-HN_{PIV-3} protein in guinea pigs.

Groups of four guinea pigs were injected intramuscularly with either 1.0 or 10.0 μg of the chimeric F_{RSV}-HN_{PIV-3} protein purified as described in Example 12 and adjuvanted with aluminum phosphate. Groups of control animals were immunized with either placebo, or live PIV-3 or RSV (administered intranasally). Guinea pigs were bled 2 and 4 weeks after the primary injection and boosted at 4 weeks with an equivalent dose of the antigen formulation. Serum samples also were taken 2 and 4 weeks after the booster dose. To assess the ability of the chimeric protein to elicit PIV-3 and RSV-specific antibody responses, sera samples were analyzed for the presence of PIV-3 specific hemagglutination inhibiting and neutralizing antibodies as well as RSV neutralizing antibodies. As summarized in Table 1 below (the Tables appear at the end of the disclosure), the sera of animals immunized with two 10 μg doses of the chimeric protein had titres of PIV-3 specific hemagglutination inhibition (HAI) and PIV-3/RSV neutralizing antibodies at the 6 and 8 week time points which were equivalent to the levels obtained following intranasal inoculation with either live PIV-3 or RSV. In addition, animals immunized with only two 1 ug doses of the chimeric protein elicited strong PIV-3 and RSV specific neutralizing antibodies. These results confirmed the immunogenicity of both the RSV and PIV-3 components of the chimeric protein and provided confirmatory evidence that a single recombinant immunogen can elicit neutralizing antibodies against both RSV and PIV-3.

Example 14:

This Example illustrates the immunogenicity and protective ability of the F_{RSV}-HN_{PIV-3} protein in cotton rats.

Groups of eight cotton rats were injected intramuscularly with either 1.0 or 10.0 ug of the chimeric F_{RSV}-HN_{PIV-3} protein (prepared as described in Example 12) adjuvanted with aluminum phosphate. Groups of control animals were immunized with either placebo (PBS + aluminum phosphate) or live PIV-3 or RSV (administered intranasally). Cotton rats were bled 4 weeks after the primary injection and boosted at 4 weeks with an equivalent dose of the antigen formulation. Serum samples were also taken 1 week after the booster dose. As shown in Table 2 below, data from the 4-week bleed demonstrated that both a 1 and 10 μg dose of the chimeric protein was capable of inducing a strong primary response. Reciprocal mean $\log_2$ PIV-3 specific HAI and PIV-3/RSV neutralizing titers were equivalent to the titres obtained with live PIV-3 and RSV. Thus, a single inoculation of the chimeric protein was sufficient to elicit neutralizing antibodies against both PIV-3 and RSV. Strong neutralizing PIV-3 and RSV titres also were observed following the booster dose (5 week bleed). These results provide additional evidence that both the RSV and PIV-3 components of the chimeric protein are highly immunogenic.

To assess the ability of the chimeric immunogen to simultaneously protect animals against both RSV and PIV-3, four cotton rats from each group were challenged intranasally with 100 $TCID_{50}$ units of either PIV-3 or RSV. Animals were killed 4 days after virus challenge. Virus titers were determined in lung ~~nomogenates~~. As shown in Table 3 below, animals immunized with either 1 or 10 μg of the chimeric F_{RSV}-HN_{PIV-3} protein were completely protected against challenge with either PIV-3 or RSV. These results provide evidence that the chimeric protein

is not only highly immunogenic but can also simultaneously protect cotton rats against disease caused by both PIV-3 and RSV infection.

Example 15:

This Example illustrates the construction of a Bluescript M13-SK vector containing the chimeric F_{PIV-3}-G_{RSV} gene. This chimeric gene construct contains the 5′ untranslated region of a mutated PIV-3 F gene but lacks the nucleotide sequence coding for the hydrophobic anchor and cytoplasmic tail domains of both a mutated PIV-3 F and the native RSV G genes. The steps involved in constructing this plasmid are outlined in Figures 17 and 18.

The first step (Fig. 17) involved in preparing the PIV-3 F component of the chimeric F_{PIV-3}-G_{RSV} gene construct was to eliminate the putative pre-termination sites within the 18 nucleotide long sequence 5′ CAAGAAAAAGGAATAAAA 3′ (SEQ ID No: 17) located between positions 857 and 874 of the non PCR-amplified PIV-3 F gene and positions 847 and 864 of the PCR-amplified PIV-3 F gene (see Figure 1). To this end, the PIV-F cDNA of the non-PCR amplified PIV-3 F gene was cut at the BsaAI and EcoRI sites. The BsaAI-EcoRI PIV F gene fragment was cloned into the EcoRI site of a Bluescript M13-SK vector using an EcoRI-BsaAI linker. The 857-874 target region of the PIV-3 F gene (non-PCR amplified) then was mutated by oligonucleotide-mediated mutagenesis using the method of Morinaga et al. [1984, Biotechnology 2: 636-639]. Plasmid pPI3Fc (Example 1) was cut with ScaI in the Amp^r gene and dephosphorylated with alkaline phosphatase (plasmid #1). A second sample of plasmid pPI3Fc was cut with BstEII and NsiI to produce a 3.9 Kb restricted plasmid, lacking the 0.9 Kb BstEII-NsiI fragment of the PIV-3 F gene (plasmid #2). A mutagenic 78-mer synthetic oligonucleotide (#2721 shown in Fig. 17-SEQ ID No: 18)) containing the sequence 5′ CAGGAGAAGGGTATCAAG 3′ (SEQ ID

No: 19) was synthesized to specifically mutate the 857-874 DNA segment without changing the F protein sequence. This oligonucleotide was added to plasmid DNAs #1 and #2, denatured at 100°C for 3 min. and renatured by gradual cooling. The mixture then was incubated in the presence of DNA polymerase, dNTPs and T4 ligase and transformed into HB101 cells. Bacteria containing the 1.8 Kb mutated PIV-3 F gene were isolated on YT agar plates containing 100 μg/ml ampicillin. Hybridization with the oligonucleotide probe 5′ AGGAGAAGGGTATCAAG 3′ (SEQ ID No: 20) was used to confirm the presence of the mutated PIV-3 F gene. The mutated gene sequence was confirmed by DNA sequencing. The plasmid containing the mutated PIV-3 gene was designated pPI3Fm.

The second step (Fig. 18) in the engineering of the chimeric gene construct involved constructing a Bluescript based vector to contain the truncated PIV-3 Fm gene lacking the nucleotide sequence coding for the transmembrane anchor domain and cytoplasmic tail of the PIV-3 F protein linked in tandem with the RSV G gene lacking both the 5′ leader sequence and the nucleotide sequence coding for the transmembrane anchor domain and cytoplasmic tail of the G glycoprotein.

To prepare this chimeric gene, the orientation of the mutated PIV-F gene in plasmid pPI3Fm first was reversed by EcoRI digestion and religation to generate plasmid pPI3Fmr. To prepare the PIV-3 F gene component of the chimeric gene, plasmid pPI3Fmr was cut with NotI and BsrI to release the 1.7 Kb truncated PIV-3 F gene. To prepare the RSV G component, the 0.95 Kb RSV-G gene lacking both the 5′ leader sequence and the DNA segment encoding the G protein anchor domain and cytoplasmic tail was released from plasmid pRSVG (Example 1) by cutting the polylinker with EcoRI and the gene with BamHI. The 0.95 Kb EcoRI-BamHI RSV G gene fragment was subcloned into the EcoRI-BamHI sites of a restricted Bluescript

vector, pM13-SK, to produce plasmid pRSVGt. The 0.95 Kb EcoRI-BamHI G gene fragment and the 1.5 Kb NotI-BsrI truncated PIV-3 F gene were linked via a BsrI-BamHI oligonucleotide cassette (SEQ ID No: 9) restoring the F and G gene coding sequences and cloned into the pRSVGt vector restricted with BamHI and NotI in a 3-way ligation. The plasmid thus generated was designated pFG.

Example 16:

This Example outlines the construction of the pD2 baculovirus transfer vector (described in Example 8) containing the chimeric F_{PIV-3}-G_{RSV} gene consisting of a mutated PIV-3 F gene lacking the hydrophobic anchor and cytoplasmic coding regions linked to the RSV G gene lacking both the 5′ leader sequence and the nucleotide sequences encoding the transmembrane anchor domain and cytoplasmic tail of the G protein.

To prepare this construct, plasmid pFG (Example 15) was cut with EcoRI to release the 2.6 Kb F_{PIV-3}-G_{RSV} chimeric gene. The 2.6 Kb EcoRI restricted chimeric gene fragment then was sub-cloned into the EcoRI site of the dephosphorylated pD2 vector to generate the 12.1 Kb plasmid pD2F-G (ATCC 75389).

Example 17:

This Example outlines the preparation of plaque-purified recombinant baculoviruses containing the chimeric F_{PIV-3}-G_{RSV} gene.

Spodoptera frugiperda (Sf9) cells were co-transfected with 2 ug of pD2F-G plasmid DNA (Example 16) and 1 ug of linear wild-type AcNPV DNA (obtained from Invitrogen). Recombinant baculoviruses containing the F_{PIV-3}-G_{RSV} gene were plaque-purified twice according to the procedure outlined in Example 10.

Example 18:

This Example illustrates the presence of the chimeric F_{PIV-3}-G_{RSV} protein in the supernatant of Sf9 and High 5 cells infected with the recombinant baculoviruses.

Sf9 and High 5 cells were infected with recombinant baculoviruses containing the F_{PIV-3}-G_{RSV} gene (Example 16) at a m.o.i. of 5 to 10 pfu/cell. The supernatant of cells infected with the recombinant viruses tested positive for expressed protein in the PIV-3 F specific ELISA. Supernatants of infected cells reacted with both anti-F PIV-3 and anti-G RSV monoclonal antibodies in immunoblots. These results confirm the presence of the chimeric F_{PIV-3}-G_{RSV} protein in the supernatants of infected Sf9 and High 5 cells.

Example 19:

This Example outlines the preparation of recombinant vaccinia viruses expressing the F_{PIV-3} - F_{RSV} and F_{RSV} - HN_{PIV-3} genes.

Vaccinia virus recombinant viruses expressing the F_{PIV-3}-F_{RSV} (designated vP1192) and F_{RSV}-HN_{PIV-3} (designated vP1195) genes were produced at Virogenetics Corporation (Troy, NY) (an entity related to assignee hereof) using the COPAK host-range selection system. Insertion plasmids used in the COPAK host-range selection system contained the vaccinia K1L host-range gene [Perkus et al., (1990) Virology 179:276-286] and the modified vaccinia H6 promoter [Perkus et al. (1989), J. Virology 63:3829-3836]. In these insertion plasmids, the K1L gene, H6 promoter and polylinker region are situated between Copenhagen strain vaccinia flanking arms replacing the ATI region [open reading frames (ORFs) A25L, A26L; Goebel et al., (1990), Virology 179: 247-266; 517-563]. COPAK insertion plasmids are designed for use in in vivo recombination using the rescue virus NYVAC (vP866) (Tartaglia et al., (1992) Virology 188: 217-232). Selection of recombinant viruses was done on rabbit kidney cells.

Recombinant viruses, vP1192 and vP1195 were generated using insertion plasmids pES229A-6 and PSD.RN, respectively. To prepare plasmid pES229A-6 containing

the F_{PIV-3}-F_{RSV} gene, the COPAK-H6 insertion plasmid pSD555 was cut with SmaI and dephosphorylated with intestinal alkaline phosphatase. The 2.6 Kb F_{PIV-3}-F_{RSV} gene was retrieved from plasmid pES60A (Example 4) by cutting the plasmid with EcoRI and BamHI. The 2.6 Kb EcoRI-BamHI F_{PIV-3}-F_{RSV} gene was blunt ended with Klenow polymerase, isolated from a low melting point agarose gel and cloned into the SmaI site of the COPAK-H6 insertion plasmid pSD555 to generate plasmid pES229A-6. This positioned the F_{PIV-3}-F_{RSV} ORF such that the 5′ end is nearest the H6 promoter.

To prepare plasmid PSD.RN, the pSD555 vector first was cut with SmaI and BamHI. Plasmid pM RF-HN (Example 9) containing the truncated F_{RSV}-HN_{PIV-3} gene was cut with ClaI, blunt ended with Klenow polymerase and then cut with BamHI. The 3.3 Kb F_{RSV}-HN_{PIV-3} gene was cloned into the SmaI-BamHI sites of the pSD555 vector to generate plasmid PSD.RN. This positioned the F_{RSV}-HN_{PIV-3} ORF such that the H6 5′ end is nearest the H6 promoter.

Plasmids pES229A-6 and PSD.RN were used in in vitro recombination experiments in vero cells with NYVAC (vP866) as the rescuing virus. Recombinant progeny virus was selected on rabbit kidney (RK)-13 cells (ATCC #CCL37). Several plaques were passaged two times on RK-13 cells. Virus containing the chimeric genes were confirmed by standard in situ plaque hybridization [Piccini et al. (1987), Methods in Enzymology, 153:545-563] using radiolabeled probes specific for the PIV and RSV inserted DNA sequences. Plaque purified virus containing the F_{PIV-3}-F_{RSV} and F_{RSV}-HN_{PIV-3} chimeric genes were designated vP1192 and vP1195, respectively.

Radioimmunoprecipitation was done to confirm the expression of the chimeric genes in vP1192 and vP1195 infected cells. These assays were performed with lysates prepared from infected Vero cells [according to the procedure of Taylor et al., (1990) J. Virology 64, 1441-

1450] using guinea pig monospecific PIV-3 anti-HN and anti-F antiserum and rabbit anti-RSV F antiserum. Both the anti-PIV F and anti-RSV F antisera precipitated a protein with an apparent molecular weight of approximately 90 koa from vP1192 infected Vero cells. Both anti-RSV F and guinea pig anti-PIV HN antisera precipitated a protein with an apparent molecular weight of approximately 100 kDa from vP1195 infected cells. These results confirmed the production of the F_{PIV-3}-F_{RSV} and F_{RSV}-HN_{PIV-3} chimeric proteins in Vero cells infected with the recombinant poxviruses.

SUMMARY OF DISCLOSURE

In summary of the disclosure, the present invention provides multimeric hybrid genes which produce chimeric proteins capable of eliciting protection against infection by a plurality of pathogens, particularly PIV and RSV. Modifications are possible within the scope of this invention.

Table 1 Secondary antibody response of guinea pigs immunized with the chimeric F_{RSV}-HN_{PIV-3} protein

Antigen Formulation	Dose (ug)	HAI Titre[a] (log_2 ± s.e.)		Neutralization Titre[b] (log_2 ± s.e.)			
		PIV-3		PIV-3		RSV	
		6 wk Bleed	8 wk Bleed	6 wk Bleed	8 wk Bleed	6 wk Bleed	8 wk Bleed
Buffer	-	<1.0 ± 0.0	<1.0 ± 0.0	<1.0 ± 0.0	<1.0 ± 0.0	<1.0 ± 0.0	<1.0 ± 0.0
F_{RSV}-HN_{PIV-3}	10.0	9.1 ± 0.3	9.1 ± 0.3	7.1 ± 0.3	7.1 ± 0.5	5.5 ± 0.9	4.5 ± 1.2
	1.0	7.0 ± 2.0	7.3 ± 2.2	5.0 ± 1.5	4.5 ± 1.4	4.5 ± 0.5	3.0 ± 1.0
Live PIV-3		8.6 ± 0.7	7.3 ± 0.6	7.0 ± 0.4	7.3 ± 0.6	N/A	N/A
Live RSV		N/A[c]	N/A	N/A	N/A	5.5 ± 1.5	5.0 ± 1.0

[a] Reciprocal mean log_2 serum dilution which inhibits erythrocyte agglutination by 4 hemagglutinating units of PIV-3

[b] Reciprocal mean log_2 serum dilution which blocks hemadsorption of 100 $TCID_{50}$ units of PIV-3 or RSV

[c] N/A - not applicable

TABLE 2

Table 2: Serum antibody response of cotton rats immunized with the chimeric F_{RSV}-HN_{PIV-3} protein[a]

Antigen Formulation	Dose (ug)	HAI Titre[b] (log_2 ± s.d.)		Neutralization Titre[c] (log_2 ± s.d.)			
		PIV-3		PIV-3		RSV	
		4 wk Bleed	5 wk Bleed	4 wk Bleed	5 wk Bleed	4 wk Bleed	5 wk Bleed
Buffer	-	2.8 ± 0.5	<3.0 ± 0.0	<1.0 ± 1.0	<1.0 ± 0.0	1.8 ± 0.3	0.8 ± 0.7
F_{RSV}-HN_{PIV-3}	10.0	9.5 ± 1.3	10.5 ± 0.6	>9.0 ± 0.0	>9.0 ± 0.0	5.2 ± 1.1	5.8 ± 0.9
	1.0	9.3 ± 1.0	10.3 ± 0.5	>9.0 ± 0.0	>9.0 ± 0.0	5.0 ± 0.7	5.8 ± 1.2
Live PIV-3		7.0 ± 0.0	8.5 ± 0.7	>9.0 ± 0.0	9.2 ± 0.7	N/A	N/A
Live RSV		N/A[d]	N/A	N/A	N/A	5.5 ± 0.6	8.5 ± 0.6

[a] Each value represents the mean titre of antisera from 8 animals.

[b] Reciprocal mean log_2 serum dilution which inhibits erythrocyte agglutination by 4 hemagglutinating units of PIV-3

[c] Reciprocal mean log_2 serum dilution which blocks hemadsorption of 100 $TCID_{50}$ units of PIV-3 or RSV

[d] N/A - not applicable

Table 3. Response of immunized cotton rats to PIV/RSV challenge[a]

Antigen Formulation	Dose (ug)	Mean virus lung titre $\log_{10}$/g lung ± s.d.	
		RSV	PIV-3
Buffer	-	3.7 ± 0.3	3.4 ± 0.3
F_{RSV}-HN_{PIV-3}	10.0	≤1.5 ± 0.0	≤1.5 ± 0.0
F_{RSV}-HN_{PIV-3}	1.0	≤1.5 ± 0.0	≤1.5 ± 0.0
Live RSV		≤1.5 ± 0.0	≤1.5 ± 0.0
Live PIV-3		≤1.5 ± 0.0	≤1.5 ± 0.0

[a] Animals were challenged intranasally with 100 $TCID_{50}$ units of PIV-3 or RSV and killed 4 days later. Each value represents the mean virus lung titre of 4 animals.

CLAIMS

What we claim is:

1. A multimeric hybrid gene, comprising a gene sequence coding for an antigenic region of a protein from a first pathogen linked to a gene sequence coding for an antigenic region of a protein from a second pathogen.

2. The hybrid gene of claim 1 wherein said first and second pathogens are selected from bacterial and viral pathogens.

3. The hybrid gene of claim 2 wherein both said first and second pathogens are viral pathogens.

4. The hybrid gene of claim 1 wherein said first and second pathogens are selected from those causing different respiratory tract diseases.

5. The hybrid gene of claim 4 wherein said first and second pathogens causing different respiratory tract diseases are selected from the paramoxyviridae family of viruses.

6. The hybrid gene of claim 1 wherein at least one of said gene sequences is mutated while retaining antigenicity.

7. The hybrid gene of claim 6 wherein said mutation is at a putative pre-termination site.

8. The hybrid gene of claim 1 wherein said first pathogen is parainfluenza virus (PIV) and said second pathogen is respiratory syncytial virus (RSV).

9. The hybrid gene of claim 1, comprising at least one gene sequence coding for a parainfluenza virus (PIV) protein linked to at least one gene sequence coding for a respiratory syncytial virus (RSV) protein.

10. The hybrid gene of claim 9, wherein said parainfluenza virus protein is selected from PIV-3 F and HN proteins and said respiratory syncytial virus protein is selected from RSV G and F proteins.

11. The hybrid gene of claim 1 consisting of a gene sequence coding for a human PIV-3 F or HN protein or an

immunogenic epitope-containing fragment thereof linked to a gene sequence coding for a human RSV G or F protein or an immunogenic epitope-containing fragment thereof.

12. The hybrid gene of claim 11 which is selected from F_{PIV-3} - F_{RSV}, F_{RSV} - HN_{PIV-3} and F_{PIV-3} - G_{RSV} hybrid genes.

13. The hybrid gene of claim 1 contained in an expression vector.

14. The hybrid gene of claim 13 in the form of plasmid pAC DR7, pD2 RF-HN or pD2 F-G.

15. The hybrid gene of claim 1 further comprising at least one gene encoding at least one immunogenic and/or immunostimulating molecule.

16. Cells containing the multimeric hybrid gene of claim 1 for expression of a chimeric protein encoded by said gene.

17. The cells of claim 16 which are bacterial cells, mammalian cells, insect cells, yeast cells or fungal cells.

18. A chimeric protein, comprising an antigenic region of a protein from a first pathogen linked to an antigenic region of a protein from a second pathogen.

19. The protein of claim 18, wherein said first and second pathogens are selected from bacterial and viral pathogens.

20. The protein of claim 19 wherein both said first and second pathogens are viral pathogens.

21. The protein of claim 18, wherein said first and second pathogens are selected from those causing different respiratory tract diseases.

22. The protein of claim 21 wherein said first and second pathogens causing different respiratory tract diseases are selected from the paramoxyviridae family of viruses.

23. The protein of claim 18, wherein said first pathogen is parainfluenza virus (PIV) and said second pathogen is respiratory syncytial virus (RSV).

24. The protein of claim 18 comprising at least one parainfluenza virus (PIV) protein linked to at least one respiratory syncytial virus (RSV) protein.

25. The protein of claim 24, wherein said PIV protein is selected from PIV-3 F and HN proteins and said RSV protein is selected from RSV G and F proteins.

26. The protein of claim 18 consisting of a human parainfluenza virus-3 (PIV-3) F or HN protein or an immunogenic epitope-containing fragment thereof linked to a human respiratory syncytial virus (RSV) G or F protein or an immunogenic epitope-containing fragment thereof.

27. The protein of claim 26 which is selected from F_{PIV-3} - F_{RSV}, F_{RSV} - HN_{PIV-3} and F_{PIV-3} - G_{RSV} chimeric proteins.

28. A process for preparation of a chimeric protein which comprises:

isolating a gene sequence coding for an antigenic region of a protein from a first pathogen,

isolating a gene sequence coding for an antigenic region of a protein from a second pathogen,

linking said gene sequences to form a multimeric

hybrid gene, and expressing the multimeric hybrid gene in a cellular expression system

29. The process of claim 28 wherein said multimeric hybrid gene comprises a gene sequence coding for a PIV-F or HN protein or an immunogenic epitope-containing fragment thereof linked to a gene sequence coding for a human RSV G or F protein or an epitope-containing fragment thereof.

30. The process of claim 29 wherein said multimeric hybrid gene is selected from F_{PIV-3} - F_{RSV}, F_{RSV} - HN_{PIV-3} and F_{PIV-3} - G_{RSV} hybrid genes.

31. The process of claim 30 wherein said multimeric hybrid gene is contained in an expression vector comprising plasmid pAC QR7, pD2 RF-HN or pD2 F-G.

32. The process of claim 28 wherein said cellular expression system is provided by bacterial cells,

mammalian cells, insect cells, yeast cells or fungal cells.

33. The process of claim 32 including separating a chimeric protein from a culture of said cellular expression system and purifying the separated chimeric protein.

34. A live vector for antigen delivery containing the gene of claim 1.

35. The live vector of claim 34 which is a viral vector.

36. The live vector of claim 35 wherein said viral vector is selected from poxviral, adenoviral and retroviral viral vectors.

37. The live vector of claim 34 which is a bacterial vector.

38. The live vector of claim 37 wherein said bacterial vector is selected from salmonella and mycobacteria.

39. A vaccine against diseases caused by multiple pathogenic infections, comprising a chimeric protein comprising an antigen region of a protein from a first pathogen linked to an antigenic region of a protein from a second pathogen, and a physiologically-acceptable carrier therefor.

40. The vaccine of claim 39, wherein said first and second pathogens are selected from bacterial and viral pathogens.

41. The vaccine of claim 39, which also contains at least one other immunogenic and/or immunostimulating molecule.

42. The vaccine of claim 40 wherein both said first and second pathogens are viral pathogens.

43. The vaccine of claim 39, wherein said first and second pathogens are selected from those causing upper and lower respiratory tract diseases.

44. The vaccine of claim 39, wherein said first pathogen is parainfluenza virus (PIV) and said second pathogen is respiratory syncytial virus (RSV).

45. The vaccine of claim 39 against infection by both parainfluenza virus (PIV) and respiratory syncytial virus (RSV), comprising a recombinant multimeric protein containing at least one segment consisting of a PIV protein or an immunogenic epitope-containing fragment thereof linked to at least one segment consisting of a RSV protein or an immunogenic epitope-containing fragment thereof, and a carrier therefor.

46. The vaccine of claim 45 wherein said recombinant multimeric protein is a recombinant chimeric protein containing a segment consisting of a PIV-3 F or HN protein or an immunogenic epitope-containing fragment thereof linked to a segment consisting of an RSV G or F protein or an immunogenic epitope-containing fragment thereof.

47. The vaccine of claim 46 containing at least one additional protein of PIV or RSV or chimeric protein thereof.

48. The vaccine of claim 39 wherein said carrier comprises an adjuvant.

49. The vaccine of claim 39 wherein said carrier is an ISCOM, a liposome or a microparticle.

50. The vaccine of claim 46 formulated to be administered in an injectable form, intranasally or orally.

51. The vaccine of claim 39 further comprising means for delivering said multimeric protein specifically to cells of the immune system.

52. The vaccine of claim 51 wherein said delivery means comprises a toxin molecule or an antibody.

53. A vaccine against diseases caused by multiple pathogenic infection, comprising a live vector as claimed in claim 34, and a physiologically-acceptable carrier therefor.

54. A method of immunizing a host against diseases caused by multiple pathogenic infections, which comprises

administering to a host an effective amount of a vaccine as claimed in claim 28 or 53.

55. The method of claim 54 wherein said vaccine is against diseases caused by parainfluenza virus (PIV) and respiratory syncytial virus (RSV).

56. The method of claim 55 wherein said host is selected from infants, young children, pregnant women, women of child-bearing age and susceptible persons.

57. A diagnostic reagent for detecting infection by a plurality of different pathogens in a host, comprising the chimeric protein claimed in claim 18.

58. A method of detecting infection by a plurality of different pathogens in a host, which comprises using said chimeric protein claimed in claim 18.

Abstract of the Disclosure

Multimeric hybrid genes encoding the corresponding chimeric protein comprise a gene sequence coding for an antigenic region of a protein from a first pathogen linked to a gene sequence coding for an antigenic region of a protein from a second pathogen. The pathogens particularly are parainfluenza virus (PIV) and respiratory syncytial virus (RSV). A single recombinant immunogen is capable of protecting infants and similar susceptible individuals against diseases caused by both PIV and RSV.

FIG.1A. NUCLEOTIDE SEQUENCE OF THE PIV-3 F GENE (PCR-AMPLIFIED)

```
AAGTCAATACCAACAACTATTAGCAGTCATACGTGCAAGAACAAGAAAGAAGAGATTCAA
TTCAGTTATGGTTGTTGATAATCGTCAGTATGCACGTTCTTGTTCTTTCTTCTCTAAGTT
          10        20        30        40        50        60

AAAGCTAAATAAGAGAAATCAAAACAAAAGGTATAGAACACCCGAACAACAAAATCAAAA
TTTCGATTTATTCTCTTTAGTTTTGTTTTCCATATCTTGTGGGCTTGTTGTTTTAGTTTT
          70        80        90        100       110       120

CATCCAATCCATTTTAAACAAAAATTCCAAAAGAGACCGGCAACACAACAAGCACCAAAC
GTAGGTTAGGTAAAATTTGTTTTTAAGGTTTTCTCTGGCCGTTGTGTTGTTCGTGGTTTG
          130       140       150       160       170       180
          <-------- SP --------
     MET PRO THR [LEU] ILE LEU LEU ILE ILE THR THR MET ILE MET ALA [SER] SER CYS GLN
ACAATGCCAACTTTAATACTGCTAATTATTACAACAATGATTATGGCATCTTCCTGCCAA
TGTTACGGTTGAAATTATGACGATTAATAATGTTGTTACTAATACCGTAGAAGGACGGTT
          190       200       210       220       230       240

 ILE ASP ILE THR LYS LEU GLN HIS VAL GLY VAL LEU VAL ASN SER PRO LYS GLY MET LYS
ATACATATCACAAAACTACAGCATGTAGGTGTATTGGTCAACAGTCCCAAAGGGATGAAG
TATGTATAGTGTTTTGATGTCGTACATCCACATAACCAGTTGTCAGGGTTTCCCTACTTC
          250       260       270       280       290       300

 ILE SER GLN ASN PHE GLU THR ARG TYR LEU ILE LEU SER LEU ILE PRO LYS ILE GLU ASP
ATATCACAAAACTTCGAAACAAGATATCTAATTTTGAGCCTCATACCAAAAATAGAAGAC
TATAGTGTTTTGAAGCTTTGTTCTATAGATTAAAACTCGGAGTATGGTTTTTATCTTCTG
          310       320       330       340       350       360

 SER ASN SER CYS GLY ASP GLN GLN ILE LYS GLN TYR LYS ARG LEU LEU ASP ARG LEU ILE
TCTAACTCTTGTGGTGACCAACAGATCAAACAATACAAGAGGTTATTGGATAGACTGATC
AGATTGAGAACACCACTGGTTGTCTAGTTTGTTATGTTCTCCAATAACCTATCTGACTAG
          370       380       390       400       410       420
```

FIG. 1B.

```
ILE PRO LEU TYR ASP GLY LEU ARG LEU GLN LYS ASP VAL ILE VAL THR ASN GLN GLU SER
ATCCCTCTATATGATGGATTAAGATTACAGAAAGATGTGATAGTAACCAATCAAGAATCC
TAGGGAGATATACTACCTAATTCTAATGTCTTTCTACACTATCATTGGTTAGTTCTTAGG
      430       440       450       460       470       480

                                  F2-F1 CLEAVAGE SITE
ASN GLU ASN THR ASP PRO ARG THR ARG ARG | SER PHE GLY GLY VAL ILE GLY THR ILE ALA
AATGAAAACACTGATCCCAGAACAAGACGATCCTTTGGAGGGGTAATTGGAACCATTGCT
TTACTTTTGTGACTAGGGTCTTGTTCTGCTAGGAAACCTCCCCATTAACCTTGGTAACGA
      490       500       510       520       530       540

LEU GLY VAL ALA THR SER ALA GLN ILE THR ALA ALA VAL ALA LEU VAL GLU ALA LYS GLN
CTGGGAGTAGCAACCTCAGCACAAATTACAGCGGCAGTTGCTCTGGTTGAAGCCAAGCAG
GACCCTCATCGTTGGAGTCGTGTTTAATGTCGCCGTCAACGAGACCAACTTCGGTTCGTC
      550       560       570       580       590       600

ALA LYS SER ASP ILE GLU LYS LEU LYS GLU ALA ILE ARG ASP THR ASN LYS ALA VAL GLN
GCAAAATCACACATCGAAAAACTCAAAGAAGCAATCAGGGACACAAACAAAGCAGTGCAG
CGTTTTAGTGTGTAGCTTTTTGAGTTTCTTCGTTAGTCCCTGTGTTTGTTTCGTCACGTC
      610       620       630       640       650       660

SER VAL GLN SER SER ILE GLY ASN LEU ILE VAL ALA ILE LYS SER VAL GLN ASP TYR VAL
TCAGTTCAGAGCTCTATAGGAAATTTAATAGTAGCAATTAAATCAGTCCAAGATTATGTC
AGTCAAGTCTCGAGATATCCTTTAAATTATCATCGTTAATTTAGTCAGGTTCTAATACAG
      670       680       690       700       710       720

ASN ASN GLU ILE VAL PRO SER ILE ALA ARG LEU GLY CYS GLU ALA ALA GLY LEU GLN LEU
AACAACGAAATCGTGCCATCGATTGCTAGACTAGGTTGTGAAGCAGCAGGACTTCAATTA
TTGTTGCTTTAGCACGGTAGCTAACGATCTGATCCAACACTTCGTCGTCCTGAAGTTAAT
      730       740       750       760       770       780

GLY ILE ALA LEU THR GLN HIS TYR SER GLU LEU THR ASN ILE PHE GLY ASP ASN ILE GLY
GGAATTGCATTAACACAGCATTACTCAGAATTAACAAACATATTTGGTGATAACATAGGA
CCTTAACGTAATTGTGTCGTAATGAGTCTTAATTGTTTGTATAAACCACTATTGTATCCT
      790       800       810       820       830       840
```

FIG.1C.

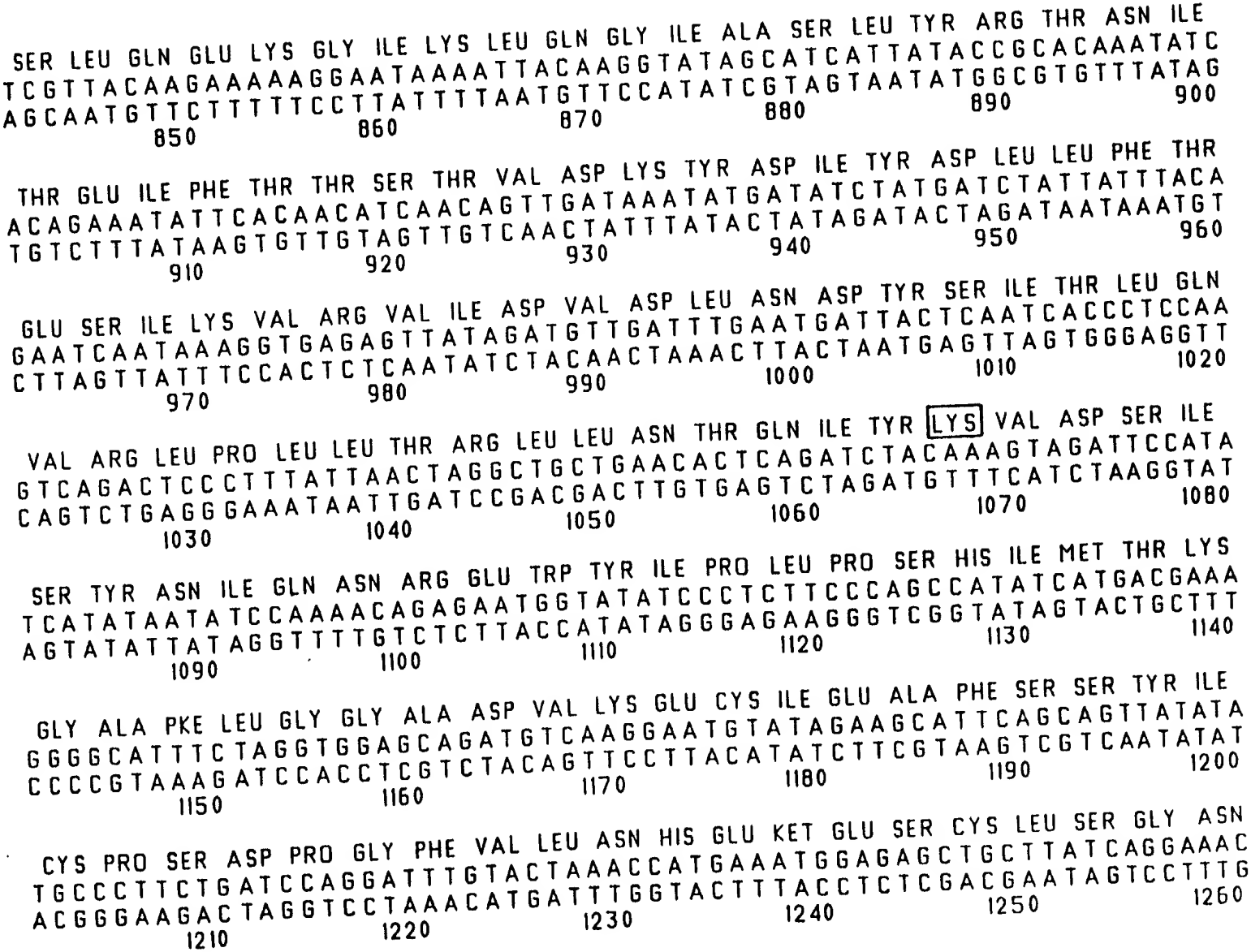


FIG.1D.

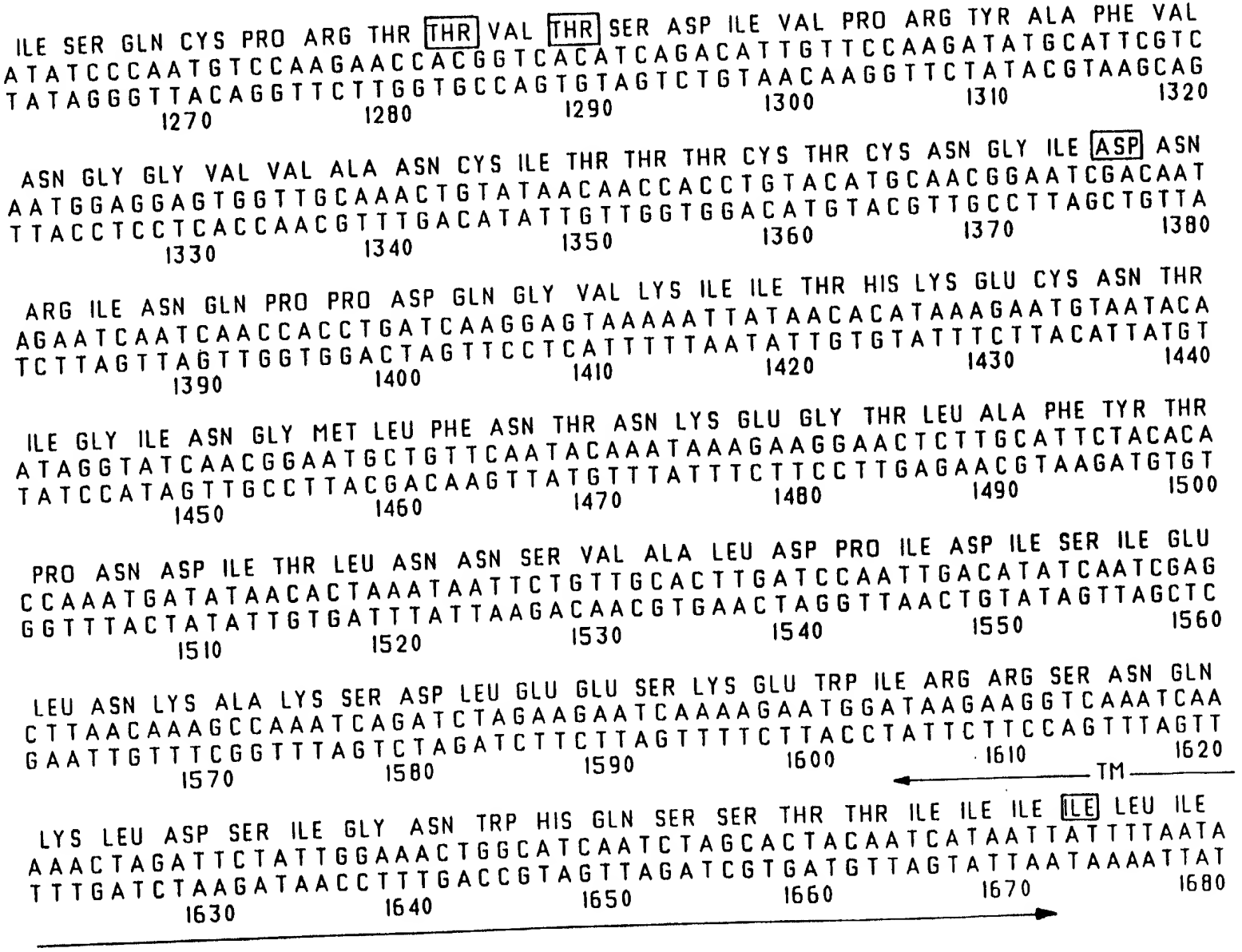

ILE SER GLN CYS PRO ARG THR THR VAL THR SER ASP ILE VAL PRO ARG TYR ALA PHE VAL
ATATCCCAATGTCCAAGAACCACGGTCACATCAGACATTGTTCCAAGATATGCATTCGTC
TATAGGGTTACAGGTTCTTGGTGCCAGTGTAGTCTGTAACAAGGTTCTATACGTAAGCAG
1270 1280 1290 1300 1310 1320
ASN GLY GLY VAL VAL ALA ASN CYS ILE THR THR THR CYS THR CYS ASN GLY ILE ASP ASN
AATGGAGGAGTGGTTGCAAACTGTATAACAACCACCTGTACATGCAACGGAATCGACAAT
TTACCTCCTCACCAACGTTTGACATATTGTTGGTGGACATGTACGTTGCCTTAGCTGTTA
1330 1340 1350 1360 1370 1380
ARG ILE ASN GLN PRO PRO ASP GLN GLY VAL LYS ILE ILE THR HIS LYS GLU CYS ASN THR
AGAATCAATCAACCACCTGATCAAGGAGTAAAAATTATAACACATAAAGAATGTAATACA
TCTTAGTTAGTTGGTGGACTAGTTCCTCATTTTTAATATTGTGTATTTCTTACATTATGT
1390 1400 1410 1420 1430 1440
ILE GLY ILE ASN GLY MET LEU PHE ASN THR ASN LYS GLU GLY THR LEU ALA PHE TYR THR
ATAGGTATCAACGGAATGCTGTTCAATACAAATAAAGAAGGAACTCTTGCATTCTACACA
TATCCATAGTTGCCTTACGACAAGTTATGTTTATTTCTTCCTTGAGAACGTAAGATGTGT
1450 1460 1470 1480 1490 1500
PRO ASN ASP ILE THR LEU ASN ASN SER VAL ALA LEU ASP PRO ILE ASP ILE SER ILE GLU
CCAAATGATATAACACTAAATAATTCTGTTGCACTTGATCCAATTGACATATCAATCGAG
GGTTTACTATATTGTGATTTATTAAGACAACGTGAACTAGGTTAACTGTATAGTTAGCTC
1510 1520 1530 1540 1550 1560
LEU ASN LYS ALA LYS SER ASP LEU GLU GLU SER LYS GLU TRP ILE ARG ARG SER ASN GLN
CTTAACAAAGCCAAATCAGATCTAGAAGAATCAAAAGAATGGATAAGAAGGTCAAATCAA
GAATTGTTTCGGTTTAGTCTAGATCTTCTTAGTTTTCTTACCTATTCTTCCAGTTTAGTT
1570 1580 1590 1600 1610 1620
TM
LYS LEU ASP SER ILE GLY ASN TRP HIS GLN SER SER THR THR ILE ILE ILE ILE LEU ILE
AAACTAGATTCTATTGGAAACTGGCATCAATCTAGCACTACAATCATAATTATTTTAATA
TTTGATCTAAGATAACCTTTGACCGTAGTTAGATCGTGATGTTAGTATTAATAAAATTAT
1630 1640 1650 1660 1670 1680

FIG.1E

```
MET ILE ILE ILE LEU PHE ILE ILE ASN VAL THR ILE ILE [THR] ILE ALA [ILE] LYS TYR TYR
ATGATCATTATATTGTTTATAATTAATGTAACGATAATTACAATTGCAATTAAGTATTAC
TACTAGTAATATAACAAATATTAATTACATTGCTATTAATGTTAACGTTAATTCATAATG
     1690      1700      1710      1720      1730      1740

ARG ILE GLN LYS ARG ASN ARG VAL ASP GLN ASN ASP LYS PRO TYR VAL LEU THR ASN LYS
AGAATTCAAAAGAGAAATCGAGTGGATCAAAATGACAAGCCATATGTACTAACAAACAAA
TCTTAAGTTTTCTCTTTAGCTCACCTAGTTTTACTGTTCGGTATACATGATTGTTTGTTT
     1750      1760      1770      1780      1790      1800

TGACATATCTATAGATCATTAGATATTAAAATTATAAAAAACTT
ACTGTATAGATATCTAGTAATCTATAATTTTAATATTTTTTGAA
     1810      1820      1830      1840
```

NUCLEOTIDE SEQUENCE OF THE PIV-3 F GENE. THE cDNA SEQUENCE IS SHOWN IN THE PLUS (mRNA) STRAND SENSE IN THE 5' TO 3' DIRECTION. THE SIGNAL PEPTIDE (SP) AND THE TRANSMEMBRANE (TM) ANCHOR DOMAIN ARE UNDERLINED. THE PREDICTED F2-F1 CLEAVAGE SITE IS INDICATED BY THE ARROW (↓). AMINO ACIDS DIFFERING FROM THE PUBLISHED PRIMARY SEQUENCE OF THE PROTEIN ENCODED BY THE PIV-3 F GENE ARE BOXED.

RESTRICTION MAP OF THE PIV-3 F GENE

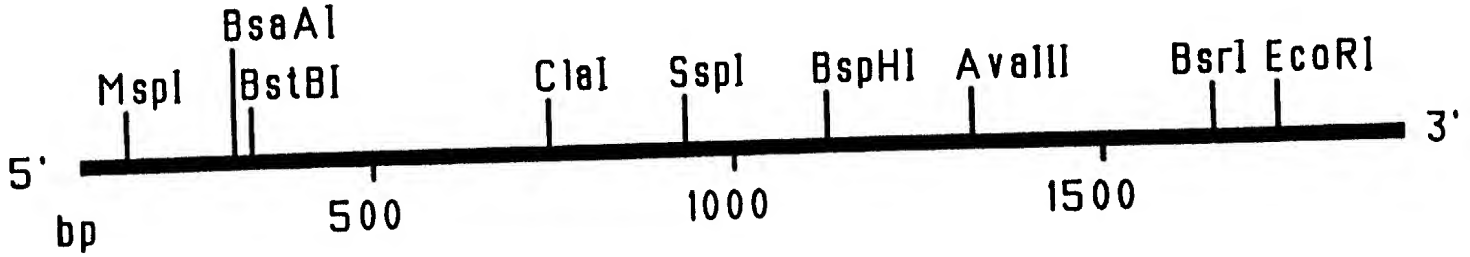
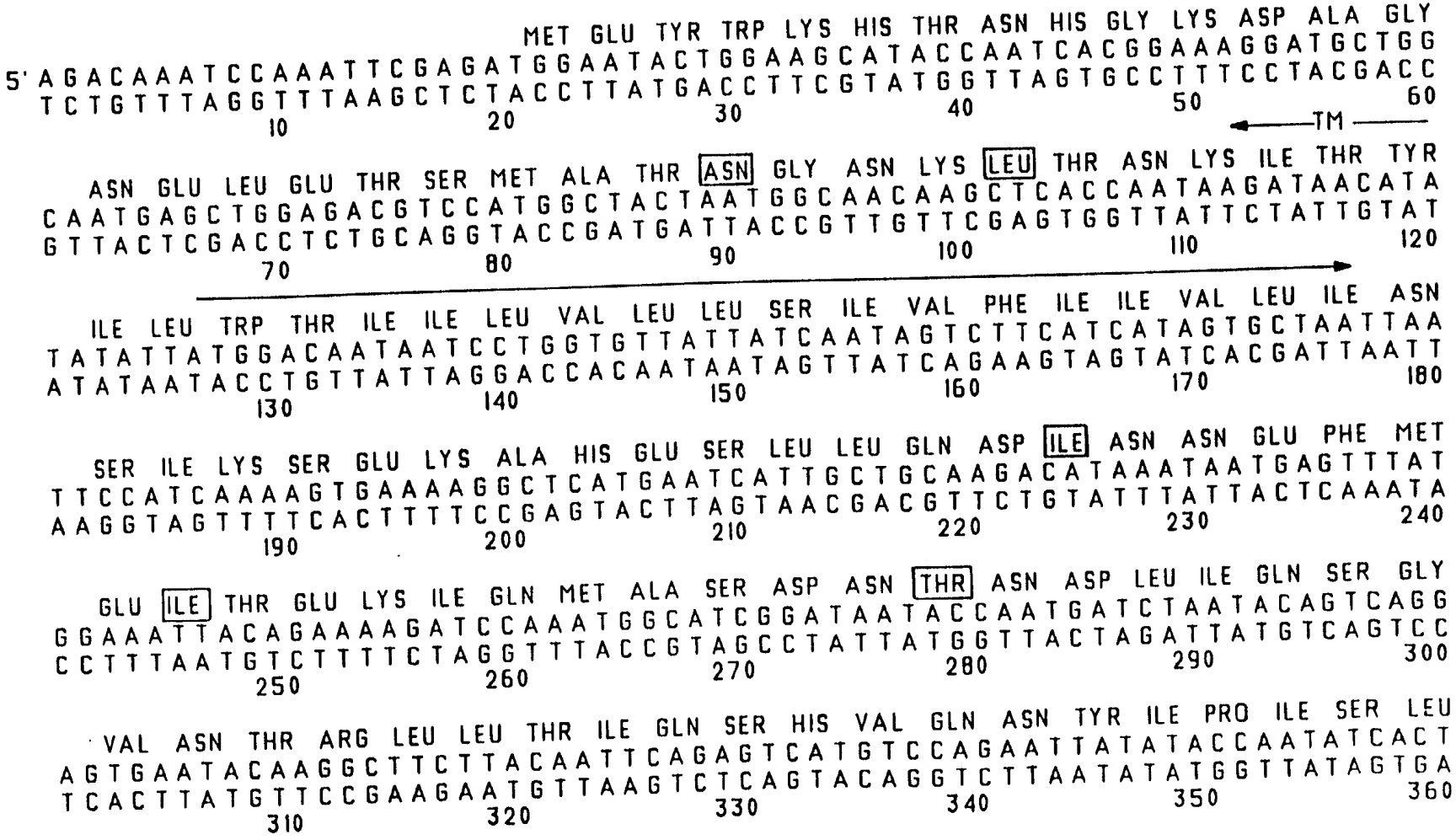


FIG.2.

FIG.3A. NUCLEOTIDE SEQUENCE OF THE PIV-3 HN GENE.



```
THR GLN GLN MET SER ASP LEU ARG LYS PHE ILE SER GLU ILE THR ILE ARG ASN ASP ASN
GACACAACAGATGTCAGATCTTAGGAAATTCATTAGTGAAATTACAATTAGAAATGATAA
CTGTGTTGTCTACAGTCTAGAATCCTTTAAGTAATCACTTTAATGTTAATCTTTACTATT
        370       380       390       400       410       420

[GLN] GLU VAL [LEU] PRO GLN ARG ILE THR HIS ASP [VAL] GLY ILE LYS PRO LEU ASN PRO ASP
TCAAGAAGTGCTGCCACAAAGAATAACACATGATGTGGGTATAAAACCTTTAAATCCAGA
AGTTCTTCACGACGGTGTTTCTTATTGTGTACTACACCCATATTTTGGAAATTTAGGTCT
        430       440       450       460       470       480

ASP PHE TRP ARG CYS THR SER GLY LEU PRO SER LEU MET LYS THR PRO LYS ILE ARG LEU
TGATTTTTGGAGATGCACGTCTGGTCTTCCATCTTTAATGAAAACTCCAAAAATAAGGTT
ACTAAAAACCTCTACGTGCAGACCAGAAGGTAGAAATTACTTTTGAGGTTTTTATTCCAA
        490       500       510       520       530       540

MET PRO GLY PRO GLY LEU LEU ALA MET PRO THR THR VAL ASP GLY CYS [ILE] ARG THR PRO
AATGCCAGGGCCGGGATTATTAGCTATGCCAACGACTGTTGATGGCTGTATCAGAACTCC
TTACGGTCCCGGCCCTAATAATCGATACGGTTGCTGACAACTACCGACATAGTCTTGAGG
        550       560       570       580       590       600

SER LEU VAL ILE ASN ASP LEU ILE TYR ALA TYR THR SER ASN LEU ILE THR ARG GLY CYS
GTCCTTAGTTATAAATGATCTGATTTATGCTTATACCTCAAATCTAATTACTCGAGGTTG
CAGGAATCAATATTTACTAGACTAAATACGAATATGGAGTTTAGATTAATGAGCTCCAAC
        610       620       630       640       650       660

GLN ASP ILE GLY LYS SER TYR GLN VAL LEU GLN ILE GLY ILE ILE THR VAL ASN SER ASP
TCAGGATATAGGAAAATCATATCAAGTCTTACAGATAGGGATAATAACTGTAAACTCAGA
AGTCCTATATCCTTTTAGTATAGTTCAGAATGTCTATCCCTATTATTGACATTTGAGTCT
        670       680       690       700       710       720

LEU VAL PRO ASP LEU ASN PRO ARG ILE SER HIS THR PHE ASN ILE ASN ASP ASN ARG LYS
CTTGGTACCTGACTTAAATCCCAGGATCTCTCATACTTTTAACATAAATGACAATAGGAA
GAACCATGGACTGAATTTAGGGTCCTAGAGAGTATGAAAATTGTATTTACTGTTATCCTT
        730       740       750       760       770       780
```

FIG.3B.

```
   SRE CYS SER LEU ALA LEU LEU ASN THR ASP VAL TYR GLN LEU CYS SER THR PRO LYS VAL
G T C A T G T T C T C T A G C A C T C C T A A A T A C A G A T G T A T A T C A A C T G T G T T C A A C T C C C A A A G T
C A G T A C A A G A G A T C G T G A G G A T T T A T G T C T A C A T A T A G T T G A C A C A A G T T G A G G G T T T C A
          790                800                810                820                830                840

   ASP GLU ARG SER ASP TYR ALA SER SER GLY ILE GLU ASP ILE VAL LEU ASP ILE VAL ASN
T G A T G A A A G A T C A G A T T A T G C A T C A T C A G G C A T A G A A G A T A T T G T A C T T G A T A T T G T C A A
A C T A C T T T C T A G T C T A A T A C G T A G T A G T C C G T A T C T T C T A T A A C A T G A A C T A T A A C A G T T
          850                860                870                880                890                900

   [TYR] ASP GLY SER ILE SER THR THR ARG PHE LYS ASN ASN ASN ILE SER PHE ASP GLN PRO
T T A T G A T G G C T C A A T C T C A A C A A C A A G A T T T A A G A A T A A T A A C A T A A G C T T T G A T C A A C C
A A T A C T A C C G A G T T A G A G T T G T T G T T C T A A A T T C T T A T T A T T G T A T T C G A A A C T A G T T G G
          910                920                930                940                950                960

   TYR ALA ALA LEU TYR PRO SER VAL GLY PRO GLY ILE TYR TYR LYS GLY LYS ILE ILE PHE
T T A T G C T G C A C T A T A C C C A T C T G T T G G A C C A G G G A T A T A C T A C A A A G G C A A A A T A A T A T T
A A T A C G A C G T G A T A T G G G T A G A C A A C C T G G T C C C T A T A T G A T G T T T C C G T T T T A T T A T A A
          970                980                990               1000               1010               1020

   LEU GLY TYR GLY GLY LEU GLU HIS PRO ILE ASN GLU ASN [VAL] ILE CYS ASN THR THR GLY
T C T C G G G T A T G G A G G T C T T G A A C A T C C A A T A A A T G A G A A T G T A A T C T G C A A C A C A A C T G G
A G A G C C C A T A C C T C C A G A A C T T G T A G G T T A T T T A C T C T T A C A T T A G A C G T T G T G T T G A C C
          1030               1040               1050               1060               1070               1080

   CYS PRO GLY LYS THR GLN ARG ASP CYS ASN GLN ALA SER HIS SER PRO TRP PHE SER ASP
G T G T C C C G G G A A A A C A C A G A G A G A C T G C A A T C A G G C A T C T C A T A G T C C A T G G T T T T C A G A
C A C A G G G C C C T T T T G T G T C T C T C T G A C G T T A G T C C G T A G A G T A T C A G G T A C C A A A A G T C T
          1090               1100               1110               1120               1130               1140

   ARG ARG MET VAL ASN SER ILE ILE VAL VAL ASP LYS GLY LEU ASN SER ILE PRO LYS LEU
T A G G A G G A T G G T C A A C T C T A T C A T T G T T G T T G A C A A A G G C T T A A A C T C A A T T C C A A A A T T
A T C C T C C T A C C A G T T G A G A T A G T A A C A A C A A C T G T T T C C G A A T T T G A G T T A A G G T T T T A A
          1150               1160               1170               1180               1190               1200
```

FIG.3C.

```
   LYS VAL TRP THR ILE SER MET ARG GLN ASN TYR TRP GLY SER GLU GLY ARG LEU LEU LEU
GAAGGTATGGACGATATCTATGAGACAGAATTACTGGGGGTCAGAAGGAAGGTTACTTCT
CTTCCATACCTGCTATAGATACTCTGTCTTAATGACCCCCAGTCTTCCTTCCAATGAAGA
         1210      1220      1230      1240      1250      1260

   LEU GLY ASN LYS ILE TYR ILE TYR THR ARG SER THR SER TRP HIS SER LYS LEU GLN LEU
ACTAGGTAACAAGATCTATATATATACAAGATCCACAAGTTGGCATAGCAAGTTACAATT
TGATCCATTGTTCTAGATATATATATGTTCTAGGTGTTCAACCGTATCGTTCAATGTTAA
         1270      1280      1290      1300      1310      1320

   GLY ILE ILE ASP ILE THR ASP TYR SER ASP ILE ARG ILE LYS TRP THR TRP HIS ASN VAL
AGGAATAATTGATATTACTGATTACAGTGATATAAGGATAAAATGGACATGGCATAATGT
TCCTTATTAACTATAATGACTAATGTCACTATATTCCTATTTTACCTGTACCGTATTACA
         1330      1340      1350      1360      1370      1380

   LEU SER ARG PRO GLY ASN ASN GLU CYS PRO TRP GLY HIS SER CYS PRO ASP GLY CYS ILE
GCTATCAAGACCAGGAAACAATGAATGTCCATGGGGACATTCATGTCCAGATGGATGTAT
CGATAGTTCTGGTCCTTTGTTACTTACAGGTACCCCTGTAAGTACAGGTCTACCTACATA
         1390      1400      1410      1420      1430      1440

   THR GLY VAL TYR THR ASP ALA TYR PRO LEU ASN PRO THR GLY SER ILE VAL SER SER VAL
AACAGGAGTATATACTGATGCATATCCACTCAATCCCACAGGGAGCATTGTGTCATCTGT
TTGTCCTCATATATGACTACGTATAGGTGAGTTAGGGTGTCCCTCGTAACACAGTAGACA
         1450      1460      1470      1480      1490      1500

   ILE LEU ASP SER GLN LYS SER ARG VAL ASN PRO VAL ILE THR TYR SER THR [ALA] THR GLU
CATATTAGATTCACAAAAATCGAGAGTGAACCCAGTCATAACTTACTCAACAGCAACCGA
GTATAATCTAAGTGTTTTTAGCTCTCACTTGGGTCAGTATTGAATGAGTTGTCGTTGGCT
         1510      1520      1530      1540      1550      1560

   ARG VAL ASN GLU LEU ALA ILE [ARG] ASN ARG THR LEU SER ALA GLY TYR THR THR THR SER
AAGAGTAAACGAGCTGGCCATCCGAAACAGAACACTCTCAGCTGGATATACAACAACAAG
TTCTCATTTGCTCGACCGGTAGGCTTTGTCTTGTGAGAGTCGACCTATATGTTGTTGTTC
         1570      1580      1590      1600      1610      1620
```

FIG.3D.

```
   CYS  ILE  THR  HIS  TYR  ASN  LYS  GLY  TYR  CYS  PHE  HIS  ILE  VAL  GLU  ILE  ASN  GLN  LYS  SER
CTGCATCACACACTATAACAAAGGATATTGTTTTCATATAGTAGAAATAAATCAGAAAAG
GACGTAGTGTGTGATATTGTTTCCTATAACAAAAGTATATCATCTTTATTTAGTCTTTTC
              1630          1640          1650          1660          1670          1680

   LEU  ASN  THR  LEU  GLN  PRO  MET  LEU  PHE  LYS  THR  GLU  VAL  PRO  LYS  SER  CYS  SER  ***
CTTAAACACACTTCAACCCATGTTGTTCAAGACAGAGGTTCCAAAAAGCTGCAGTTAATC
GAATTTGTGTGAAGTTGGGTACAACAAGTTCTGTCTCCAAGGTTTTTCGACGTCAATTAG
              1690          1700          1710          1720          1730          1740

ATAATTAACCGCAATATGCATTAACCTATCTATAATACAAGTATATGATAAGTAATCAGC
TATTAATTGGCGTTATACGTAATTGGATAGATATTATGTTCATATACTATTCATTAGTCG
              1750          1760          1770          1780          1790          1800

AATCAGACAATAGACAAAAGGGAAATATAAAAA
TTAGTCTGTTATCTGTTTTCCCTTTATATTTTT
              1810          1820          1830
```

NUCLEOTIDE SEQUENCE OF THE PIV-3 HN GENE. THE cDNA SEQUENCE IS SHOWN IN THE PLUS (mRNA) STRAND SENSE IN THE 5' TO 3' DIRECTION. THE TRANSMEMBRANE (TM) ANCHOR DOMAIN IS UNDERLINED. AMINO ACIDS DIFFERING FROM THE PUBLISHED PRIMARY SEQUENCE OF THE PROTEIN ENCODED BY THE PIV-3 HN GENE ARE BOXED.

FIG.3E.

RESTRICTION MAP OF THE PIV-3 HN GENE

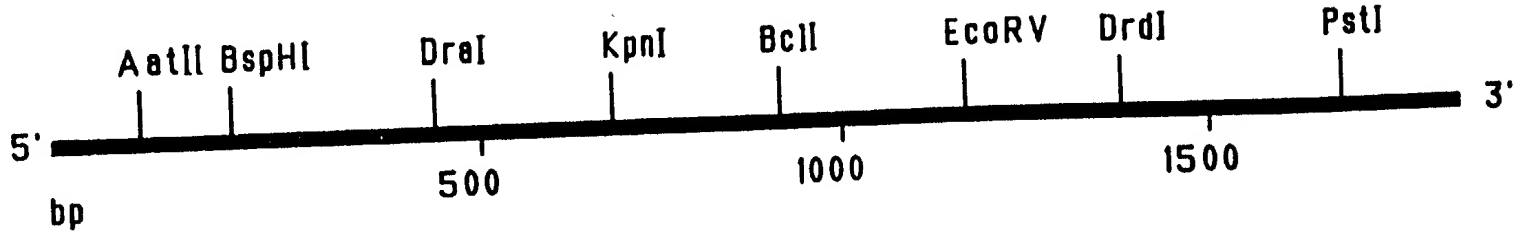
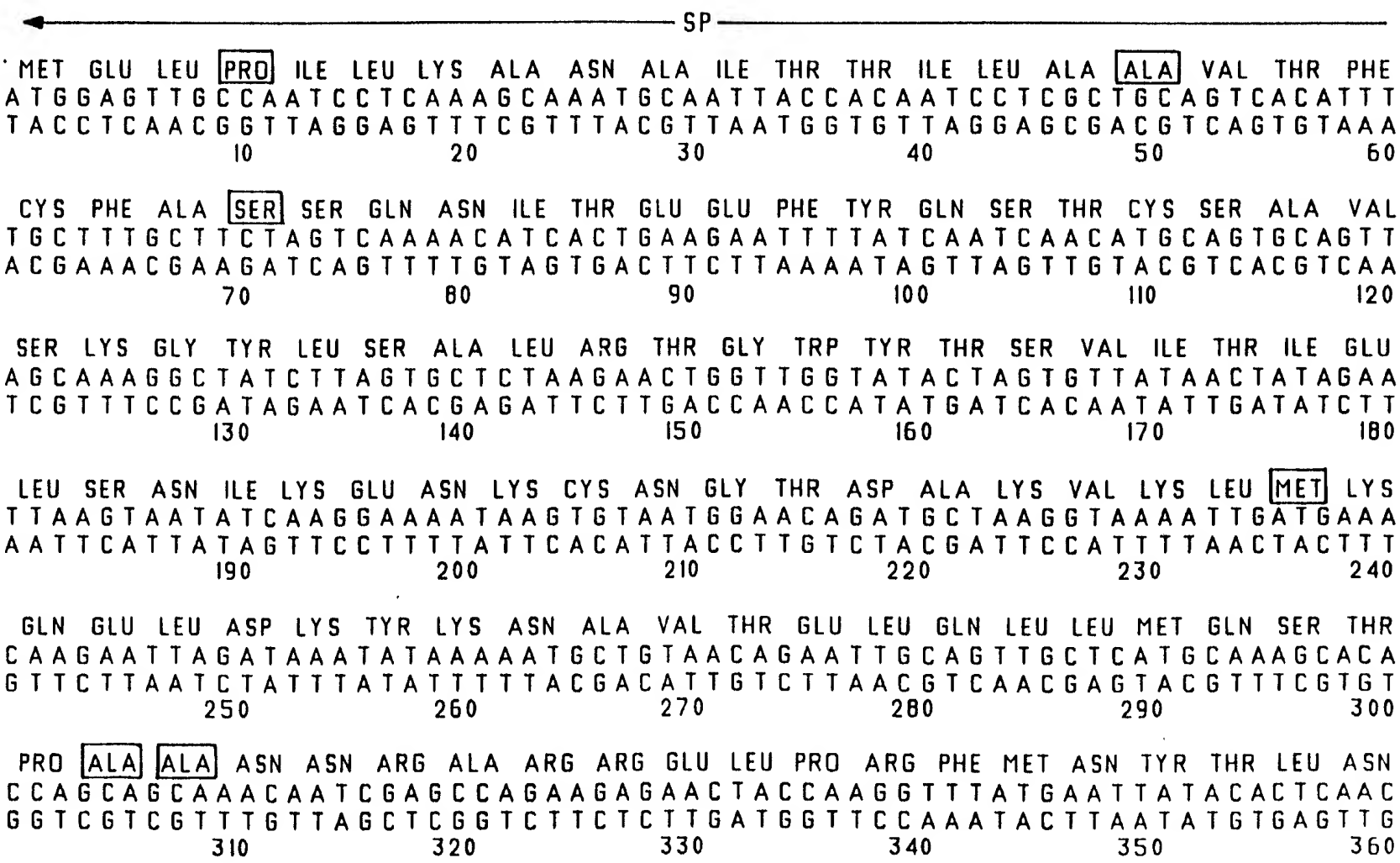


FIG.4.

FIG.5A. NUCLEOTIDE SEQUENCE OF THE RSV F GENE.



F2-F1 CLEAVAGE SITE

ASN [THR] LYS LYS THR ASN VAL THR LEU SER LYS LYS ARG LYS ARG ARG↓PHE LEU GLY PHE
AATACCAAAAAAAACCAATGTAACATTAAGCAAGAAAAGGAAAAGAAGATTTCTTGGTTTT
TTATGGTTTTTTTGGTTACATTGTAATTCGTTCTTTTCCTTTTCTTCTAAAGAACCAAAA
370 380 390 400 410 420

LEU LEU GLY VAL GLY SER ALA ILE ALA SER GLY [ILE] ALA VAL SER LYS VAL LEU HIS LEU
TTGTTAGGTGTTGGATCTGCAATCGCCAGTGGCATTGCTGTATCTAAGGTCCTGCACTTA
AACAATCCACAACCTAGACGTTAGCGGTCACCGTAACGACATAGATTCCAGGACGTGAAT
430 440 450 460 470 480

GLU GLY GLU VAL ASN LYS ILE LYS SER ALA LEU LEU SER THR ASN LYS ALA VAL VAL SER
GAAGGAGAAGTGAACAAGATCAAAAGTGCTCTACTATCCACAAACAAGGCCGTAGTCAGT
CTTCCTCTTCACTTGTTCTAGTTTTCACGAGATGATAGGTGTTTGTTCCGGCATCAGTCA
490 500 510 520 530 540

LEU SER ASN GLY VAL SER VAL LEU THR SER LYS VAL LEU ASP LEU LYS ASN TYR ILE ASP
TTATCAAATGGAGTTAGTGTCTTAACCAGCAAAGTGTTAGACCTCAAAAACTATATAGAT
AATAGTTTACCTCAATCACAGAATTGGTCGTTTCACAATCTGGAGTTTTTGATATATCTA
550 560 570 580 590 600

LYS GLN LEU LEU PRO ILE VAL ASN LYS ARG SER CYS [ARG] ILE SER ASN ILE GLU THR VAL
AAACAATTGTTACCTATTGTGAATAAGCGAAGCTGCAGAATATCAAATATAGAAACTGTG
TTTGTTAACAATGGATAACACTTATTCGCTTCGACGTCTTATAGTTTATATCTTTGACAC
610 620 630 640 650 660

ILE GLU PHE GLN HIS LYS ASN ASN ARG LEU LEU GLU ILE THR ARG GLU PHE SER VAL ASN
ATAGAGTTCCAACACAAGAACAACAGACTACTAGAGATTACCAGGGAATTTAGTGTTAAT
TATCTCAAGGTTGTGTTCTTGTTGTCTGATGATCTCTAATGGTCCCTTAAATCACAATTA
670 680 690 700 710 720

ALA GLY VAL THR THR PRO VAL SER THR TYR MET LEU THR ASN SER GLU LEU LEU SER LEU
GCAGGTGTAACTACACCTGTAAGCACTTACATGTTAACTAATAGTGAATTATTGTCATTA
CGTCCACATTGATGTGGACATTCGTGAATGTACAATTGATTATCACTTAATAACAGTAAT
730 740 750 760 770 780

FIG.5B.

```
ILE ASN ASP MET PRO ILE THR ASN ASP GLN LYS LYS LEU MET SER ASN ASN VAL GLN ILE
ATCAATGATATGCCTATAACAAATGATCAGAAAAAGTTAATGTCCAACAATGTTCAAATA
TAGTTACTATACGGATATTGTTTACTAGTCTTTTTCAATTACAGGTTGTTACAAGTTTAT
          790        800        810        820        830        840

VAL ARG GLN GLN SER TYR SER ILE MET SER ILE ILE LYS GLU GLU VAL LEU ALA TYR VAL
GTTAGACAGCAAAGTTACTCTATCATGTCCATAATAAAAGAGGAAGTCTTAGCATATGTA
CAATCTGTCGTTTCAATGAGATAGTACAGGTATTATTTTCTCCTTCAGAATCGTATACAT
          850        860        870        880        890        900

VAL GLN LEU PRO LEU TYR GLY VAL ILE ASP THR PRO CYS TRP LYS LEU HIS THR SER PRO
GTACAATTACCACTATATGGTGTGATAGATACACCTTGTTGGAAATTACACACATCCCCT
CATGTTAATGGTGATATACCACACTATCTATGTGGAACAACCTTTAATGTGTGTAGGGGA
          910        920        930        940        950        960

LEU CYS THR THR ASN THR LYS GLU GLY SER ASN ILE CYS LEU THR ARG THR ASP ARG GLY
CTATGTACAACCAACACAAAAGAAGGGTCAAACATCTGTTTAACAAGAACTGACAGAGGA
GATACATGTTGGTTGTGTTTTCTTCCCAGTTTGTAGACAAATTGTTCTTGACTGTCTCCT
          970        980        990       1000       1010       1020

TRP TYR CYS ASP ASN ALA GLY SER VAL SER PHE PHE PRO GLN ALA GLU THR CYS LYS VAL
TGGTACTGTGACAATGCAGGATCAGTATCTTTCTTCCCACAAGCTGAAACATGTAAAGTT
ACCATGACACTGTTACGTCCTAGTCATAGAAAGAAGGGTGTTCGACTTTGTACATTTCAA
         1030       1040       1050       1060       1070       1080

GLN SER ASN ARG VAL PHE CYS ASP THR MET ASN SER LEU THR LEU PRO SER GLU VAL ASN
CAATCGAATCGAGTATTTTGTGACACAATGAACAGTTTAACATTACCAAGTGAAGTAAAT
GTTAGCTTAGCTCATAAAACACTGTGTTACTTGTCAAATTGTAATGGTTCACTTCATTTA
         1090       1100       1110       1120       1130       1140

LEU CYS ASN VAL ASP ILE PHE ASN PRO LYS TYR ASP CYS LYS ILE MET THR SER LYS THR
CTCTGCAATGTTGACATATTCAATCCCAAATATGATTGTAAAATTATGACTTCAAAAACA
GAGACGTTACAACTGTATAAGTTAGGGTTTATACTAACATTTTAATACTGAAGTTTTTGT
         1150       1160       1170       1180       1190       1200
```

FIG.5C.

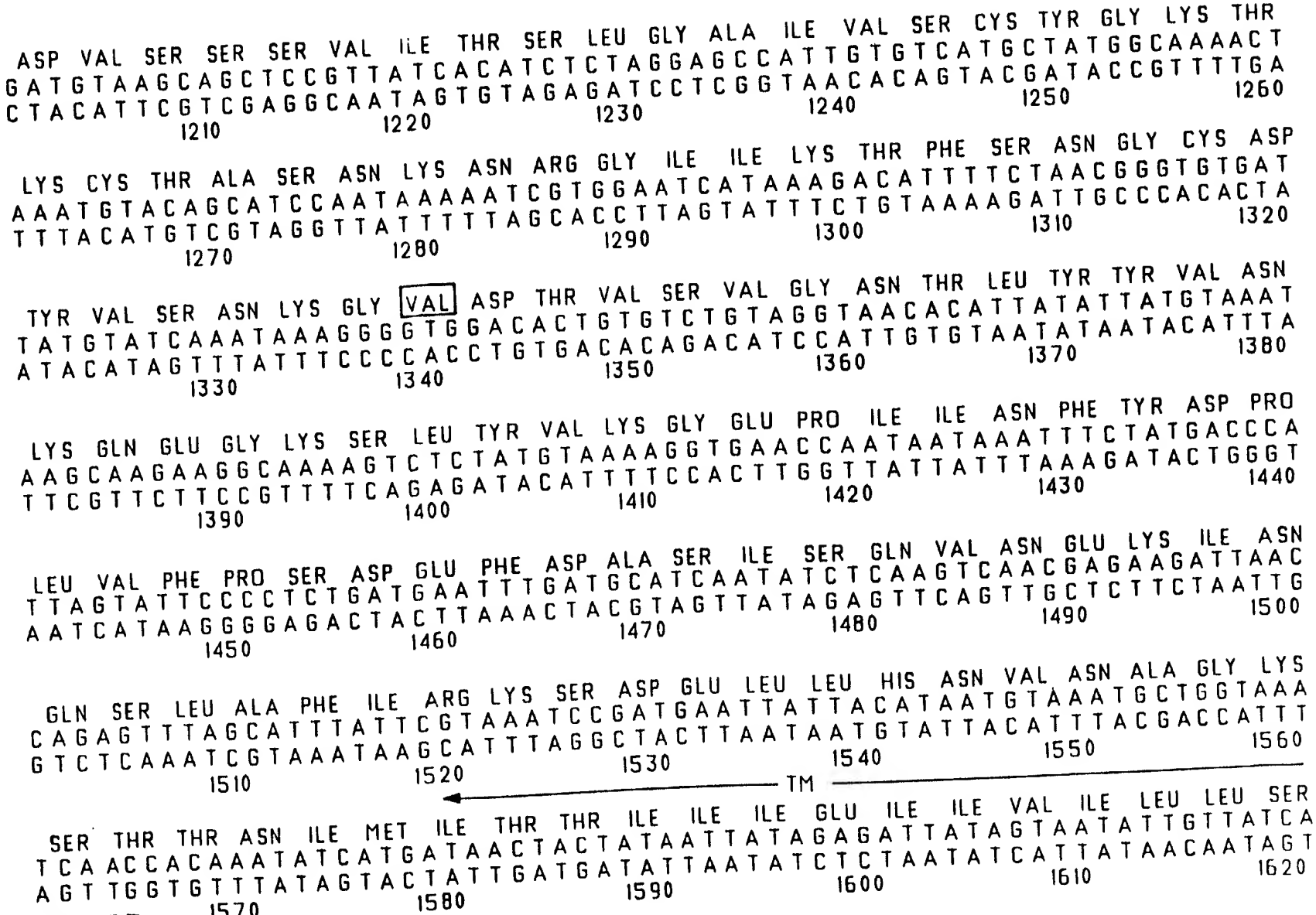
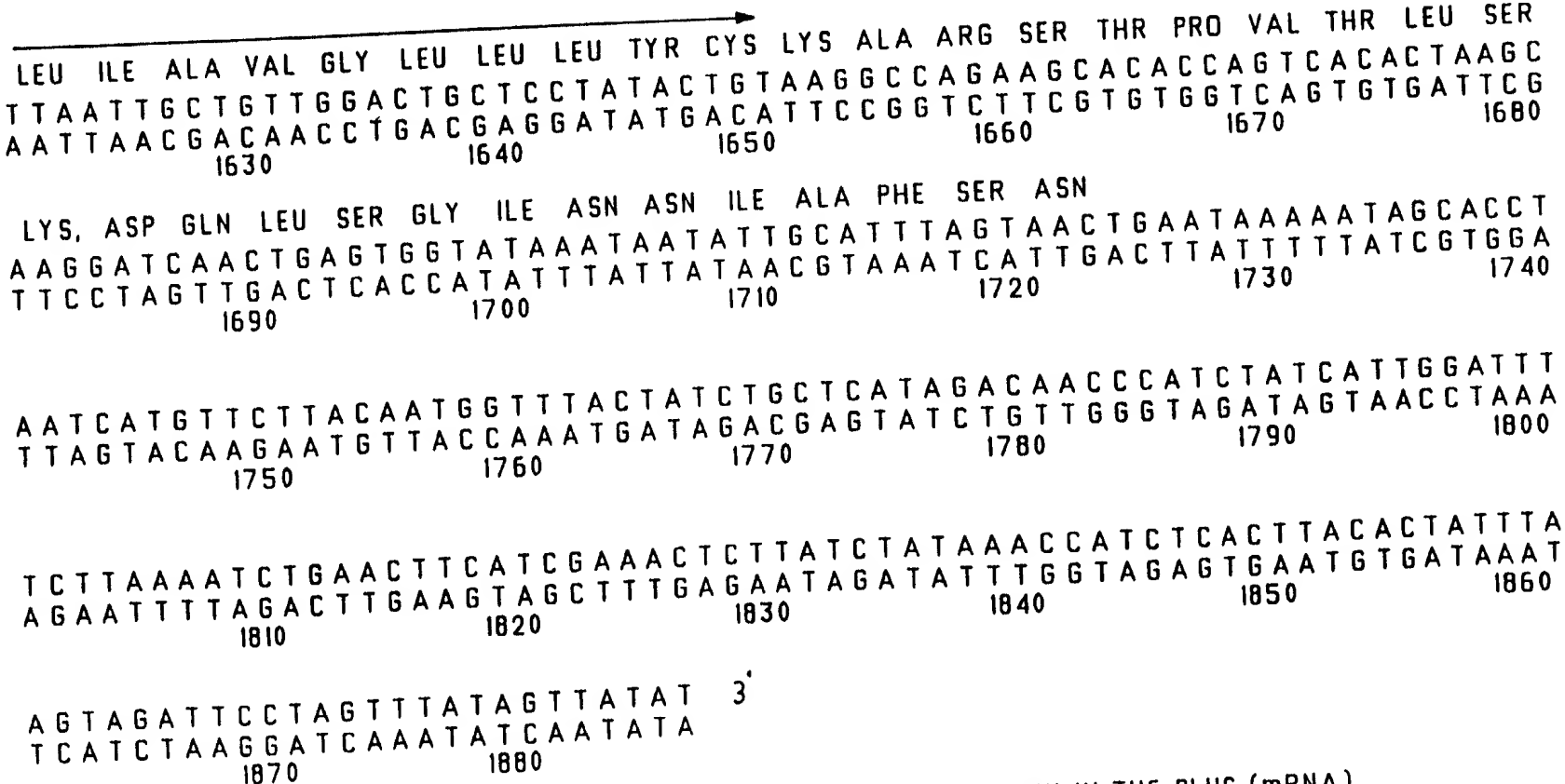


FIG.5D.



NUCLEOTIDE SEQUENCE OF THE RSV F GENE.THE cDNA SEQUENCE IS SHOWN IN THE PLUS (mRNA) STRAND SENSE IN THE 5' TO 3' DIRECTION.THE SIGNAL PEPTIDE (SP) AND THE TRANSMEMBRANE (TM) ANCHOR DOMAIN ARE UNDERLINED.THE PREDICTED F2-F1 CLEAVAGE SITE IS INDICATED BY THE ARROW (↓).AMINO ACIDS DIFFERING FROM THE PUBLISHED PRIMARY SEQUENCE OF THE PROTEIN ENCODED BY THE RSV F GENE ARE BOXED.

FIG.5E.

RESTRICTION MAP OF THE RSV F GENE

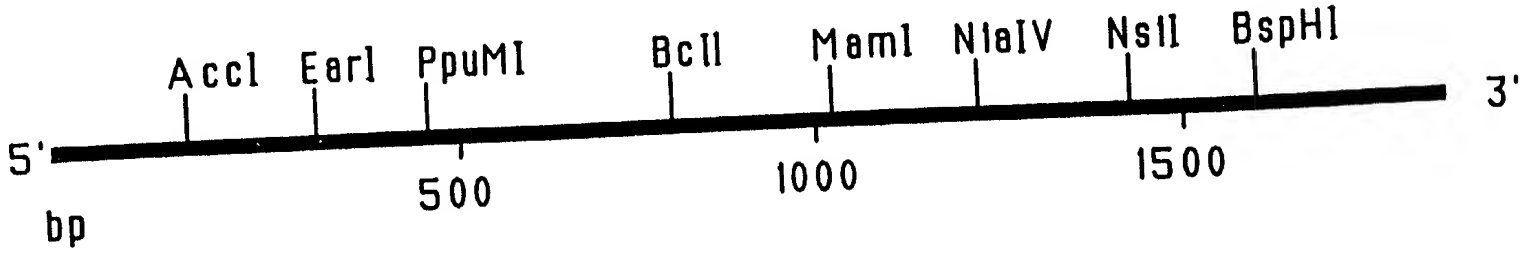


FIG.6.

FIG.7A. NUCLEOTIDE SEQUENCE OF THE RSV G GENE

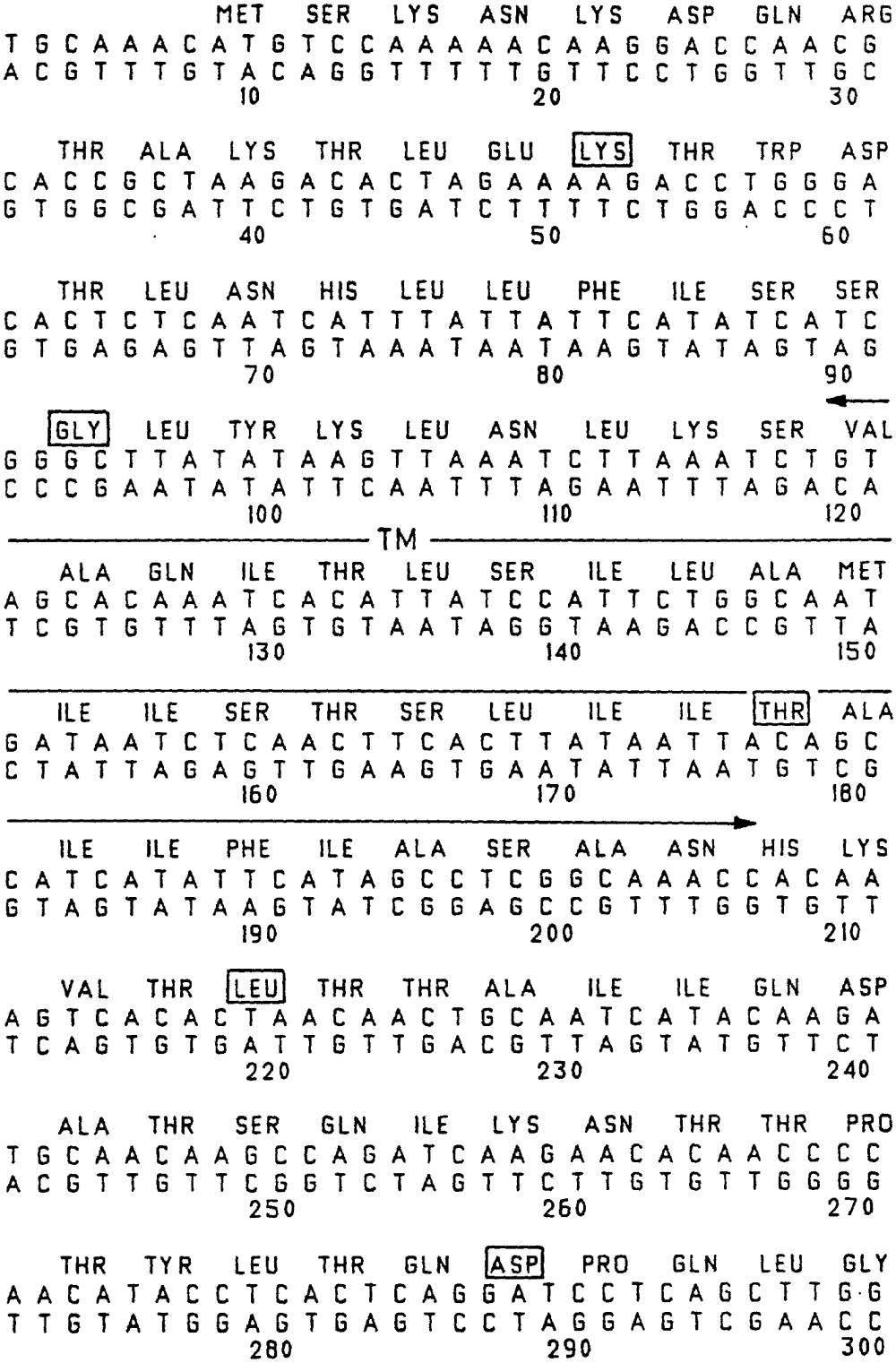


FIG.7B.

```
   ILE  SER  PHE  SER  ASN  LEU  SER  GLU  ILE  THR
A A T C A G C T T C T C C A A T C T G T C T G A A A T T A C
T T A G T C G A A G A G G T T A G A C A G A C T T T A A T G
                  310                 320                 330

   SER  GLN  THR  THR  THR  ILE  LEU  ALA  SER  THR
A T C A C A A A C C A C C A C C A T A C T A G C T T C A A C
T A G T G T T T G G T G G T G G T A T G A T C G A A G T T G
                  340                 350                 360

   THR  PRO  GLY  VAL  LYS  SER  ASN  LEU  GLN  PRO
A A C A C C A G G A G T C A A G T C A A A C C T G C A A C C
T T G T G G T C C T C A G T T C A G T T T G G A C G T T G G
                  370                 380                 390

   THR  THR  VAL  LYS  THR  LYS  ASN  THR  THR  THR
C A C A A C A G T C A A G A C T A A A A A C A C A A C A A C
G T G T T G T C A G T T C T G A T T T T T G T G T T G T T G
                  400                 410                 420

   THR  GLN  THR  GLN  PRO  SER  LYS  PRO  THR  THR
A A C C C A A A C A C A A C C C A G C A A G C C C A C T A C
T T G G G T T T G T G T T G G G T C G T T C G G G T G A T G
                  430                 440                 450

   LYS  GLN  ARG  GLN  ASN  LYS  PRO  PRO  ASN  LYS
A A A A C A A C G C C A A A A C A A A C C A C C A A A C A A
T T T T G T T G C G G T T T T G T T T G G T G G T T T G T T
                  460                 470                 480

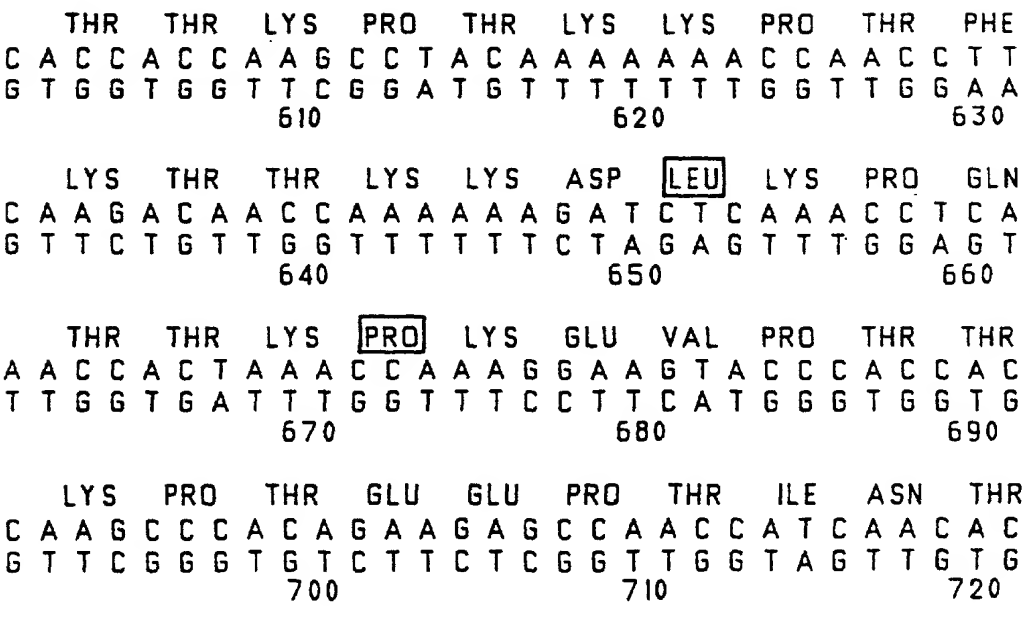
   PRO  ASN  ASN  ASP  PHE  HIS  PHE  GLU  VAL  PHE
A C C C A A T A A T G A T T T T C A C T T C G A A G T G T T
T G G G T T A T T A C T A A A A G T G A A G C T T C A C A A
                  490                 500                 510

   ASN  PHE  VAL  PRO  CYS  SER  ILE  CYS  SER  ASN
T A A C T T T G T A C C C T G C A G C A T A T G C A G C A A
A T T G A A A C A T G G G A C G T C G T A T A C G T C G T T
                  520                 530                 540

   ASN  PRO  THR  CYS  TRP  ALA  ILE  CYS  LYS  ARG
C A A T C C A A C C T G C T G G G C T A T C T G C A A A A G
G T T A G G T T G G A C G A C C C G A T A G A C G T T T T C
                  550                 560                 570

   ILE  PRO  ASN  LYS  LYS  PRO  GLY  LYS  LYS  THR
A A T A C C A A A C A A A A A A C C A G G A A A G A A A A C
T T A T G G T T T G T T T T T T G G T C C T T T C T T T T G
                  580                 590                 600
```

FIG.7C.


THR THR LYS PRO THR LYS LYS PRO THR PHE
C A C C A C C A A G C C T A C A A A A A A A C C A A C C T T
G T G G T G G T T C G G A T G T T T T T T T G G T T G G A A
610 620 630
LYS THR THR LYS LYS ASP LEU LYS PRO GLN
C A A G A C A A C C A A A A A A A G A T C T C A A A C C T C A
G T T C T G T T G G T T T T T T C T A G A G T T T G G A G T
640 650 660
THR THR LYS PRO LYS GLU VAL PRO THR THR
A A C C A C T A A A C C A A A G G A A G T A C C C A C C A C
T T G G T G A T T T G G T T T C C T T C A T G G G T G G T G
670 680 690
LYS PRO THR GLU GLU PRO THR ILE ASN THR
C A A G C C C A C A G A A G A G C C A A C C A T C A A C A C
G T T C G G G T G T C T T C T C G G T T G G T A G T T G T G
700 710 720
THR LYS THR ASN ILE THR THR THR LEU LEU
C A C C A A A A C A A A C A T C A C A A C T A C A C T G C T
G T G G T T T T G T T T G T A G T G T T G A T G T G A C G A
730 740 750
THR ASN ASN THR THR GLY ASN PRO LYS LEU
C A C C A A C A A C A C C A C A G G A A A T C C A A A A C T
G T G G T T G T T G T G G T G T C C T T T A G G T T T T G A
760 770 780
THR SER GLN MET GLU THR PHE HIS SER THR
C A C A A G T C A A A T G G A A A C C T T C C A C T C A A C
G T G T T C A G T T T A C C T T T G G A A G G T G A G T T G
790 800 810
SER SER GLU GLY ASN LEU SER PRO SER GLN
C T C C T C C G A A G G C A A T C T A A G C C C T T C T C A
G A G G A G G C T T C C G T T A G A T T C G G G A A G A G T
820 830 840
VAL SER THR THR SER GLU HIS PRO SER GLN
A G T C T C C A C A A C A T C C G A G C A C C C A T C A C A
T C A G A G G T G T T G T A G G C T C G T G G G T A G T G T
850 860 870
PRO SER SER PRO PRO ASN THR THR ARG GLN
A C C C T C A T C T C C A C C C A A C A C A A C A C G C C A
T G G G A G T A G A G G T G G G T T G T G T T G T G C G G T
880 890 900

```
  ***
G T A G T T A T T A A A A A A A A A A A A
C A T C A A T A A T T T T T T T T T T T T
              910                    920
```

NUCLEOTIDE SEQUENCE OF THE RSV G GENE. THE cDNA SEQUENCE IS SHOWN IN THE PLUS (mRNA) STRAND SENSE IN THE 5' TO 3' DIRECTION. THE TRANSMEMBRANE (TM) ANCHOR DOMAIN IS UNDERLINED. AMINO ACIDS DIFFERING FROM THE PUBLISHED PRIMARY SEQUENCE OF THE PROTEIN ENCODED BY THE RSV G GENE ARE BOXED.

FIG.7D.

RESTRICTION MAP OF RSV G GENE

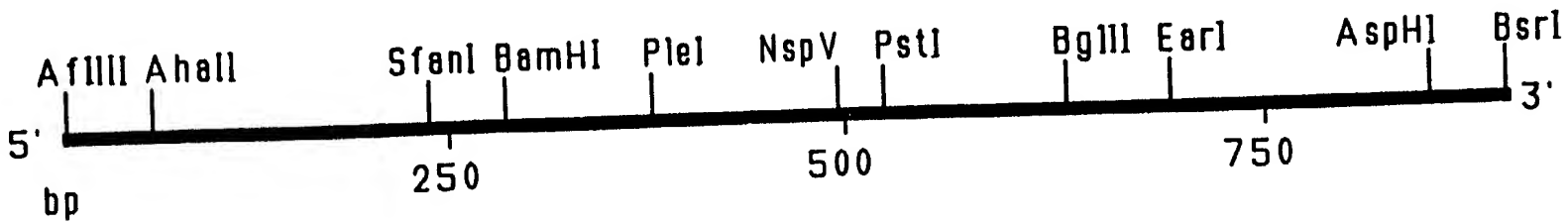


FIG.8.

Construction of a Bluescript-based expression vector containing the chimeric F_{PIV-3} -F_{RSV} gene with the 5' untranslated region of the PIV-3 F gene intact but lacking the nucleotide sequences coding for the hydrophobic anchor domains and cytoplasmic tails of both the PIV-3 and RSV F genes.

Step 1: Preparation of the plasmid containing the modified PIV-3 F gene

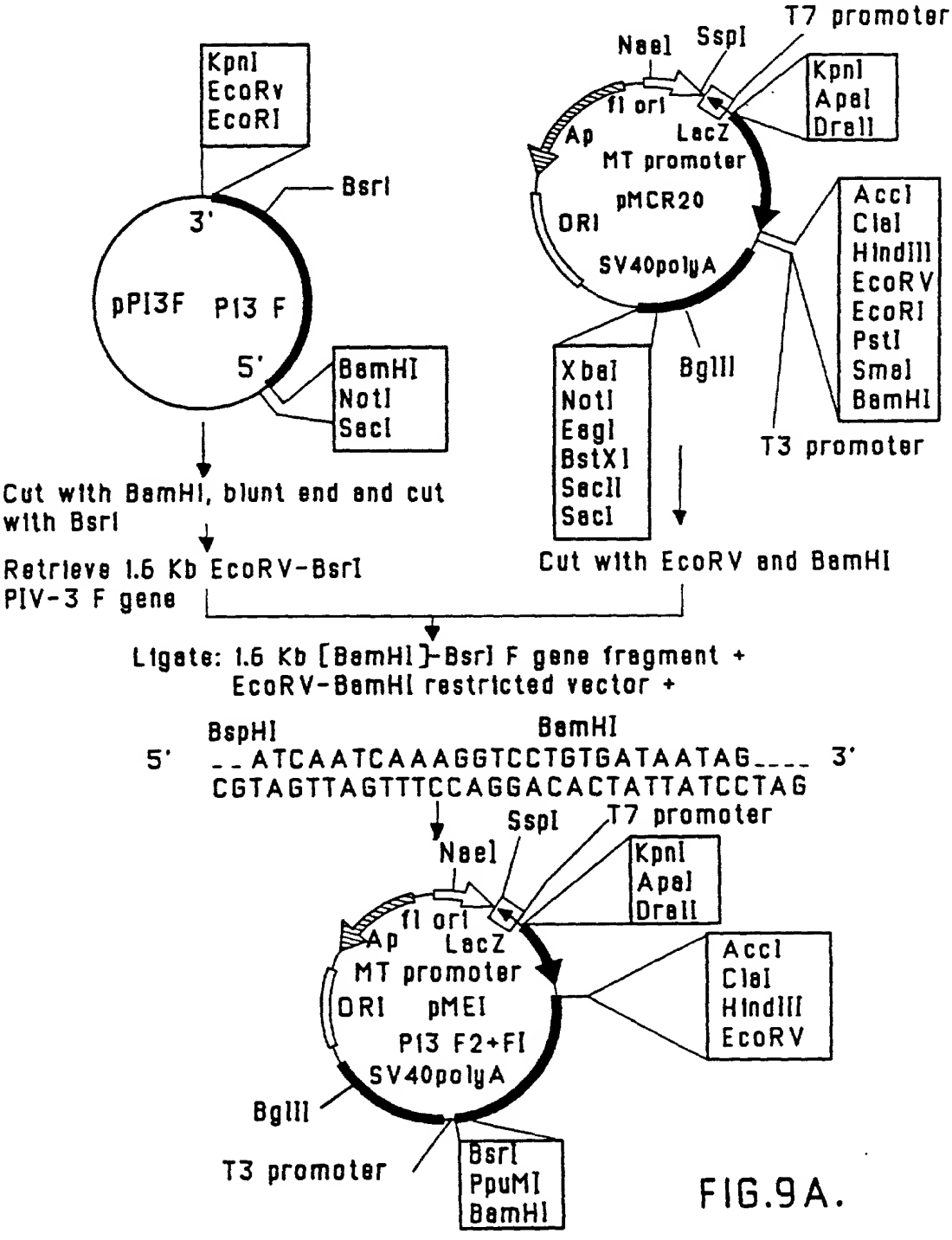
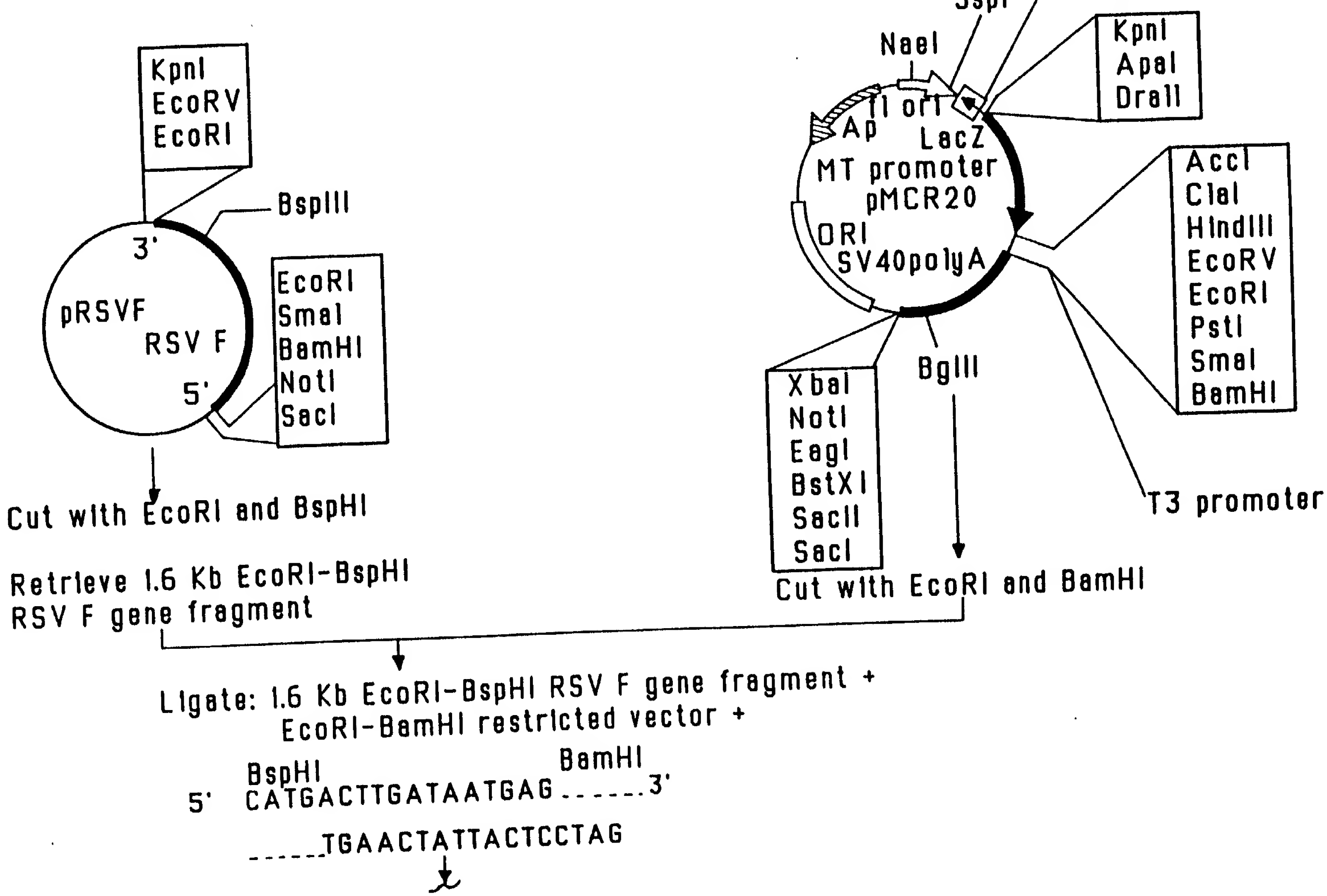


FIG.9A.

FIG.9B. Step 2: Preparation of the plasmid containing the modified RSV F gene



Ligate: 1.6 Kb EcoRI-BspHI RSV F gene fragment +
EcoRI-BamHI restricted vector +

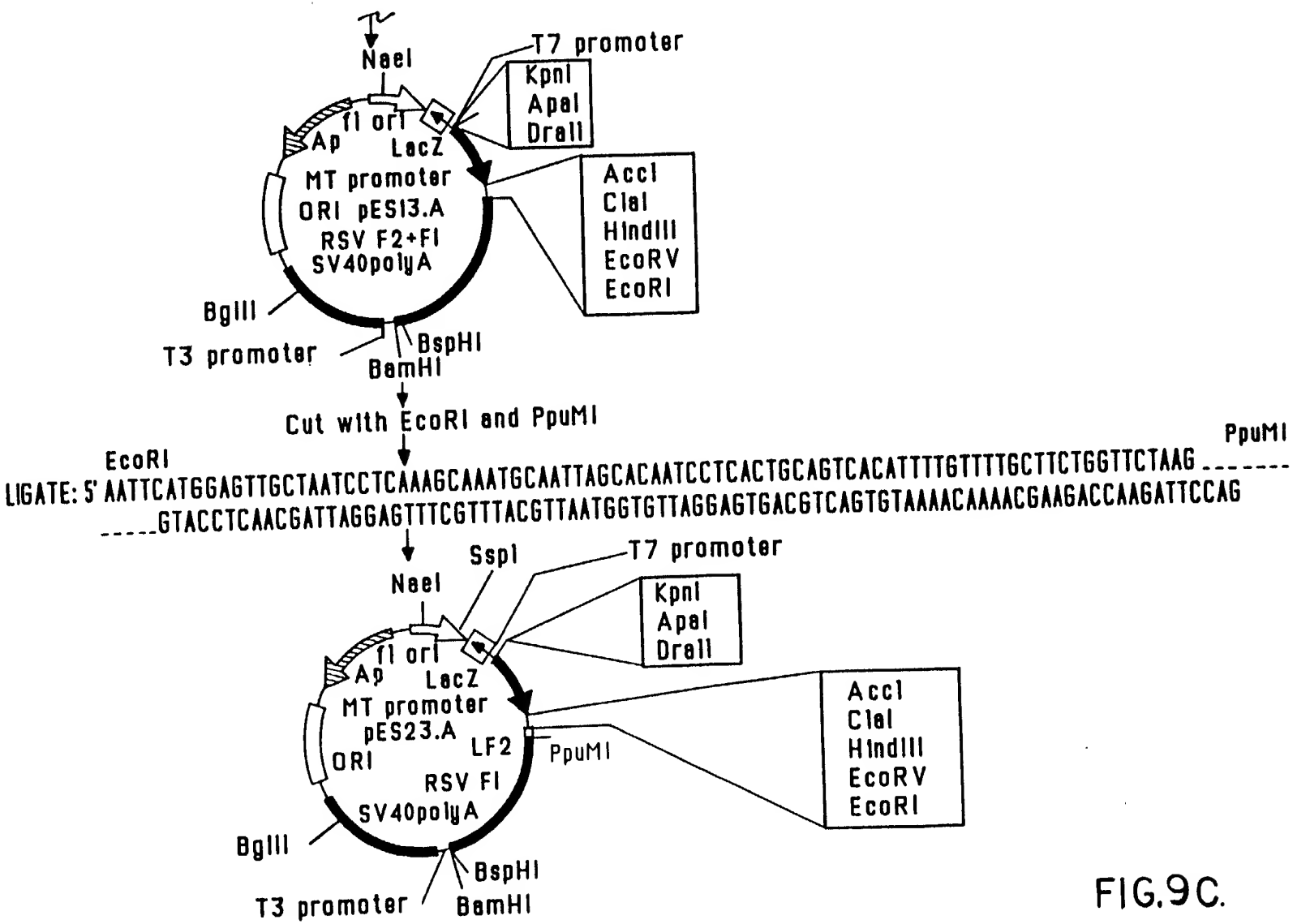
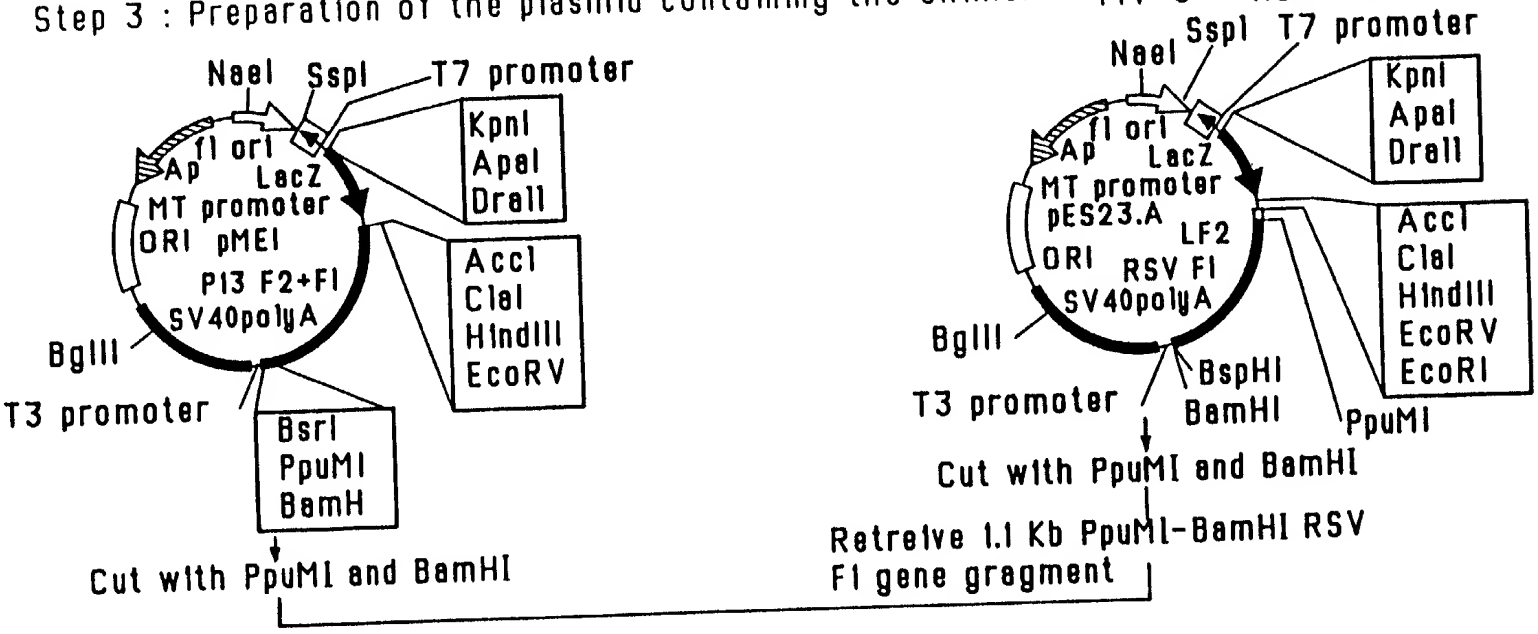

NaeI
T7 promoter
KpnI
ApaI
DraII
f1 ori
Ap
LacZ
MT promoter
ORI pES13.A
RSV F2+FI
SV40polyA
AccI
ClaI
HindIII
EcoRV
EcoRI
BglII
T3 promoter
BspHI
BamHI
Cut with EcoRI and PpuMI
EcoRI
PpuMI
LIGATE: 5' AATTCATGGAGTTGCTAATCCTCAAAGCAAATGCAATTAGCACAATCCTCACTGCAGTCACATTTTGTTTTGCTTCTGGTTCTAAG -------
-----GTACCTCAACGATTAGGAGTTTCGTTTACGTTAATGGTGTTAGGAGTGACGTCAGTGTAAAACAAAACGAAGACCAAGATTCCAG
SspI
NaeI
T7 promoter
KpnI
ApaI
DraII
f1 ori
Ap
LacZ
MT promoter
pES23.A
ORI
LF2
PpuMI
RSV FI
SV40polyA
AccI
ClaI
HindIII
EcoRV
EcoRI
BglII
BspHI
T3 promoter
BamHI

FIG.9C.

Step 3 : Preparation of the plasmid containing the chimeric F_{PIV-3} - F_{RSV} gene



Ligate: PpuMI-BamHI restricted vector +
1.1 Kd PpuMI-BamHI RSV F1 fragment

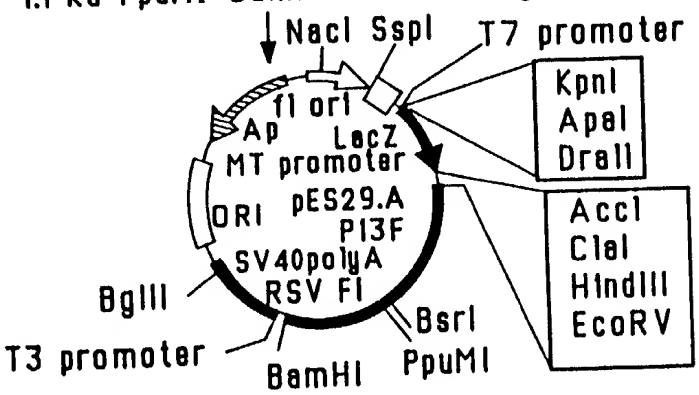


FIG.9D.

Construction of a Bluescript-based expression vector containing the PIV-3 F gene lacking the 5' untranslated sequence and transmembrane anchor and cytoplasmic tail coding regions.

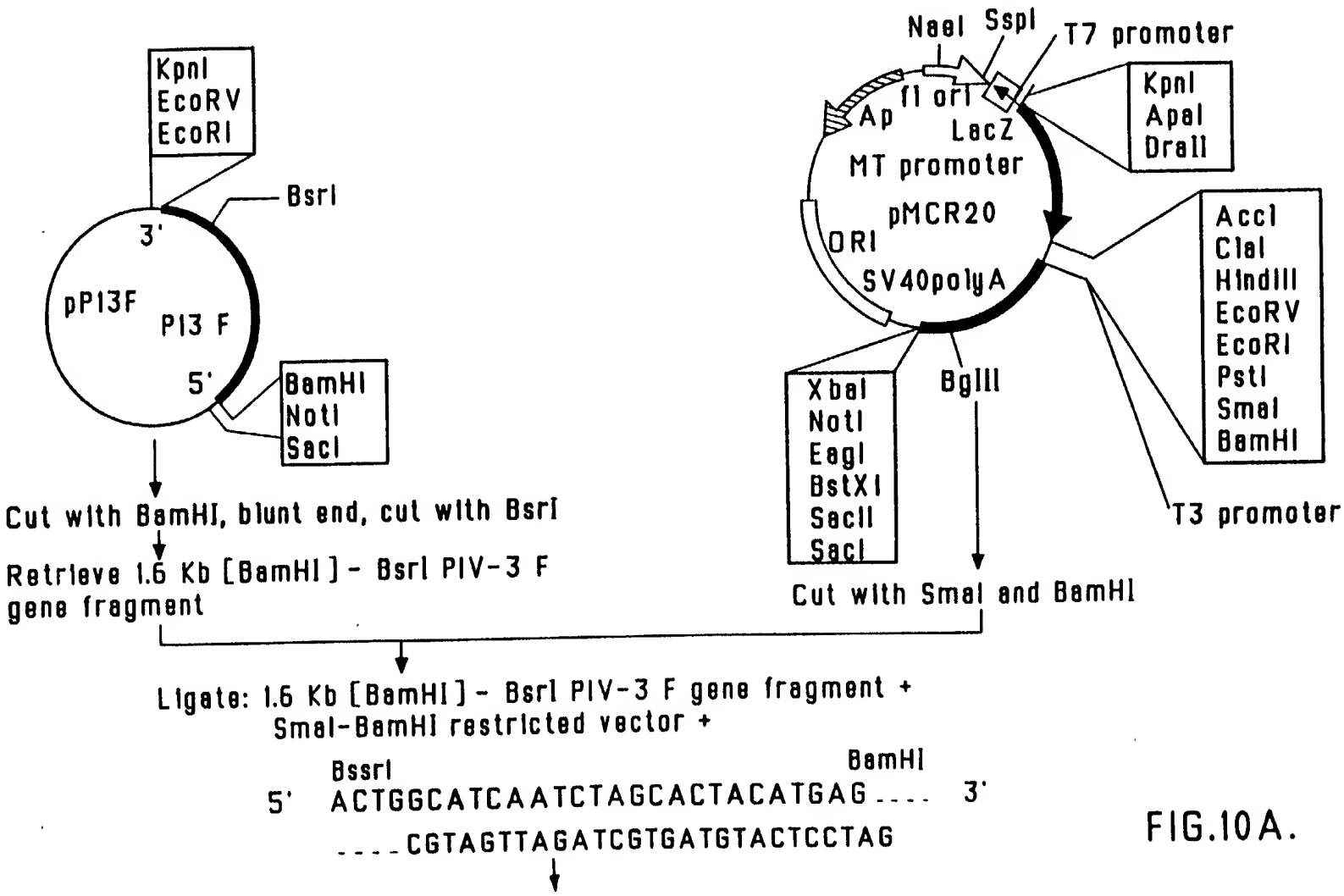
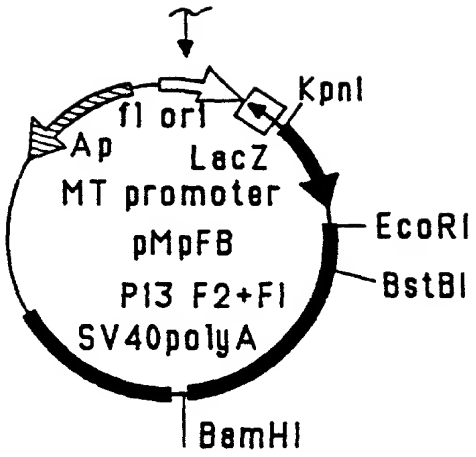


FIG.10A.

FIG.10B.



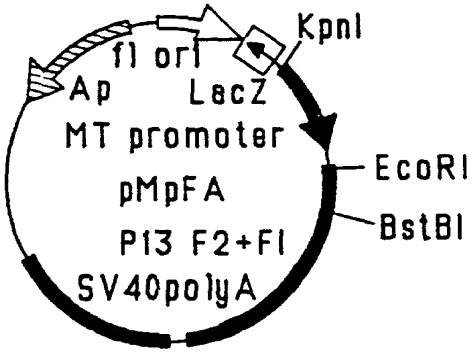
Cut with EcoRI and BstBI

Retreive: EcoRI-BstBI restricted vector

Ligate: EcoRI-BstBI restricted vector +

EcoRI PpuMI

AATTCATGCCAACTTTAATACTGCTAATTATTACAACAATGATTATGG
CATCTTCCTGCCAAATAGATATCACAAAACTACAGCAATGTAGGTGTA
TTGGTCAACAGTCCCAAAGGGATGAAGATATCACAAAACTT____ 3'
____GTACGGTTGAAATTATGACGATTAATAATGTTGTTACTAATACC
GTAGAAGGACGGTTTATCTATAGTGTTTTGATGTCGTACATCCACATA
ACCAGTTGTCAGGGTTTCCCTACTTCTATAGTGTTTTGAAGCTT



Construction of the chimeric F_{PIV-3}-F_{RSV} gene consisting of the truncated PIV-3 F gene devoid of the 5' untranslated region linked to the truncated RSV F1 gene.

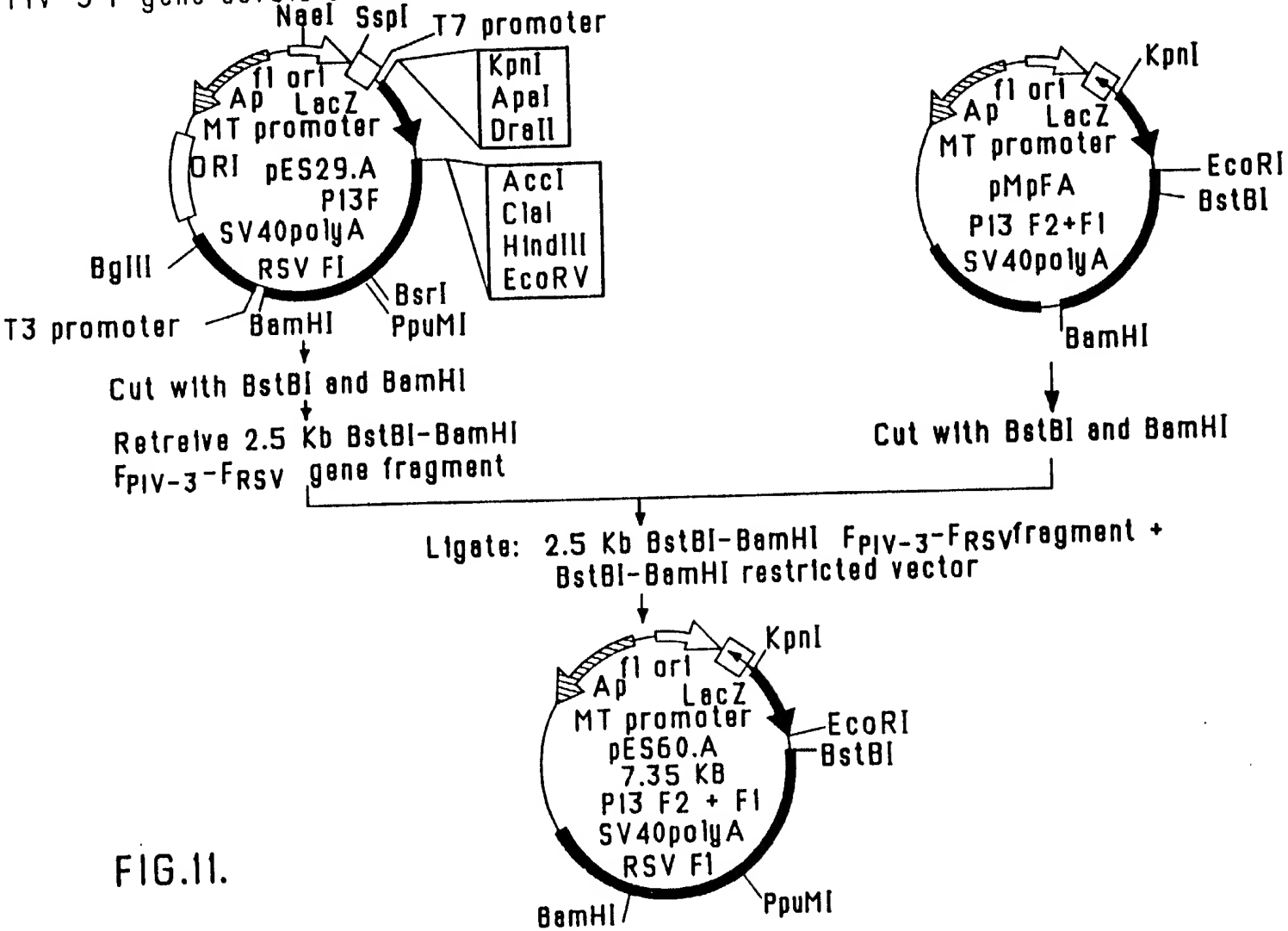
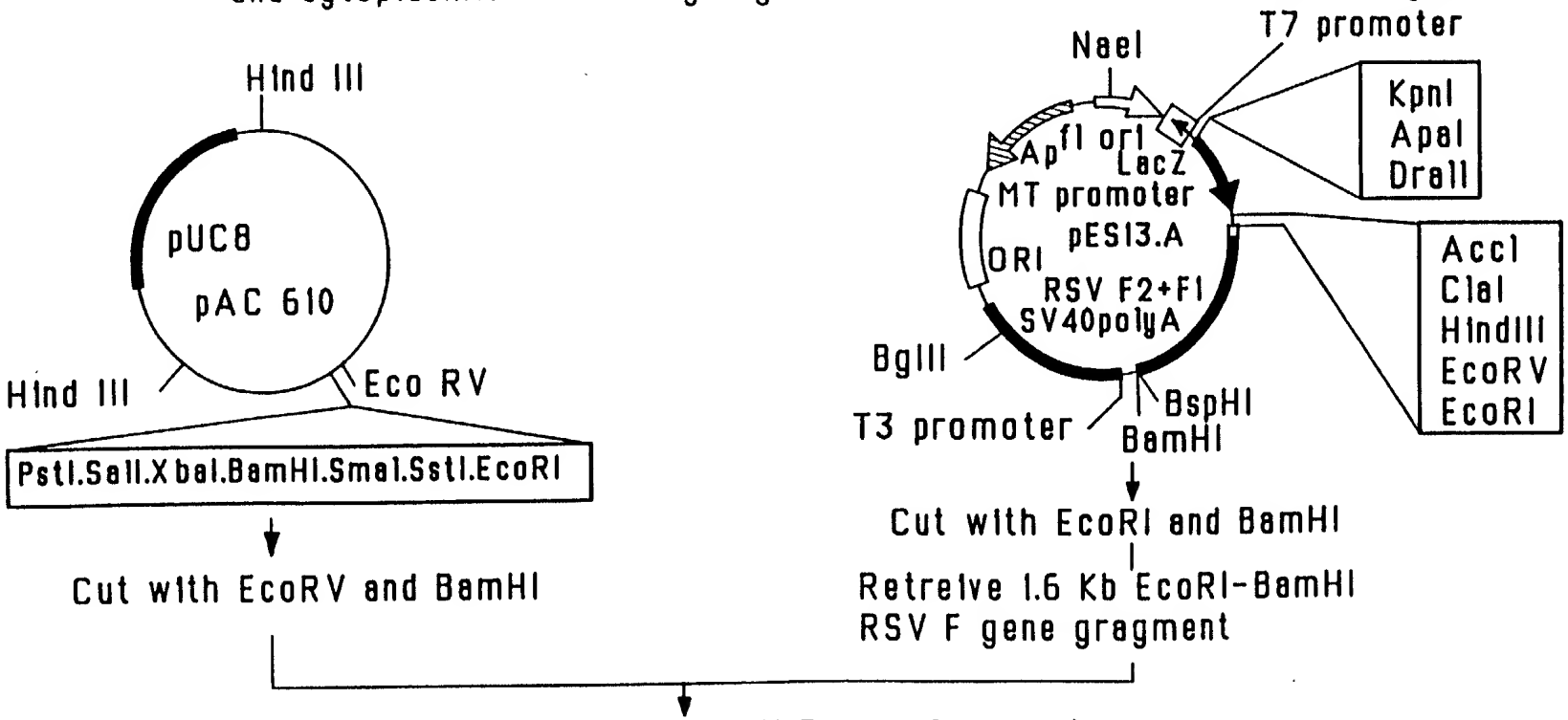


FIG.11.

FIG.12A. Construction of the modified pAc 610 baculovirus expression vector containing the chimeric F_{PIV-3}-F_{RSV} gene consisting of the PIV-3 F gene lacking both the 5' untranslated sequence as well as the transmembrane and cytoplasmic tail coding regions linked to the truncated RSV F1 gene



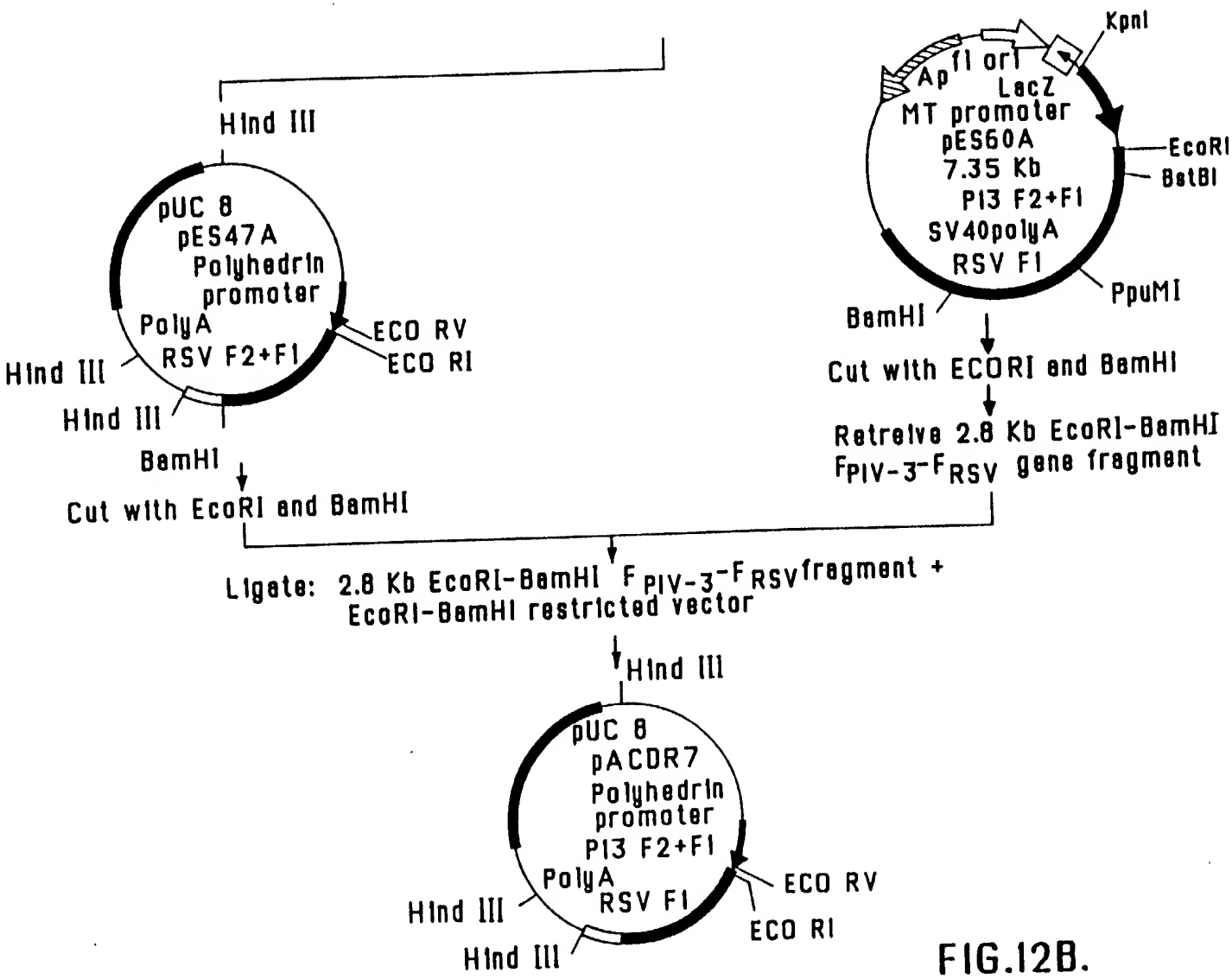

Hind III
pUC 8
pES47A
Polyhedrin
promoter
PolyA
RSV F2+F1
ECO RV
ECO RI
Hind III
Hind III
BamHI
Cut with EcoRI and BamHI
KpnI
Ap
f1 ori
LacZ
MT promoter
pES60A
7.35 Kb
P13 F2+F1
SV40polyA
RSV F1
EcoRI
BstBI
PpuMI
BamHI
Cut with ECORI and BamHI
Retreive 2.8 Kb EcoRI-BamHI
FPIV-3-FRSV gene fragment
Ligate: 2.8 Kb EcoRI-BamHI FPIV-3-FRSV fragment +
EcoRI-BamHI restricted vector
Hind III
pUC 8
pACDR7
Polyhedrin
promoter
P13 F2+F1
PolyA
RSV F1
ECO RV
ECO RI
Hind III
Hind III

FIG.12B.

FIG.13

IMMUNOBLOTS OF CELL LYSATES FROM Sf9 CELLS INFECTED WITH RECOMBINANT BACULOVIRUSES

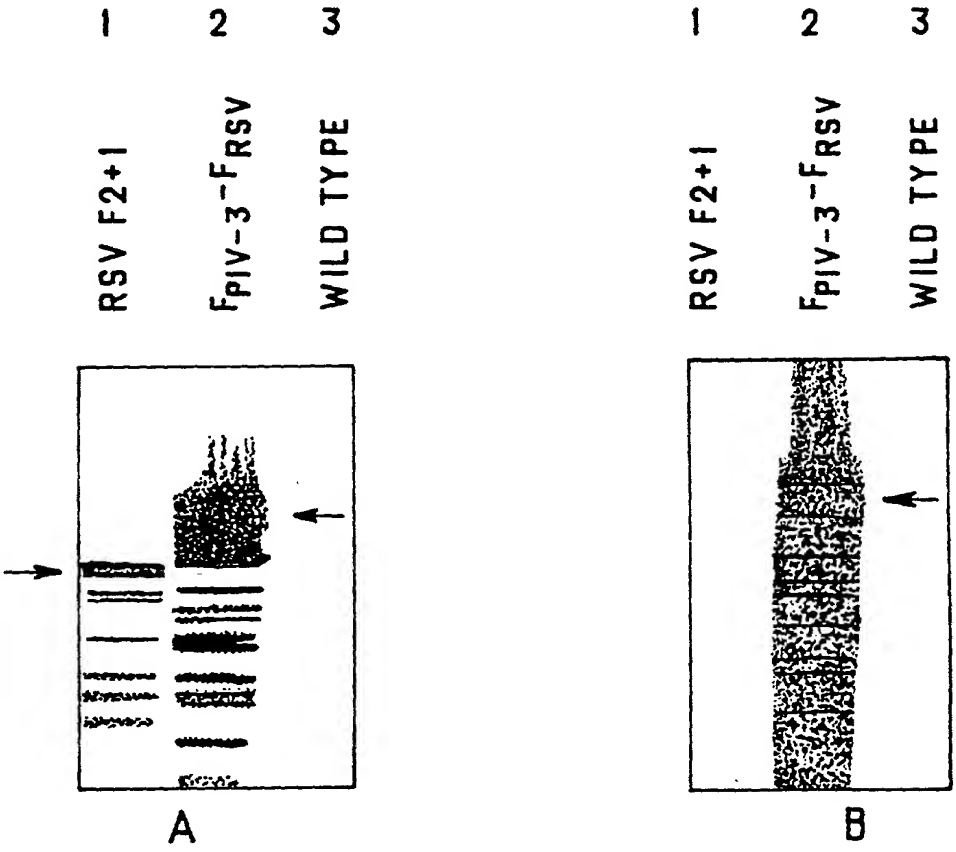


FIG 13 : Immunoblots of cell lysates from Sf9 cells infected wirth recombinant baculoviruses containing the truncated RSV F gene (Lane 1), the chimeric F_{PIV-3}-F_{RSV}gene (Lane 2) or infected with wild type virus (Lane 3) reacted with anti-F RSV Mab (panel A) and anti-F1 PIV-3 antiserum (panel B)

FIG.14. CONSTRUCTION OF THE BACULOVIRUS TRANSFER VECTOR pD2

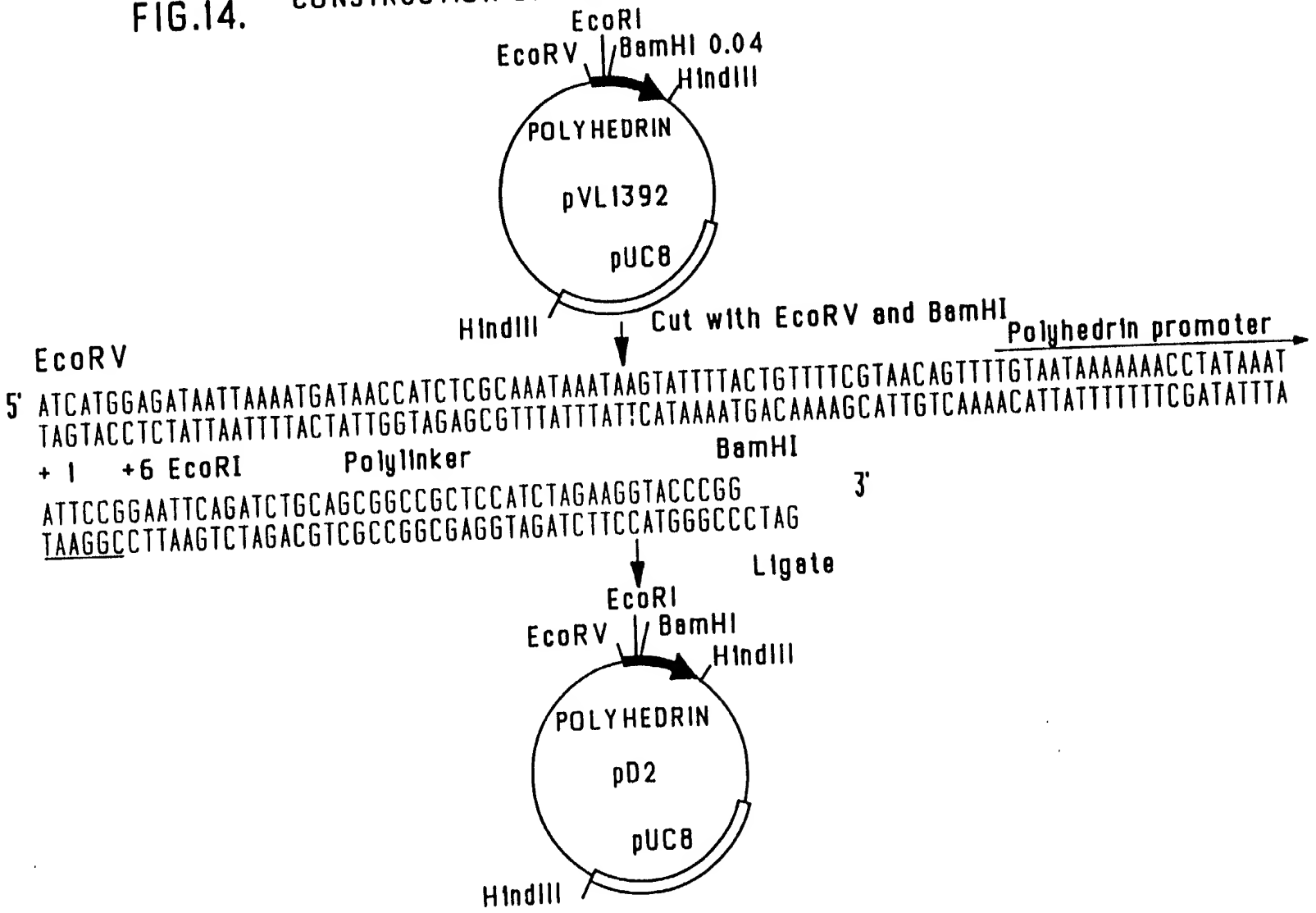


FIG.15A. CONSTRUCTION OF THE F_{RSV}-HN_{PIV3} CHIMERIC GENE

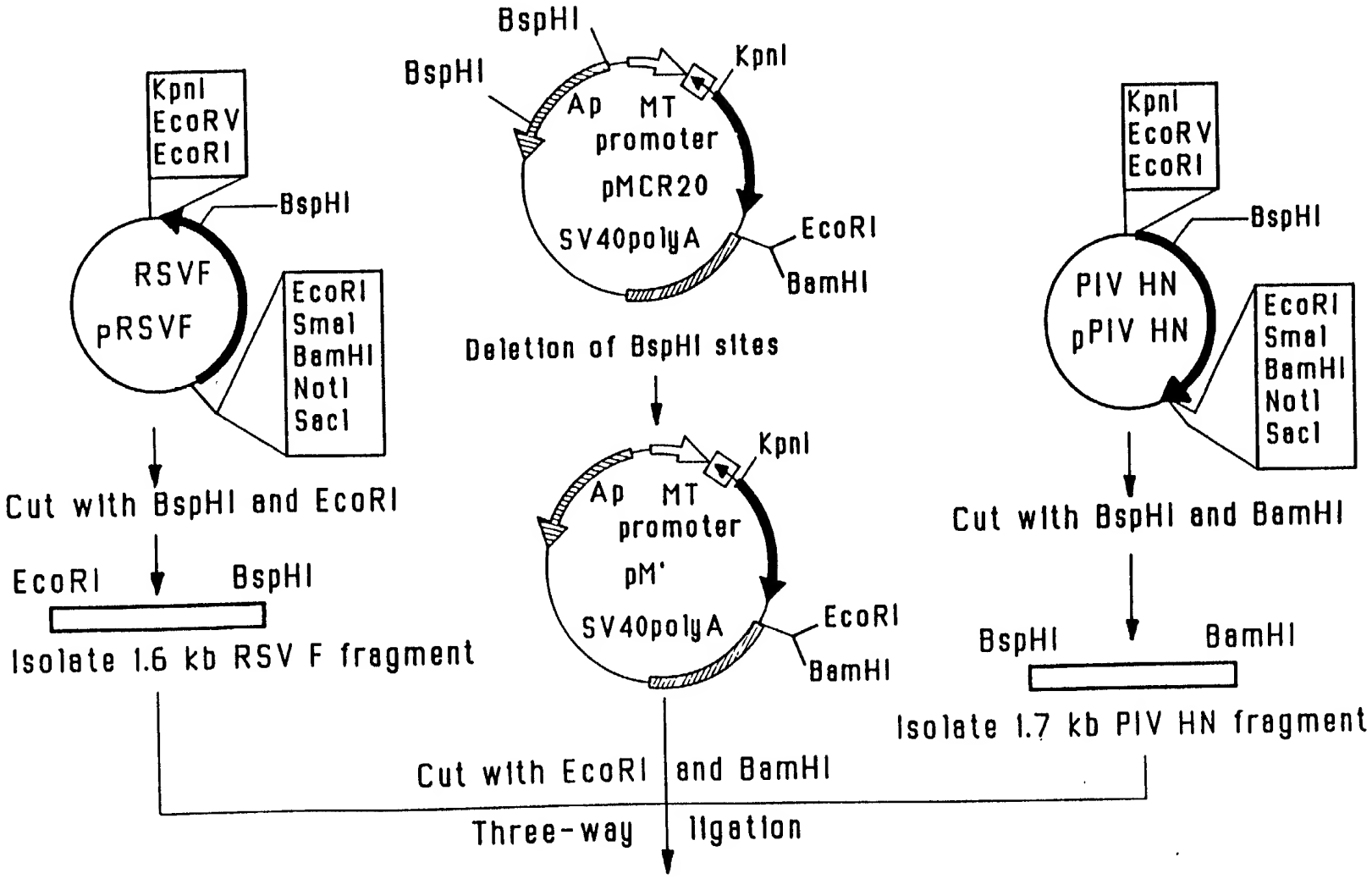


FIG.15B.

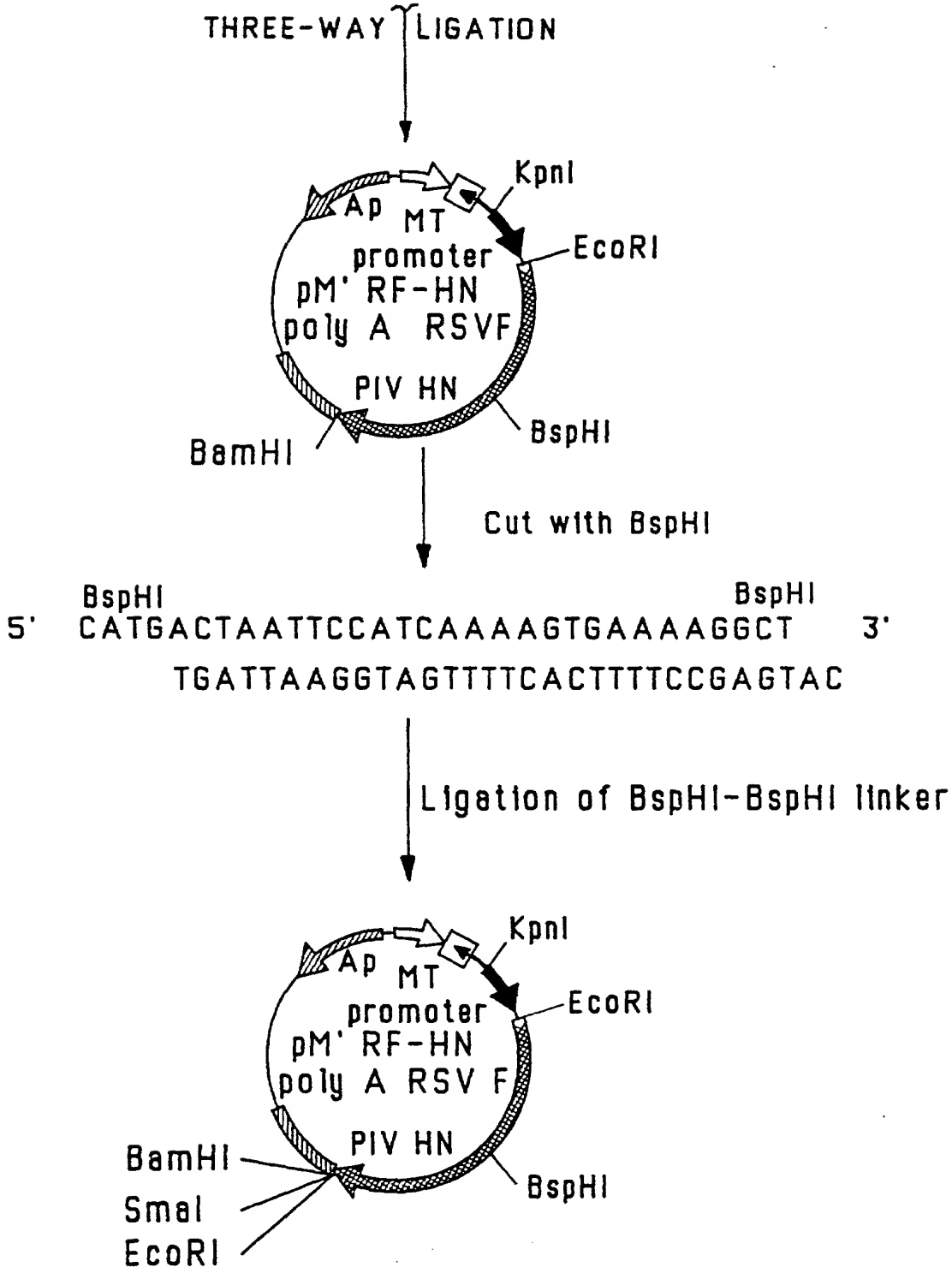

THREE-WAY LIGATION
KpnI
Ap
MT
promoter
EcoRI
pM' RF-HN
poly A RSVF
PIV HN
BamHI
BspHI
Cut with BspHI
BspHI
BspHI
5' CATGACTAATTCCATCAAAAGTGAAAAGGCT 3'
TGATTAAGGTAGTTTTCACTTTTCCGAGTAC
Ligation of BspHI-BspHI linker
KpnI
Ap
MT
promoter
EcoRI
pM' RF-HN
poly A RSV F
PIV HN
BamHI
SmaI
EcoRI
BspHI

FIG.16

SDS POLY ACRYLAMIDE GEL AND IMMUNOBLOTS OF PURIFIED F_{RSV}-HN_{PIV-3} CHIMERIC PROTEIN

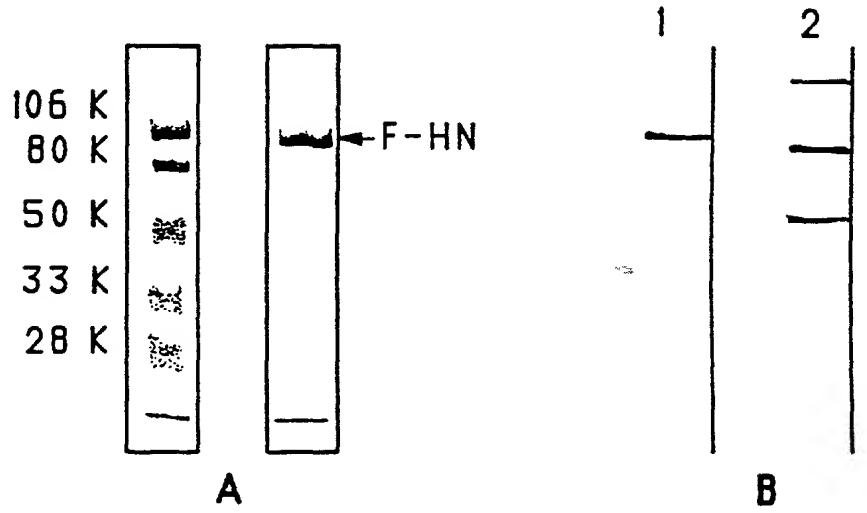


FIG 16 : A) Coomassie-stained SDS polyacrylamide gel of immunoaffinity- purified F_{RSV}-HN_{PIV-3}protein.

B) Immunoblots of F_{RSV}-HN_{PIV-3}protein reacted with an anti-F RSV Mab (lane 1) and anti-HN PIV-3 antiserum (lane 2)

FIG.17. MUTAGENESIS OF THE PIV-3 F GENE

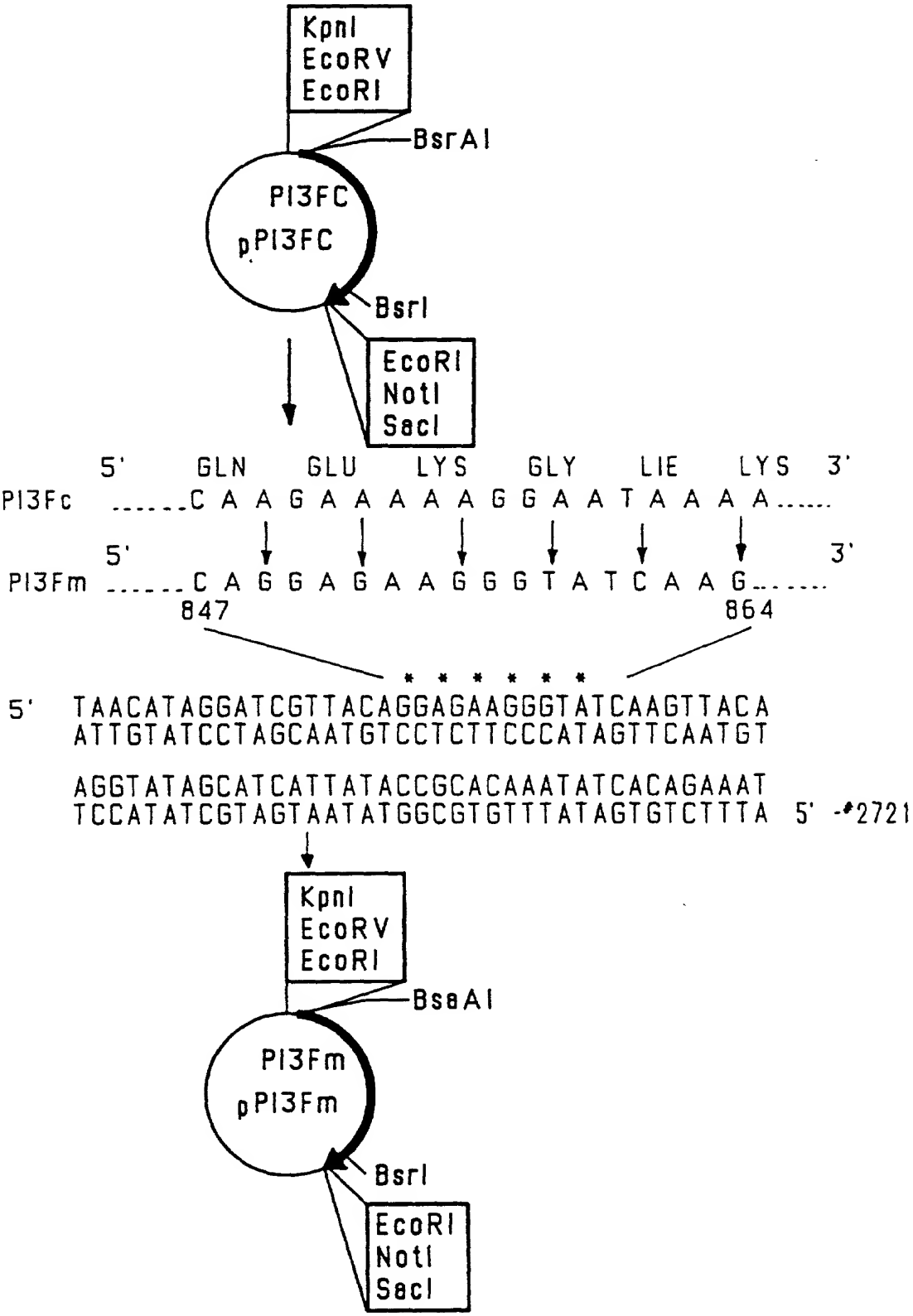
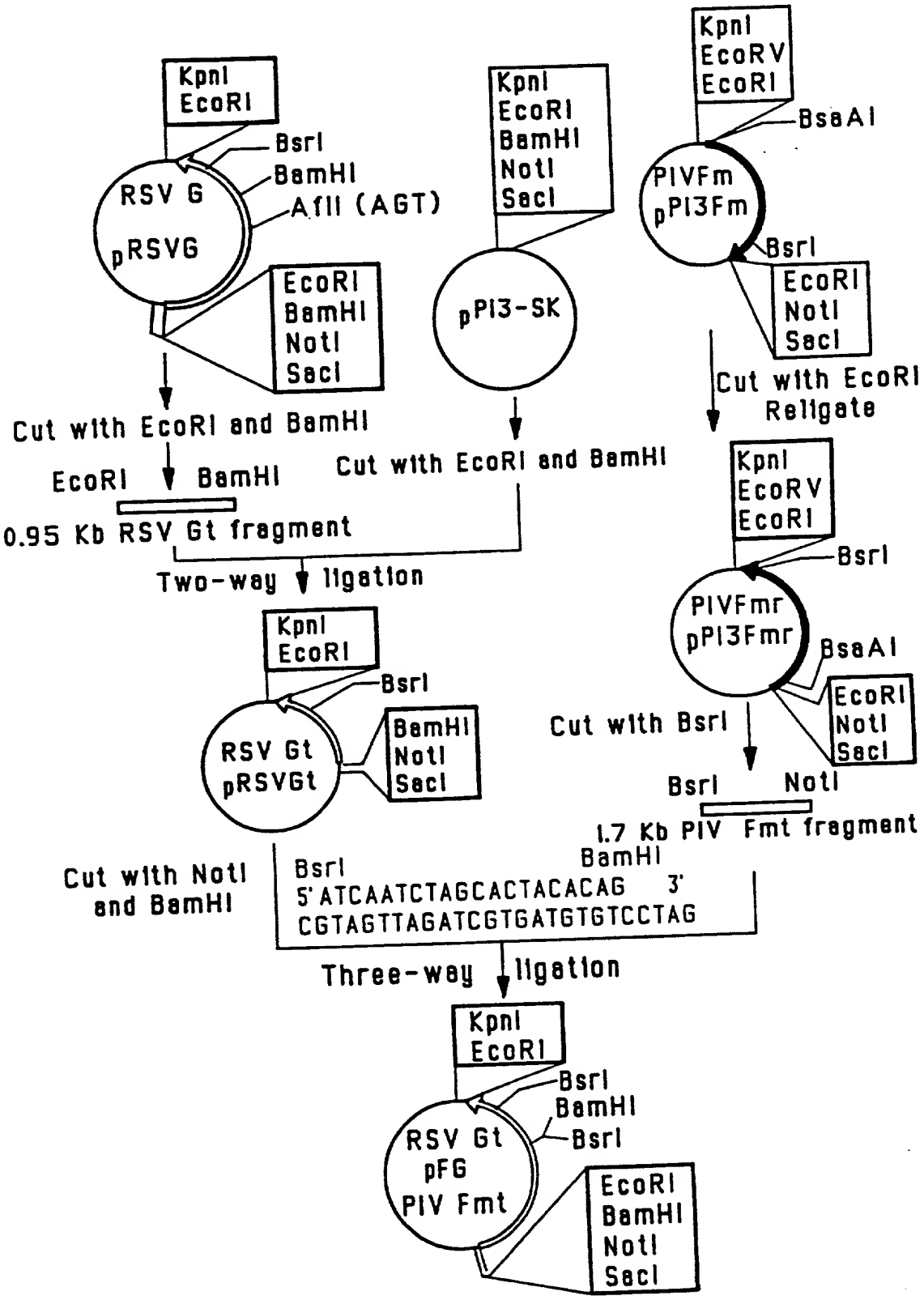


FIG.18. CONSTRUCTION OF THE F_{PIV3}-G_{RSV} CHIMERIC GENE



Combined Declaration and Power of Attorney for United States Patent Application

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name;

I believe I am the original, first and joint/sole inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled: **CHIMERIC IMMUNOGENS**, the specification of which is attached hereto.

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, S.1.56(a).

I hereby claim foreign priority benefits under Title 35, United States Code, S.119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed Yes	No
9200117.1 (Number)	GB (Country)	06/01/92 (Day/Month/Year Filed)	X	

I hereby claim the benefit under Title 35, United States Code, S.120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, S.112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, S.1.56(a) which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

(Appln. Serial No.)	(Filing Date)	(Status) (patented, pending, abandoned)

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that wilful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such wilful false statements may jeopardize the validity of the application or any patent issued thereon.

I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

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Reg. No. 27,093; Stephen J. Perry, Reg. No. 32,107; Patricia A. Rae, Reg. No. 33,570 and John R. Orange, Reg. No. 29,725.

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at
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(416) 595-1155

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Date

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Citizenship: Canadian

Post Office Address: 16 Monro Boulevard, Willowdale, Ontario, Canada, L4Z 1M5

Full name of second inventor: Run-Pan Du

Inventor's signature ______________ Feb 1 1993
Date

Residence: Thornhill, Ontario, Canada

Citizenship: Canadian

Post Office Address: 299 Chelwood Drive, Thornhill, Ontario, Canada, L4J 7Y8

Full name of third inventor: Mary E. Ewasyshyn

Inventor's signature ______________ Feb 1 1993
Date

Residence: Willowdale, Ontario, Canada

Citizenship: Canadian

Post Office Address: 120 Torresdale, Apt. 1506, Willowdale, Ontario, Canada, M2R 3N7

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EWASYSHYN, Mary E

(ii) TITLE OF INVENTION: INFECTION DETECTION METHOD USING CHIMERIC PROTEIN

(iii) NUMBER OF SEQUENCES: 38

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(C) CITY: Toronto
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(E) COUNTRY: Canada
(F) ZIP: M5G 1R7

(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/467,961
(B) FILING DATE: 06-JUN-1995

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/001,554
(B) FILING DATE: 06-JAN-1993

(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: GB 9200117.1
(B) FILING DATE: 06-JAN-1992

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(C) REFERENCE/DOCKET NUMBER: 1038-1000 MIS:jb

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(A) TELEPHONE: (416) 595-1155
(B) TELEFAX: (416) 595-1163

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1844 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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CATCCAATCC ATTTTAAACA AAAATTCCAA AAGAGACCGG CAACACAACA AGCACCAAAC 180

ACAATGCCAA CTTTAATACT GCTAATTATT ACAACAATGA TTATGGCATC TTCCTGCCAA 240

ATAGATATCA CAAAACTACA GCATGTAGGT GTATTGGTCA ACAGTCCCAA AGGGATGAAG 300

ATATCACAAA ACTTCGAAAC AAGATATCTA ATTTTGAGCC TCATACCAAA AATAGAAGAC 360

TCTAACTCTT GTGGTGACCA ACAGATCAAA CAATACAAGA GGTTATTGGA TAGACTGATC 420

ATCCCTCTAT ATGATGGATT AAGATTACAG AAAGATGTGA TAGTAACCAA TCAAGAATCC 480

AATGAAAACA CTGATCCCAG AACAAGACGA TCCTTTGGAG GGGTAATTGG AACCATTGCT 540

CTGGGAGTAG CAACCTCAGC ACAAATTACA GCGGCAGTTG CTCTGGTTGA AGCCAAGCAG 600

GCAAAATCAG ACATCGAAAA ACTCAAAGAA GCAATCAGGG ACACAAACAA AGCAGTGCAG 660

TCAGTTCAGA GCTCTATAGG AAATTTAATA GTAGCAATTA AATCAGTCCA AGATTATGTC 720

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ACAGAAATAT TCACAACATC AACAGTTGAT AAATATGATA TCTATGATCT ATTATTTACA 960

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TCATATAATA TCCAAAACAG AGAATGGTAT ATCCCTCTTC CCAGCCATAT CATGACGAAA 1140

GGGGCATTTC TAGGTGGAGC AGATGTCAAG GAATGTATAG AAGCATTCAG CAGTTATATA 1200

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CTTAACAAAG CCAAATCAGA TCTAGAAGAA TCAAAAGAAT GGATAAGAAG GTCAAATCAA 1620

AAACTAGATT CTATTGGAAA CTGGCATCAA TCTAGCACTA CAATCATAAT TATTTTAATA 1680

ATGATCATTA TATTGTTTAT AATTAATGTA ACGATAATTA CAATTGCAAT TAAGTATTAC 1740

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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 539 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Asn Ser Pro Lys Gly Met Lys Ile Ser Gln Asn Phe Glu Thr Arg Tyr
        35                  40                  45

Leu Ile Leu Ser Leu Ile Pro Lys Ile Glu Asp Ser Asn Ser Cys Gly
    50                  55                  60

Asp Gln Gln Ile Lys Gln Tyr Lys Arg Leu Leu Asp Arg Leu Ile Ile
65                  70                  75                  80

Pro Leu Tyr Asp Gly Leu Arg Leu Gln Lys Asp Val Ile Val Thr Asn
                85                  90                  95

Gln Glu Ser Asn Glu Asn Thr Asp Pro Arg Thr Arg Arg Ser Phe Gly
            100                 105                 110

Gly Val Ile Gly Thr Ile Ala Leu Gly Val Ala Thr Ser Ala Gln Ile
        115                 120                 125

Thr Ala Ala Val Ala Leu Val Glu Ala Lys Gln Ala Lys Ser Asp Ile
    130                 135                 140

Glu Lys Leu Lys Glu Ala Ile Arg Asp Thr Asn Lys Ala Val Gln Ser
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145 150 155 160

Val Gln Ser Ser Ile Gly Asn Leu Ile Val Ala Ile Lys Ser Val Gln
165 170 175

Asp Tyr Val Asn Asn Glu Ile Val Pro Ser Ile Ala Arg Leu Gly Cys
180 185 190

Glu Ala Ala Gly Leu Gln Leu Gly Ile Ala Leu Thr Gln His Tyr Ser
195 200 205

Glu Leu Thr Asn Ile Phe Gly Asp Asn Ile Gly Ser Leu Gln Glu Lys
210 215 220

Gly Ile Lys Leu Gln Gly Ile Ala Ser Leu Tyr Arg Thr Asn Ile Thr
225 230 235 240

Glu Ile Phe Thr Thr Ser Thr Val Asp Lys Tyr Asp Ile Tyr Asp Leu
245 250 255

Leu Phe Thr Glu Ser Ile Lys Val Arg Val Ile Asp Val Asp Leu Asn
260 265 270

Asp Tyr Ser Ile Thr Leu Gln Val Arg Leu Pro Leu Leu Thr Arg Leu
275 280 285

Leu Asn Thr Gln Ile Tyr Lys Val Asp Ser Ile Ser Tyr Asn Ile Gln
290 295 300

Asn Arg Glu Trp Tyr Ile Pro Leu Pro Ser His Ile Met Thr Lys Gly
305 310 315 320

Ala Phe Leu Gly Gly Ala Asp Val Lys Glu Cys Ile Glu Ala Phe Ser
325 330 335

Ser Tyr Ile Cys Pro Ser Asp Pro Gly Phe Val Leu Asn His Glu Met
340 345 350

Glu Ser Cys Leu Ser Gly Asn Ile Ser Gln Cys Pro Arg Thr Thr Val
355 360 365

Thr Ser Asp Ile Val Pro Arg Tyr Ala Phe Val Asn Gly Gly Val Val
370 375 380

Ala Asn Cys Ile Thr Thr Thr Cys Thr Cys Asn Gly Ile Asp Asn Arg
385 390 395 400

Ile Asn Gln Pro Pro Asp Gln Gly Val Lys Ile Ile Thr His Lys Glu
405 410 415

Cys Asn Thr Ile Gly Ile Asn Gly Met Leu Phe Asn Thr Asn Lys Glu
420 425 430

Gly Thr Leu Ala Phe Tyr Thr Pro Asn Asp Ile Thr Leu Asn Asn Ser
435 440 445

Val Ala Leu Asp Pro Ile Asp Ile Ser Ile Glu Leu Asn Lys Ala Lys

```
     450                 455                 460

Ser Asp Leu Glu Glu Ser Lys Glu Trp Ile Arg Arg Ser Asn Gln Lys
465                 470                 475                 480

Leu Asp Ser Ile Gly Asn Trp His Gln Ser Ser Thr Thr Ile Ile Ile
                485                 490                 495

Ile Leu Ile Met Ile Ile Ile Leu Phe Ile Ile Asn Val Thr Ile Ile
            500                 505                 510

Thr Ile Ala Ile Lys Tyr Tyr Arg Ile Gln Lys Arg Asn Arg Val Asp
        515                 520                 525

Gln Asn Asp Lys Pro Tyr Val Leu Thr Asn Lys
    530                 535
```

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1833 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

```
AGACAAATCC AAATTCGAGA TGGAATACTG GAAGCATACC AATCACGGAA AGGATGCTGG      60

CAATGAGCTG GAGACGTCCA TGGCTACTAA TGGCAACAAG CTCACCAATA AGATAACATA     120

TATATTATGG ACAATAATCC TGGTGTTATT ATCAATAGTC TTCATCATAG TGCTAATTAA     180

TTCCATCAAA AGTGAAAAGG CTCATGAATC ATTGCTGCAA GACATAAATA ATGAGTTTAT     240

GGAAATTACA GAAAAGATCC AAATGGCATC GGATAATACC AATGATCTAA TACAGTCAGG     300

AGTGAATACA AGGCTTCTTA CAATTCAGAG TCATGTCCAG AATTATATAC CAATATCACT     360

GACACAACAG ATGTCAGATC TTAGGAAATT CATTAGTGAA ATTACAATTA GAAATGATAA     420

TCAAGAAGTG CTGCCACAAA GAATAACACA TGATGTGGGT ATAAAACCTT TAAATCCAGA     480

TGATTTTTGG AGATGCACGT CTGGTCTTCC ATCTTTAATG AAAACTCCAA AAATAAGGTT     540

AATGCCAGGG CCGGGATTAT TAGCTATGCC AACGACTGTT GATGGCTGTA TCAGAACTCC     600

GTCCTTAGTT ATAAATGATC TGATTTATGC TTATACCTCA AATCTAATTA CTCGAGGTTG     660

TCAGGATATA GGAAAATCAT ATCAAGTCTT ACAGATAGGG ATAATAACTG TAAACTCAGA     720

CTTGGTACCT GACTTAAATC CCAGGATCTC TCATACTTTT AACATAAATG ACAATAGGAA     780

GTCATGTTCT CTAGCACTCC TAAATACAGA TGTATATCAA CTGTGTTCAA CTCCCAAAGT     840
```

TGATGAAAGA TCAGATTATG CATCATCAGG CATAGAAGAT ATTGTACTTG ATATTGTCAA 900

TTATGATGGC TCAATCTCAA CAACAAGATT TAAGAATAAT AACATAAGCT TTGATCAACC 960

TTATGCTGCA CTATACCCAT CTGTTGGACC AGGGATATAC TACAAAGGCA AAATAATATT 1020

TCTCGGGTAT GGAGGTCTTG AACATCCAAT AAATGAGAAT GTAATCTGCA ACACAACTGG 1080

GTGTCCCGGG AAAACACAGA GAGACTGCAA TCAGGCATCT CATAGTCCAT GGTTTTCAGA 1140

TAGGAGGATG GTCAACTCTA TCATTGTTGT TGACAAAGGC TTAAACTCAA TTCCAAAATT 1200

GAAGGTATGG ACGATATCTA TGAGACAGAA TTACTGGGGG TCAGAAGGAA GGTTACTTCT 1260

ACTAGGTAAC AAGATCTATA TATATACAAG ATCCACAAGT TGGCATAGCA AGTTACAATT 1320

AGGAATAATT GATATTACTG ATTACAGTGA TATAAGGATA AAATGGACAT GGCATAATGT 1380

GCTATCAAGA CCAGGAAACA ATGAATGTCC ATGGGGACAT TCATGTCCAG ATGGATGTAT 1440

AACAGGAGTA TATACTGATG CATATCCACT CAATCCCACA GGGAGCATTG TGTCATCTGT 1500

CATATTAGAT TCACAAAAAT CGAGAGTGAA CCCAGTCATA ACTTACTCAA CAGCAACCGA 1560

AAGAGTAAAC GAGCTGGCCA TCCGAAACAG AACACTCTCA GCTGGATATA CAACAACAAG 1620

CTGCATCACA CACTATAACA AAGGATATTG TTTTCATATA GTAGAAATAA ATCAGAAAAG 1680

CTTAAACACA CTTCAACCCA TGTTGTTCAA GACAGAGGTT CCAAAAAGCT GCAGTTAATC 1740

ATAATTAACC GCAATATGCA TTAACCTATC TATAATACAA GTATATGATA AGTAATCAGC 1800

AATCAGACAA TAGACAAAAG GGAAATATAA AAA 1833

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 572 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Glu Tyr Trp Lys His Thr Asn His Gly Lys Asp Ala Gly Asn Glu
1 5 10 15

Leu Glu Thr Ser Met Ala Thr Asn Gly Asn Lys Leu Thr Asn Lys Ile
20 25 30

Thr Tyr Ile Leu Trp Thr Ile Ile Leu Val Leu Leu Ser Ile Val Phe
35 40 45

Ile Ile Val Leu Ile Asn Ser Ile Lys Ser Glu Lys Ala His Glu Ser
50 55 60

Leu Leu Gln Asp Ile Asn Asn Glu Phe Met Glu Ile Thr Glu Lys Ile
65 70 75 80

Gln Met Ala Ser Asp Asn Thr Asn Asp Leu Ile Gln Ser Gly Val Asn
85 90 95

Thr Arg Leu Leu Thr Ile Gln Ser His Val Gln Asn Tyr Ile Pro Ile
100 105 110

Ser Leu Thr Gln Gln Met Ser Asp Leu Arg Lys Phe Ile Ser Glu Ile
115 120 125

Thr Ile Arg Asn Asp Asn Gln Glu Val Leu Pro Gln Arg Ile Thr His
130 135 140

Asp Val Gly Ile Lys Pro Leu Asn Pro Asp Asp Phe Trp Arg Cys Thr
145 150 155 160

Ser Gly Leu Pro Ser Leu Met Lys Thr Pro Lys Ile Arg Leu Met Pro
165 170 175

Gly Pro Gly Leu Leu Ala Met Pro Thr Thr Val Asp Gly Cys Ile Arg
180 185 190

Thr Pro Ser Leu Val Ile Asn Asp Leu Ile Tyr Ala Tyr Thr Ser Asn
195 200 205

Leu Ile Thr Arg Gly Cys Gln Asp Ile Gly Lys Ser Tyr Gln Val Leu
210 215 220

Gln Ile Gly Ile Ile Thr Val Asn Ser Asp Leu Val Pro Asp Leu Asn
225 230 235 240

Pro Arg Ile Ser His Thr Phe Asn Ile Asn Asp Asn Arg Lys Ser Cys
245 250 255

Ser Leu Ala Leu Leu Asn Thr Asp Val Tyr Gln Leu Cys Ser Thr Pro
260 265 270

Lys Val Asp Glu Arg Ser Asp Tyr Ala Ser Ser Gly Ile Glu Asp Ile
275 280 285

Val Leu Asp Ile Val Asn Tyr Asp Gly Ser Ile Ser Thr Thr Arg Phe
290 295 300

Lys Asn Asn Asn Ile Ser Phe Asp Gln Pro Tyr Ala Ala Leu Tyr Pro
305 310 315 320

Ser Val Gly Pro Gly Ile Tyr Tyr Lys Gly Lys Ile Ile Phe Leu Gly
325 330 335

Tyr Gly Gly Leu Glu His Pro Ile Asn Glu Asn Val Ile Cys Asn Thr
340 345 350

Thr Gly Cys Pro Gly Lys Thr Gln Arg Asp Cys Asn Gln Ala Ser His
355 360 365

Ser Pro Trp Phe Ser Asp Arg Arg Met Val Asn Ser Ile Ile Val Val
370 375 380

Asp Lys Gly Leu Asn Ser Ile Pro Lys Leu Lys Val Trp Thr Ile Ser
385 390 395 400

Met Arg Gln Asn Tyr Trp Gly Ser Glu Gly Arg Leu Leu Leu Leu Gly
405 410 415

Asn Lys Ile Tyr Ile Tyr Thr Arg Ser Thr Ser Trp His Ser Lys Leu
420 425 430

Gln Leu Gly Ile Ile Asp Ile Thr Asp Tyr Ser Asp Ile Arg Ile Lys
435 440 445

Trp Thr Trp His Asn Val Leu Ser Arg Pro Gly Asn Asn Glu Cys Pro
450 455 460

Trp Gly His Ser Cys Pro Asp Gly Cys Ile Thr Gly Val Tyr Thr Asp
465 470 475 480

Ala Tyr Pro Leu Asn Pro Thr Gly Ser Ile Val Ser Ser Val Ile Leu
485 490 495

Asp Ser Gln Lys Ser Arg Val Asn Pro Val Ile Thr Tyr Ser Thr Ala
500 505 510

Thr Glu Arg Val Asn Glu Leu Ala Ile Arg Asn Arg Thr Leu Ser Ala
515 520 525

Gly Tyr Thr Thr Thr Ser Cys Ile Thr His Tyr Asn Lys Gly Tyr Cys
530 535 540

Phe His Ile Val Glu Ile Asn Gln Lys Ser Leu Asn Thr Leu Gln Pro
545 550 555 560

Met Leu Phe Lys Thr Glu Val Pro Lys Ser Cys Ser
565 570

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1886 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATGGAGTTGC CAATCCTCAA AGCAAATGCA ATTACCACAA TCCTCGCTGC AGTCACATTT 60

TGCTTTGCTT CTAGTCAAAA CATCACTGAA GAATTTTATC AATCAACATG CAGTGCAGTT 120

AGCAAAGGCT ATCTTAGTGC TCTAAGAACT GGTTGGTATA CTAGTGTTAT AACTATAGAA 180

TTAAGTAATA TCAAGGAAAA TAAGTGTAAT GGAACAGATG CTAAGGTAAA ATTGATGAAA 240

CAAGAATTAG ATAAATATAA AAATGCTGTA ACAGAATTGC AGTTGCTCAT GCAAAGCACA 300

CCAGCAGCAA ACAATCGAGC CAGAAGAGAA CTACCAAGGT TTATGAATTA TACACTCAAC 360

AATACCAAAA AAACCAATGT AACATTAAGC AAGAAAAGGA AAAGAAGATT TCTTGGTTTT 420

TTGTTAGGTG TTGGATCTGC AATCGCCAGT GGCATTGCTG TATCTAAGGT CCTGCACTTA 480

GAAGGAGAAG TGAACAAGAT CAAAAGTGCT CTACTATCCA CAAACAAGGC CGTAGTCAGC 540

TTATCAAATG GAGTTAGTGT CTTAACCAGC AAAGTGTTAG ACCTCAAAAA CTATATAGAT 600

AAACAATTGT TACCTATTGT GAATAAGCAA AGCTGCAGAA TATCAAATAT AGAAACTGTG 660

ATAGAGTTCC AACAAAAGAA CAACAGACTA CTAGAGATTA CCAGGGAATT TAGTGTTAAT 720

GCAGGTGTAA CTACACCTGT AAGCACTTAC ATGTTAACTA ATAGTGAATT ATTGTCATTA 780

ATCAATGATA TGCCTATAAC AAATGATCAG AAAAAGTTAA TGTCCAACAA TGTTCAAATA 840

GTTAGACAGC AAAGTTACTC TATCATGTCC ATAATAAAAG AGGAAGTCTT AGCATATGTA 900

GTACAATTAC CACTATATGG TGTGATAGAT ACACCTTGTT GGAAATTACA CACATCCCCT 960

CTATGTACAA CCAACACAAA AGAAGGGTCA AACATCTGTT TAACAAGAAC TGACAGAGGA 1020

TGGTACTGTG ACAATGCAGG ATCAGTATCT TTCTTCCCAC AAGCTGAAAC ATGTAAAGTT 1080

CAATCGAATC GAGTATTTTG TGACACAATG AACAGTTTAA CATTACCAAG TGAAGTAAAT 1140

CTCTGCAATG TTGACATATT CAATCCCAAA TATGATTGTA AAATTATGAC TTCAAAAACA 1200

GATGTAAGCA GCTCCGTTAT CACATCTCTA GGAGCCATTG TGTCATGCTA TGGCAAAACT 1260

AAATGTACAG CATCCAATAA AAATCGTGGA ATCATAAAGA CATTTTCTAA CGGGTGTGAT 1320

TATGTATCAA ATAAAGGGGT GGACACTGTG TCTGTAGGTA ACACATTATA TTATGTAAAT 1380

AAGCAAGAAG GCAAAAGTCT CTATGTAAAA GGTGAACCAA TAATAAATTT CTATGACCCA 1440

TTAGTATTCC CCTCTGATGA ATTTGATGCA TCAATATCTC AAGTCAATGA GAAGATTAAC 1500

CAGAGTTTAG CATTTATTCG TAAATCCGAT GAATTATTAC ATAATGTAAA TGCTGGTAAA 1560

TCAACCACAA ATATCATGAT AACTACTATA ATTATAGTGA TTATAGTAAT ATTGTTATCA 1620

TTAATTGCTG TTGGACTGCT CCTATACTGT AAGGCCAGAA GCACACCAGT CACACTAAGC 1680

AAGGATCAAC TGAGTGGTAT AAATAATATT GCATTTAGTA ACTGAATAAA AATAGCACCT 1740

AATCATGTTC TTACAATGGT TTACTATCTG CTCATAGACA ACCCATCTAT CATTGGATTT 1800

TCTTAAAATC TGAACTTCAT CGAAACTCTT ATCTATAAAC CATCTCACTT ACACTATTTA 1860

AGTAGATTCC TAGTTTATAG TTATAT 1886

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 594 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

```
Met Glu Leu Pro Ile Leu Lys Ala Asn Ala Ile Thr Thr Ile Leu Ala
1               5                   10                  15

Ala Val Thr Phe Cys Phe Ala Ser Ser Gln Asn Ile Thr Glu Glu Phe
            20                  25                  30

Tyr Gln Ser Thr Cys Ser Ala Val Ser Lys Gly Tyr Leu Ser Ala Leu
        35                  40                  45

Arg Thr Gly Trp Tyr Thr Ser Val Ile Thr Ile Glu Leu Ser Asn Ile
    50                  55                  60

Lys Glu Asn Lys Cys Asn Gly Thr Asp Ala Lys Val Lys Leu Met Lys
65                  70                  75                  80

Gln Glu Leu Asp Lys Tyr Lys Asn Ala Val Thr Glu Leu Gln Leu Leu
                85                  90                  95

Met Gln Ser Thr Pro Ala Ala Asn Asn Arg Ala Arg Arg Glu Leu Pro
            100                 105                 110

Arg Phe Met Asn Tyr Thr Leu Asn Asn Thr Lys Lys Thr Asn Val Thr
        115                 120                 125

Leu Ser Lys Lys Arg Lys Arg Arg Phe Leu Gly Phe Leu Leu Gly Val
    130                 135                 140

Gly Ser Ala Ile Ala Ser Gly Ile Ala Val Ser Lys Val Leu His Leu
145                 150                 155                 160

Glu Gly Glu Val Asn Lys Ile Lys Ser Ala Leu Leu Ser Thr Asn Lys
                165                 170                 175

Ala Val Val Ser Leu Ser Asn Gly Val Ser Val Leu Thr Ser Lys Val
            180                 185                 190

Leu Asp Leu Lys Asn Tyr Ile Asp Lys Gln Leu Leu Pro Ile Val Asn
        195                 200                 205
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Lys Gln Ser Cys Arg Ile Ser Asn Ile Glu Thr Val Ile Glu Phe Gln
210 215 220

His Lys Asn Asn Arg Leu Leu Glu Ile Thr Arg Glu Phe Ser Val Asn
225 230 235 240

Ala Gly Val Thr Thr Pro Val Ser Thr Tyr Met Leu Thr Asn Ser Glu
245 250 255

Leu Leu Ser Leu Ile Asn Asp Met Pro Ile Thr Asn Asp Gln Lys Lys
260 265 270

Leu Met Ser Asn Asn Val Gln Ile Val Arg Gln Gln Ser Tyr Ser Ile
275 280 285

Met Ser Ile Ile Lys Glu Glu Val Leu Ala Tyr Val Val Gln Leu Pro
290 295 300

Leu Tyr Gly Val Ile Asp Thr Pro Cys Trp Lys Leu His Thr Ser Pro
305 310 315 320

Leu Cys Thr Thr Asn Thr Lys Glu Gly Ser Asn Ile Cys Leu Thr Arg
325 330 335

Thr Asp Arg Gly Trp Tyr Cys Asp Asn Ala Gly Ser Val Ser Phe Phe
340 345 350

Pro Gln Ala Glu Thr Cys Lys Val Gln Ser Asn Arg Val Phe Cys Asp
355 360 365

Thr Met Asn Ser Leu Thr Leu Pro Ser Glu Val Asn Leu Cys Asn Val
370 375 380

Asp Ile Phe Asn Pro Lys Tyr Asp Cys Lys Ile Met Thr Ser Lys Thr
385 390 395 400

Asp Val Ser Ser Ser Val Ile Thr Ser Leu Gly Ala Ile Val Ser Cys
405 410 415

Tyr Gly Lys Thr Lys Cys Thr Ala Ser Asn Lys Asn Arg Gly Ile Ile
420 425 430

Lys Thr Phe Ser Asn Gly Cys Asp Tyr Val Ser Asn Lys Gly Val Asp
435 440 445

Thr Val Ser Val Gly Asn Thr Leu Tyr Tyr Val Asn Lys Gln Glu Gly
450 455 460

Lys Ser Leu Tyr Val Lys Gly Glu Pro Ile Ile Asn Phe Tyr Asp Pro
465 470 475 480

Leu Val Phe Pro Ser Asp Glu Phe Asp Ala Ser Ile Ser Gln Val Asn
485 490 495

Glu Lys Ile Asn Leu Val Phe Pro Ser Asp Glu Phe Asp Ala Ser Ile
500 505 510

Ser Gln Val Asn Glu Lys Ile Asn Gln Ser Leu Ala Phe Ile Arg Lys
515 520 525

Ser Asp Glu Leu Leu His Asn Val Asn Ala Gly Lys Ser Thr Thr Asn
530 535 540

Ile Met Ile Thr Thr Ile Ile Ile Glu Ile Ile Val Ile Leu Leu Ser
545 550 555 560

Leu Ile Ala Val Gly Leu Leu Leu Tyr Cys Lys Ala Arg Ser Thr Pro
565 570 575

Val Thr Leu Ser Lys Asp Gln Leu Ser Gly Ile Asn Asn Ile Ala Phe
580 585 590

Ser Asn

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 920 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

TGCAAACATG TCCAAAAACA AGGACCAACG CACCGCTAAG ACACTAGAAA AGACCTGGGA 60

CACTCTCAAT CATTTATTAT TCATATCATC GGGCTTATAT AAGTTAAATC TTAAATCTGT 120

AGCACAAATC ACATTATCCA TTCTGGCAAT GATAATCTCA ACTTCACTTA TAATTACAGC 180

CATCATATTC ATAGCCTCGG CAAACCACAA AGTCACACTA ACAACTGCAA TCATACAAGA 240

TGCAACAAGC CAGATCAAGA ACACAACCCC AACATACCTC ACTCAGGATC CTCAGCTTGG 300

AATCAGCTTC TCCAATCTGT CTGAAATTAC ATCACAAACC ACCACCATAC TAGCTTCAAC 360

AACACCAGGA GTCAAGTCAA ACCTGCAACC CACAACAGTC AAGACTAAAA ACACAACAAC 420

AACCCAAACA CAACCCAGCA AGCCCACTAC AAAACAACGC CAAAACAAAC CACCAAACAA 480

ACCCAATAAT GATTTTCACT TCGAAGTGTT TAACTTTGTA CCCTGCAGCA TATGCAGCAA 540

CAATCCAACC TGCTGGGCTA TCTGCAAAAG AATACCAAAC AAAAAACCAG GAAAGAAAAC 600

CACCACCAAG CCTACAAAAA AACCAACCTT CAAGACAACC AAAAAAGATC TCAAACCTCA 660

AACCACTAAA CCAAAGGAAG TACCCACCAC CAAGCCCACA GAAGAGCCAA CCATCAACAC 720

CACCAAAACA AACATCACAA CTACACTGCT CACCAACAAC ACCACAGGAA ATCCAAAACT 780

CACAAGTCAA ATGGAAACCT TCCACTCAAC CTCCTCCGAA GGCAATCTAA GCCCTTCTCA 840

AGTCTCCACA ACATCCGAGC ACCCATCACA ACCCTCATCT CCACCCAACA CAACACGCCA 900

GTAGTTATTA AAAAAAAAAA 920

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 298 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ser Lys Asn Lys Asp Gln Arg Thr Ala Lys Thr Leu Glu Lys Thr
1 5 10 15

Trp Asp Thr Leu Asn His Leu Leu Phe Ile Ser Ser Gly Leu Tyr Lys
20 25 30

Leu Asn Leu Lys Ser Val Ala Gln Ile Thr Leu Ser Ile Leu Ala Met
35 40 45

Ile Ile Ser Thr Ser Leu Ile Ile Thr Ala Ile Ile Phe Ile Ala Ser
50 55 60

Ala Asn His Lys Val Thr Leu Thr Thr Ala Ile Ile Gln Asp Ala Thr
65 70 75 80

Ser Gln Ile Lys Asn Thr Thr Pro Thr Tyr Leu Thr Gln Asp Pro Gln
85 90 95

Leu Gly Ile Ser Phe Ser Asn Leu Ser Glu Ile Thr Ser Gln Thr Thr
100 105 110

Thr Ile Leu Ala Ser Thr Thr Pro Gly Val Lys Ser Asn Leu Gln Pro
115 120 125

Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Thr Gln Thr Gln Pro Ser
130 135 140

Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro Asn
145 150 155 160

Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile Cys
165 170 175

Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn Lys
180 185 190

Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr Phe
195 200 205

Lys Thr Thr Lys Lys Asp Leu Lys Pro Gln Thr Thr Lys Pro Lys Glu
210 215 220

Val Pro Thr Thr Lys Pro Thr Glu Glu Pro Thr Ile Asn Thr Thr Lys
225 230 235 240

Thr Asn Ile Thr Thr Thr Leu Leu Thr Asn Asn Thr Thr Gly Asn Pro
245 250 255

Lys Leu Thr Ser Gln Met Glu Thr Phe His Ser Thr Ser Ser Glu Gly
260 265 270

Asn Leu Ser Pro Ser Gln Val Ser Thr Thr Ser Glu His Pro Ser Gln
275 280 285

Pro Ser Ser Pro Pro Asn Thr Thr Arg Gln
290 295

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

ATCAATCAAA GGTCCTGTGA TAATAG 26

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CATGACTTGA TAATGAG 17

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 86 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AATTCATGGA GTTGCTAATC CTCAAAGCAA ATGCAATTAC CACAATCCTC ACTGCAGTCA 60

CATTTTGTTT TGCTTCTGGT TCTAAG 86

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

ACTGGCATCA ATCTAGCACT ACATGAG 27

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 136 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

AATTCATGCC AACTTTAATA CTGCTAATTA TTACAACAAT GATTATGGCA TCTTCCTGCC 60

AAATAGATAT CACAAAACTA CAGCATGTAG GTGTATTGGT CAACAGTCCC AAAGGGATGA 120

AGATATCACA AAACTT 136

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 94 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

ATCATGGAGA TAATTAAAAT GATAACCATC TCGCAAATAA ATAAGTATTT TACTGTTTTC 60

GTAACAGTTT TGTAATAAAA AAACCTATAA ATAG 94

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 141 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

ATCATGGAGA TAATTAAAAT GATAACCATC TCGCAAATAA ATAAGTATTT TACTGTTTTC 60

GTAACAGTTT TGTAATAAAA AAACCTATAA ATATTCCGGA ATTCAGATCT GCAGCGGCCG 120

CTCCATCTAG AAGGTACCCG G 141

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 31 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CATGACTAAT TCCATCAAAA GTGAAAAGGC T 31

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CAAGAAAAAG GAATAAAA 18

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 39 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

ATTTCTGTGA TATTTGTGCG GTATAATGAT GCTATACCT 39

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CAGGAGAAGG GTATCAAG 18

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

AGGAGAAGGG TATCAAG 17

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 94 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

ATCATGGAGA TAATTAAAAT GATAACCATC TCGCAAATAA ATAAGTATTT TACTGTTTTC 60

GTAACAGTTT TGTAATAAAA AAACCTATAA ATAG 94

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Gln Glu Lys Gly Ile Lys
1 5

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Gln Glu Lys Gly Ile Lys
1 5

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

ATCAATCTAG CACTACACAG 20

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1617 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

ATGCCAACTT TAATACTGCT AATTATTACA ACAATGATTA TGGCATCTTC CTGCCAAATA 60

GATATCACAA AACTACAGCA TGTAGGTGTA TTGGTCAACA GTCCCAAAGG GATGAAGATA 120

TCACAAAACT TCGAAACAAG ATATCTAATT TTGAGCCTCA TACCAAAAAT AGAAGACTCT 180

AACTCTTGTG GTGACCAACA GATCAAACAA TACAAGAGGT TATTGGATAG ACTGATCATC 240

CCTCTATATG ATGGATTAAG ATTACAGAAA GATGTGATAG TAACCAATCA AGAATCCAAT 300

GAAAACACTG ATCCCAGAAC AAGACGATCC TTTGGAGGGG TAATTGGAAC CATTGCTCTG 360

GGAGTAGCAA CCTCAGCACA AATTACAGCG GCAGTTGCTC TGGTTGAAGC CAAGCAGGCA 420

AAATCAGACA TCGAAAAACT CAAAGAAGCA ATCAGGGACA CAAACAAAGC AGTGCAGTCA 480

GTTCAGAGCT CTATAGGAAA TTTAATAGTA GCAATTAAAT CAGTCCAAGA TTATGTCAAC 540

AACGAAATGG TGCCATCGAT TGCTAGACTA GGTTGTGAAG CAGCAGGACT TCAATTAGGA 600

ATTGCATTAA CACAGCATTA CTCAGAATTA ACAAACATAT TTGGTGATAA CATAGGATCG 660

TTACAAGAAA AAGGAATAAA ATTACAAGGT ATAGCATCAT TATACCGCAC AAATATCACA 720

GAAATATTCA CAACATCAAC AGTTGATAAA TATGATATCT ATGATCTATT ATTTACAGAA 780

TCAATAAAGG TGAGAGTTAT AGATGTTGAT TTGAATGATT ACTCAATCAC CCTCCAAGTC 840

AGACTCCCTT TATTAACTAG GCTGCTGAAC ACTCAGATCT ACAAAGTAGA TTCCATATCA 900

TATAATATCC AAAACAGAGA ATGGTATATC CCTCTTCCCA GCCATATCAT GACGAAAGGG 960

GCATTTCTAG GTGGAGCAGA TGTCAAGGAA TGTATAGAAG CATTCAGCAG TTATATATGC 1020

CCTTCTGATC CAGGATTTGT ACTAAACCAT GAAATGGAGA GCTGCTTATC AGGAAACATA 1080

TCCCAATGTC CAAGAACCAC GGTCACATCA GACATTGTTC CAAGATATGC ATTTGTCAAT 1140

GGAGGAGTGG TTGCAAACTG TATAACAACC ACCTGTACAT GCAACGGAAT CGACAATAGA 1200

ATCAATCAAC CACCTGATCA AGGAGTAAAA ATTATAACAC ATAAAGAATG TAATACAATA 1260

GGTATCAACG GAATGCTGTT CAATACAAAT AAAGAAGGAA CTCTTGCATT CTACACACCA 1320

AATGATATAA CACTAAATAA TTCTGTTGCA CTTGATCCAA TTGACATATC AATCGAGCTT 1380

AACAAAGCCA AATCAGATCT AGAAGAATCA AAAGAATGGA TAAGAAGGTC AAATCAAAAA 1440

CTAGATTCTA TTGGAAACTG GCATCAATCT AGCACTACAA TCATAATTAT TTTAATAATG 1500

ATCATTATAT TGTTTATAAT TAATGTAACG ATAATTACAA TTGCAATTAA GTATTACAGA 1560

ATTCAAAAGA GAAATCGAGT GGATCAAAAT GACAAGCCAT ATGTACTAAC AAACAAA 1617

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1715 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

ATGGAATACT GGAAGCATAC CAATCACGGA AAGGATGCTG GCAATGAGCT GGAGACGTCC 60

ATGGCTACTA ATGGCAACAA GCTCACCAAT AAGATAACAT ATATATTATG GACAATAATC 120

CTGGTGTTAT TATCAATAGT CTTCATCATA GTGCTAATTA ATTCCATCAA AAGTGAAAAG 180

GCTCATGAAT CATTGCTGCA AGACATAAAT AATGAGTTTA TGGAAATTAC AGAAAAGATC 240

CAAATGGCAT CGGATAATAC CAATGATCTA ATACAGTCAG GAGTGAATAC AAGGCTTCTT 300

ACAATTCAGA GTCATGTCCA GAATTATATA CCAATATCAC TGACACAACA GATGTCAGAT 360

CTTAGGAAAT TCATTAGTGA AATTACAATT AGAAATGATA ATCAAGAAGT GCTGCCACAA 420

AGAATAACAC ATGATGTGGG TATAAAACCT TTAAATCCAG ATGATTTTTG GAGATGCACG 480

TCTGGTCTTC CATCTTTAAT GAAAACTCCA AAAATAAGGT TAATGCCAGG GCCGGGATTA 540

TTAGCTATGC CAACGACTGT TGATGGCTGT ATCAGAACTC CGTCCTTAGT TATAAATGAT 600

CTGATTTATG CTTATACCTC AAATCTAATT ACTCGAGGTT GTCAGGATAT AGGAAAATCA 660

TATCAAGTCT TACAGATAGG GATAATAACT GTAAACTCAG ACTTGGTACC TGACTTAAAT 720

CCCAGGATCT CTCATACTTT TAACATAAAT GACAATAGGA AGTCATGTTC TCTAGCACTC 780

CTAAATACAG ATGTATATCA ACTGTGTTCA ACTCCCAAAG TTGATGAAAG ATCAGATTAT 840

GCATCATCAG GCATAGAAGA TATTGTACTT GATATTGTCA ATTATGATGG CTCAATCTCA 900

ACAACAAGAT TTAAGAATAA TAACATAAGC TTTGATCAAC CTTATGCTGC ACTATACCCA 960

TCTGTTGGAC CAGGGATATA CTACAAAGGC AAAATAATAT TTCTCGGGTA TGGAGGTCTT 1020

GAACATCCAA TAAATGAGAA TGTAATCTGC AACACAACTG GGTGTCCCGG GAAAACACAG 1080

AGAGACTGCA ATCAGGCATC TCATAGTCCA TGGTTTTCAG ATAGGAGGAT GGTCAACTCT 1140

ATCATTGTTG TTGACAAAGG CTTAAACTCA ATTCCAAAAT TGAAGGTATG GACGATATCT 1200

ATGAGACAGA ATTACTGGGG GTCAGAAGGA AGGTTACTTC TACTAGGTAA CAAGATCTAT 1260

ATATATACAA GATCCACAAG TTGGCATAGC AAGTTACAAT TAGGAATAAT TGATATTACT 1320

GATTACAGTG ATATAAGGAT AAAATGGACA TGGCATAATG TGCTATCAAG ACCAGGAAAC 1380

AATGAATGTC CATGGGGACA TTCATGTCCA GATGGATGTA TAACAGGAGT ATATACTGAT 1440

GCATATCCAC TCAATCCCAC AGGGAGCATT GTGTCATCTG TCATATTAGA TTCACAAAAA 1500

TCGAGAGTGA ACCCAGTCAT AACTTACTCA ACAGCAACCG AAAGAGTAAA CGAGCTGGCC 1560

ATCCGAAACA GAACACTCTC AGCTGGATAT ACAACAACAA GCTGCATCAC ACACTATAAC 1620

AAAGGATATT GTTTTCATAT AGTAGAAATA AATCAGAAAA GCTTAAACAC ACTTCAACCC 1680

ATGTTGTTCA AGACAGAGGT TCCAAAAAGC TGCAG 1715

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1722 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

ATGGAGTTGC CAATCCTCAA AGCAAATGCA ATTACCACAA TCCTCGCTGC AGTCACATTT 60

TGCTTTGCTT CTAGTCAAAA CATCACTGAA GAATTTTATC AATCAACATG CAGTGCAGTT 120

AGCAAAGGCT ATCTTAGTGC TCTAAGAACT GGTTGGTATA CTAGTGTTAT AACTATAGAA 180

TTAAGTAATA TCAAGGAAAA TAAGTGTAAT GGAACAGATG CTAAGGTAAA ATTGATGAAA 240

CAAGAATTAG ATAAATATAA AAATGCTGTA ACAGAATTGC AGTTGCTCAT GCAAAGCACA 300

CCAGCAGCAA ACAATCGAGC CAGAAGAGAA CTACCAAGGT TTATGAATTA TACACTCAAC 360

AATACCAAAA AAACCAATGT AACATTAAGC AAGAAAAGGA AAAGAAGATT TCTTGGTTTT 420

TTGTTAGGTG TTGGATCTGC AATCGCCAGT GGCATTGCTG TATCTAAGGT CCTGCACTTA 480

GAAGGAGAAG TGAACAAGAT CAAAAGTGCT CTACTATCCA CAAACAAGGC CGTAGTCAGC 540

TTATCAAATG GAGTTAGTGT CTTAACCAGC AAAGTGTTAG ACCTCAAAAA CTATATAGAT 600

AAACAATTGT TACCTATTGT GAATAAGCAA AGCTGCAGAA TATCAAATAT AGAAACTGTG 660

ATAGAGTTCC AACAAAAGAA CAACAGACTA CTAGAGATTA CCAGGGAATT TAGTGTTAAT 720

GCAGGTGTAA CTACACCTGT AAGCACTTAC ATGTTAACTA ATAGTGAATT ATTGTCATTA 780

ATCAATGATA TGCCTATAAC AAATGATCAG AAAAAGTTAA TGTCCAACAA TGTTCAAATA 840

GTTAGACAGC AAAGTTACTC TATCATGTCC ATAATAAAAG AGGAAGTCTT AGCATATGTA 900

GTACAATTAC CACTATATGG TGTGATAGAT ACACCTTGTT GGAAATTACA CACATCCCCT 960

CTATGTACAA CCAACACAAA AGAAGGGTCA AACATCTGTT TAACAAGAAC TGACAGAGGA 1020

TGGTACTGTG ACAATGCAGG ATCAGTATCT TTCTTCCCAC AAGCTGAAAC ATGTAAAGTT 1080

CAATCGAATC GAGTATTTTG TGACACAATG AACAGTTTAA CATTACCAAG TGAAGTAAAT 1140

CTCTGCAATG TTGACATATT CAATCCCAAA TATGATTGTA AAATTATGAC TTCAAAAACA 1200

GATGTAAGCA GCTCCGTTAT CACATCTCTA GGAGCCATTG TGTCATGCTA TGGCAAAACT 1260

AAATGTACAG CATCCAATAA AAATCGTGGA ATCATAAAGA CATTTTCTAA CGGGTGTGAT 1320

TATGTATCAA ATAAAGGGGT GGACACTGTG TCTGTAGGTA ACACATTATA TTATGTAAAT 1380

AAGCAAGAAG GCAAAAGTCT CTATGTAAAA GGTGAACCAA TAATAAATTT CTATGACCCA 1440

TTAGTATTCC CCTCTGATGA ATTTGATGCA TCAATATCTC AAGTCAATGA GAAGATTAAC 1500

CAGAGTTTAG CATTTATTCG TAAATCCGAT GAATTATTAC ATAATGTAAA TGCTGGTAAA 1560

TCAACCACAA ATATCATGAT AACTACTATA ATTATAGTGA TTATAGTAAT ATTGTTATCA 1620

TTAATTGCTG TTGGACTGCT CCTATACTGT AAGGCCAGAA GCACACCAGT CACACTAAGC 1680

AAGGATCAAC TGAGTGGTAT AAATAATATT GCATTTAGTA AC 1722

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 894 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

ATGTCCAAAA ACAAGGACCA ACGCACCGCT AAGACACTAG AAAAGACCTG GGACACTCTC 60

AATCATTTAT TATTCATATC ATCGGGCTTA TATAAGTTAA ATCTTAAATC TGTAGCACAA 120

ATCACATTAT CCATTCTGGC AATGATAATC TCAACTTCAC TTATAATTAC AGCCATCATA 180

TTCATAGCCT CGGCAAACCA CAAAGTCACA CTAACAACTG CAATCATACA AGATGCAACA 240

AGCCAGATCA AGAACACAAC CCCAACATAC CTCACTCAGG ATCCTCAGCT TGGAATCAGC 300

TTCTCCAATC TGTCTGAAAT TACATCACAA ACCACCACCA TACTAGCTTC AACAACACCA 360

GGAGTCAAGT CAAACCTGCA ACCCACAACA GTCAAGACTA AAAACACAAC AACAACCCAA 420

ACACAACCCA GCAAGCCCAC TACAAAACAA CGCCAAAACA AACCACCAAA CAAACCCAAT 480

AATGATTTTC ACTTCGAAGT GTTTAACTTT GTACCCTGCA GCATATGCAG CAACAATCCA 540

ACCTGCTGGG CTATCTGCAA AAGAATACCA AACAAAAAAC CAGGAAAGAA AACCACCACC 600

AAGCCTACAA AAAAACCAAC CTTCAAGACA ACCAAAAAAG ATCTCAAACC TCAAACCACT 660

AAACCAAAGG AAGTACCCAC CACCAAGCCC ACAGAAGAGC CAACCATCAA CACCACCAAA 720

ACAAACATCA CAACTACACT GCTCACCAAC AACACCACAG GAAATCCAAA ACTCACAAGT 780

CAAATGGAAA CCTTCCACTC AACCTCCTCC GAAGGCAATC TAAGCCCTTC TCAAGTCTCC 840

ACAACATCCG AGCACCCATC ACAACCCTCA TCTCCACCCA ACACAACACG CCAG 894

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 32 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

CGTAGTTAGT TTCCAGGACA CTATTATCCT AG 32

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

TGAACTATTA CTCCTAG 17

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 85 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

GTACCTCAAC GATTAGGAGT TTCGTTTACG TTAATGGTGT TAGGAGTGAC GTCAGTGTAA 60

AACAAAACGA AGACCAAGAT TCCAG 85

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

CGTAGTTAGA TCGTGATGTA CTCCTAG 27

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 136 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GTACGGTTGA AATTATGACG ATTAATAATG TTGTTACTAA TACCGTAGAA GGACGGTTTA 60

TCTATAGTGT TTTGATGTCG TACATCCACA TAACCAGTTG TCAGGGTTTC CCTACTTCTA 120

TAGTGTTTTG AAGCTT 136

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 98 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

TAGTACCTCT ATTAATTTTA CTATTGGTAG AGCGTTTATT TATTCATAAA ATGACAAAAG 60

CATTGTCAAA ACATTATTTT TTTGGATATT TATCTTAA 98

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 145 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

TAGTACCTCT ATTAATTTTA CTATTGGTAG AGCGTTTATT TATTCATAAA ATGACAAAAG 60

CATTGTCAAA ACATTATTTT TTTGGATATT TATAAGGCCT TAAGTCTAGA CGTCGCCGGC 120

GAGGTAGATC TTCCATGGGC CCTAG 145

(2) INFORMATION FOR SEQ ID NO:36:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 31 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

TGATTAAGGT AGTTTTCACT TTTCCGAGTA C 31

(2) INFORMATION FOR SEQ ID NO:37:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 39 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

TAAAGACACT ATAAACACGC CATATTACTA CGATATGGA 39

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

AGGACAAAAG